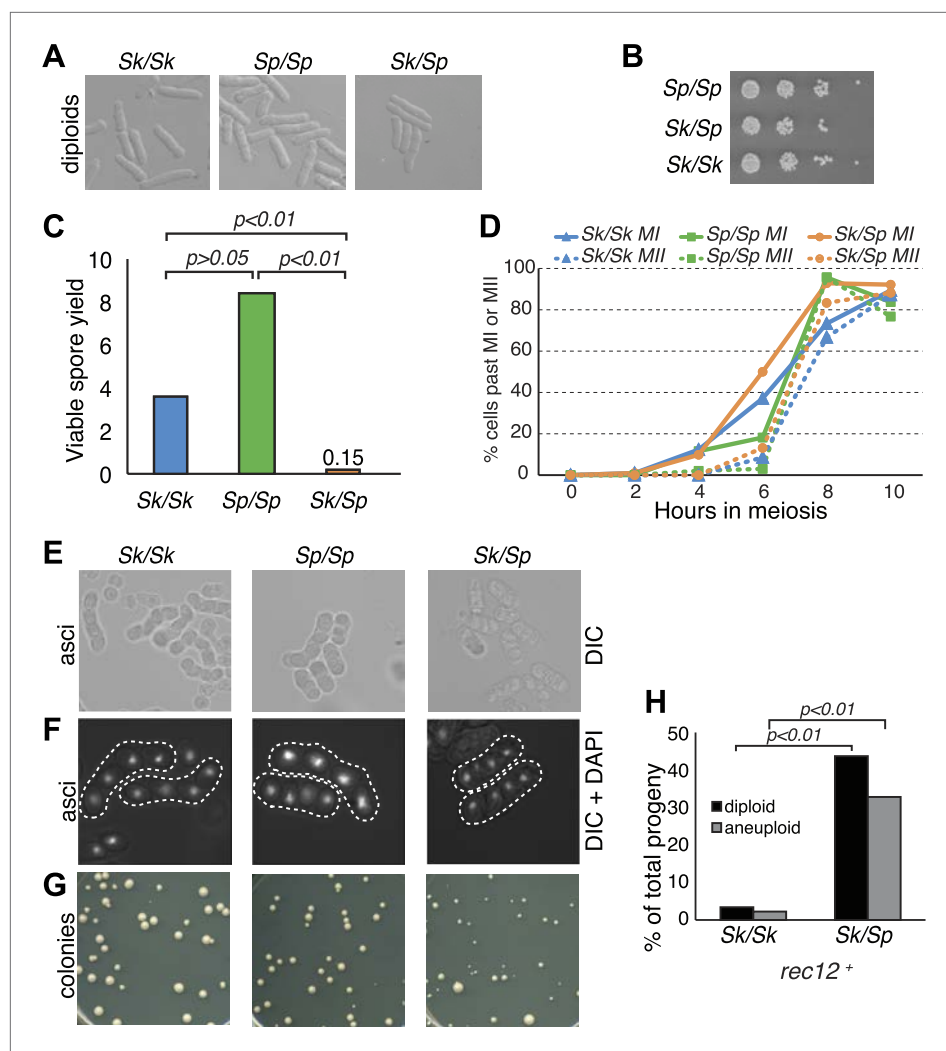


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## Figures and figure supplements

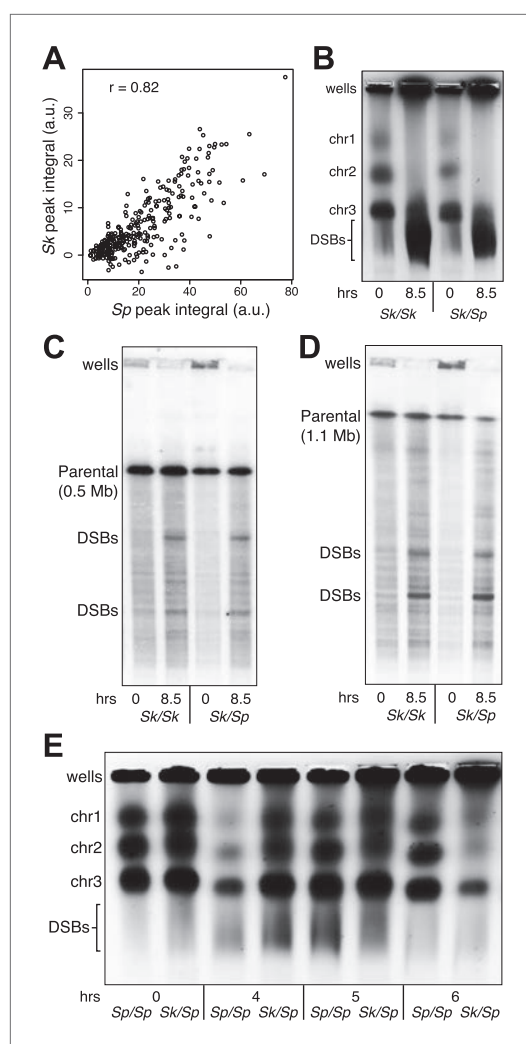
Genome rearrangements and pervasive meiotic drive cause hybrid infertility in fission yeast

**Sarah E Zanders, et al.**



**Figure 1.** *Sk/Sp* hybrids are healthy but exhibit low fertility. (A) *Sk/Sp* hybrid diploids are morphologically similar to pure species diploids. (B) *Sk/Sp* hybrid diploids show no gross growth defects relative to pure species controls. (C) Viable spore yield tests show that *Sk/Sp* fertility is low relative to pure species controls (averages of  $n \geq 5$  experiments, p-values obtained using t test). This assay does not directly measure viable spores per meiosis, so values can exceed 4. (D) *Sk/Sp* hybrids complete both meiotic divisions with timing similar to that of pure species controls (representative experiment of 3,  $n \geq 200$  cells for each data point). (E and F) The asci produced by *Sk/Sp* hybrids contain spores that are more irregular and transparent than pure species asci. (G) The viable spores produced by *Sk/Sp* hybrids often grow into small irregularly sized and shaped colonies. (H) The majority of the viable spores produced by *Sk/Sp* hybrids are aneuploid or diploid (p-values obtained using G-test,  $n > 200$  for each). These data are also shown in **Figure 5A**.

DOI: [10.7554/eLife.02630.003](https://doi.org/10.7554/eLife.02630.003)

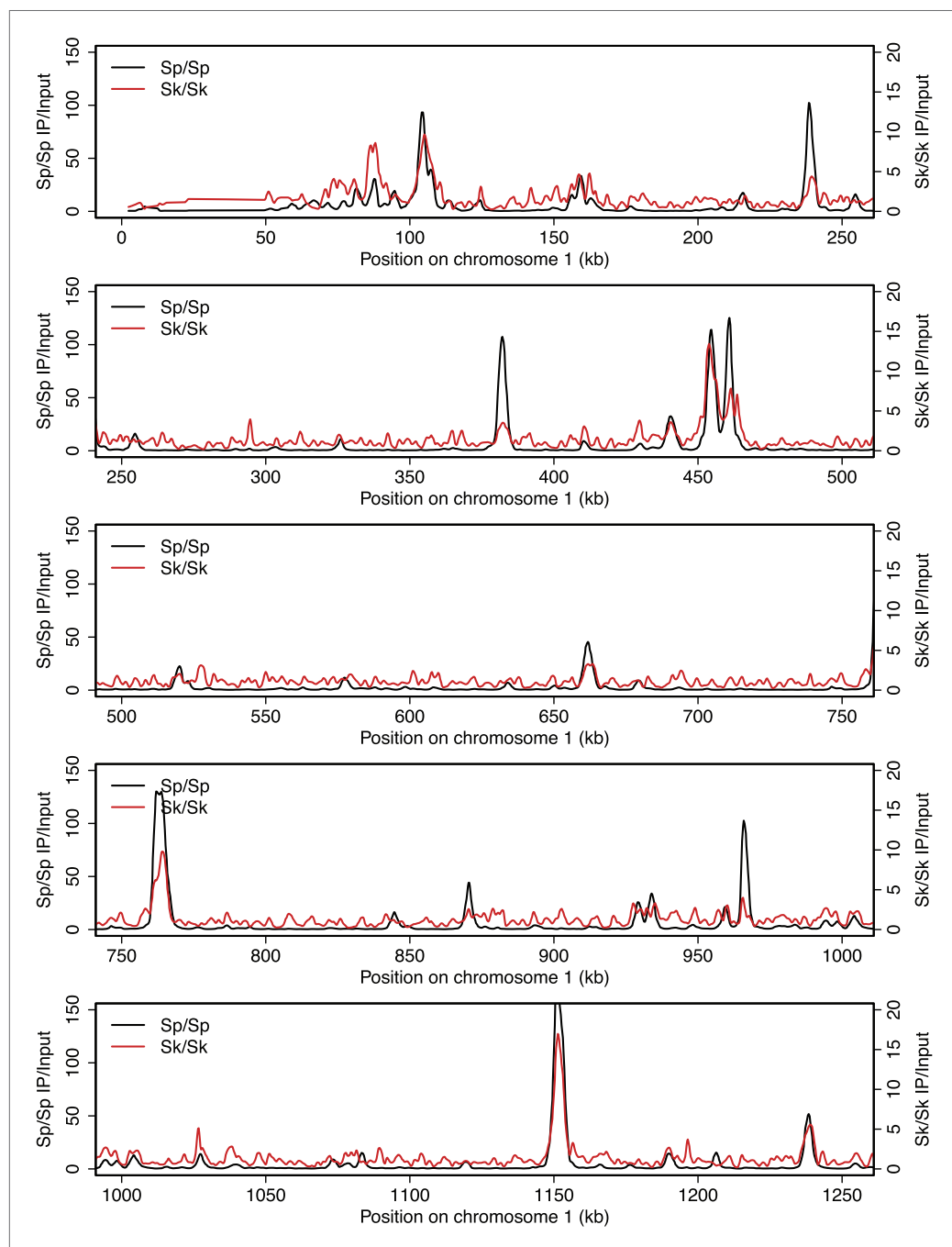


**Figure 2.** DSB hotspot divergence and repair in *Sk/Sp* hybrids. **(A)** We used ChIP–chip of Rec12-FLAG from *rad50S* *Sk* meiotic cultures to assay DSB hotspots and compared the profile to the published DSB hotspot maps of *Sp* (Fowler et al., 2013). We then compared the Rec12-enrichment in *Sk* at 286 defined *Sp* hotspots and found a strong correlation between Rec12 enrichments between the two species at these sites. **(B)** *Sk/Sp* cells are proficient at inducing DSBs. Ethidium bromide stained pulsed-field gel of diploids at 0 and 8.5 hr after inducing meiosis in liquid cultures. These diploids are *rad50S* mutants, so DSBs form normally but are not repaired. As DSBs are formed, the three full-sized chromosome bands disappear and the DNA runs as smaller broken fragments on the gel. **(C and D)** We find that DSBs are formed at similar locations and similar frequencies in *Sk/Sp* and *Sk/Sk*. Southern blots of pulsed-field gels to obtain a closer view of DSB formation in *rad50S* diploids probed to visualize two *NotI* restriction fragments known as *NotI* J [shown in (C)] and *NotI* D [shown in (D)]. Prior to DSB formation, most of the DNA runs as a single large band. After all break formation (8.5 hr) smaller cut fragments become apparent at the same Figure 2. Continued on next page

Figure 2. Continued

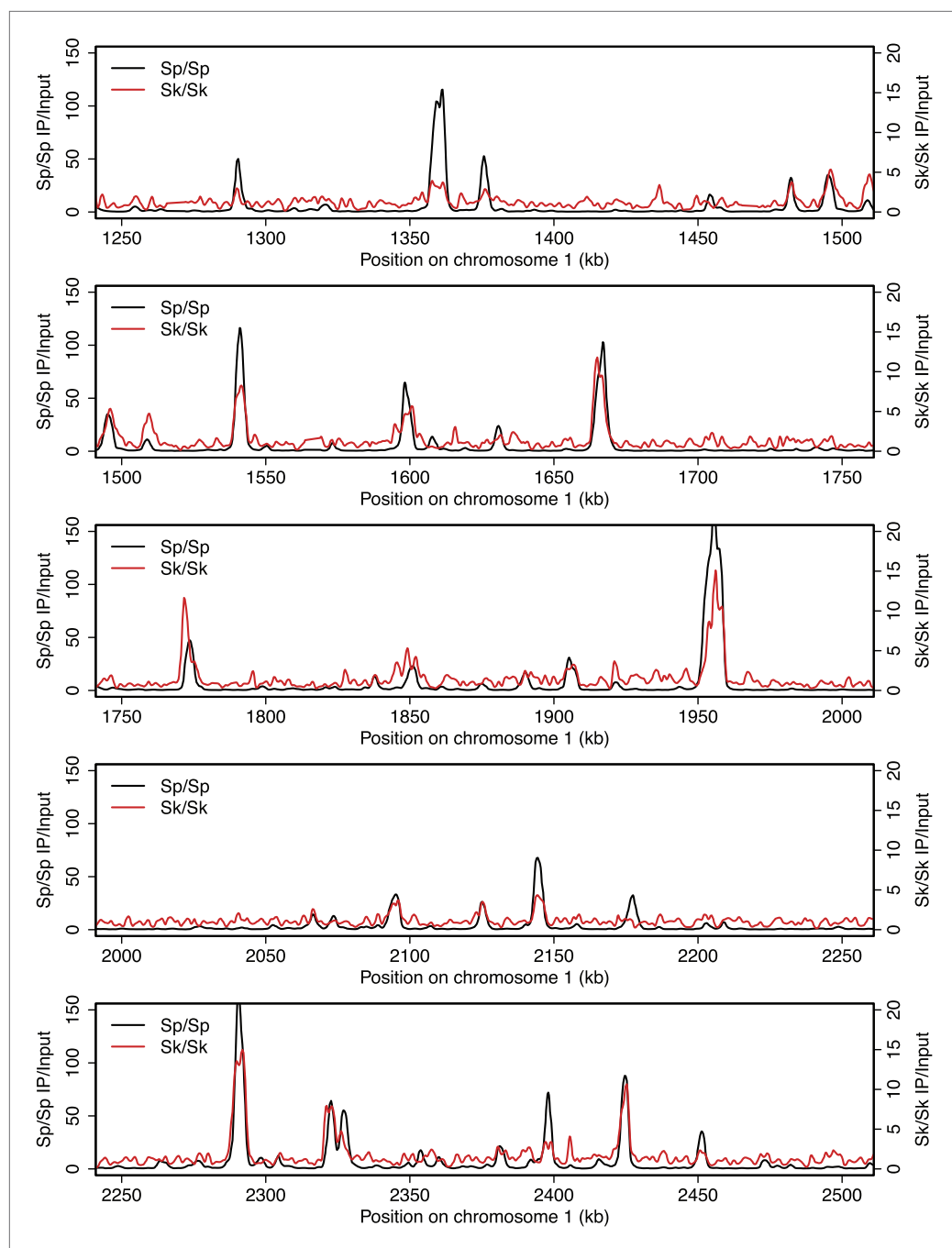
sites in *Sk/Sp* and *Sk/Sk*. (E) DSBs are efficiently repaired in *Sk/Sp*. Ethidium bromide stained pulsed-field gel of *rad50<sup>+</sup>* diploids at the given times after the induction of meiosis show that DSBs do not accumulate more in *Sk/Sp* than the *Sp/Sp* control during meiotic prophase. Together with those in (B) these data demonstrate that *Sk/Sp* cells form and efficiently repair DSBs.

DOI: [10.7554/eLife.02630.004](https://doi.org/10.7554/eLife.02630.004)



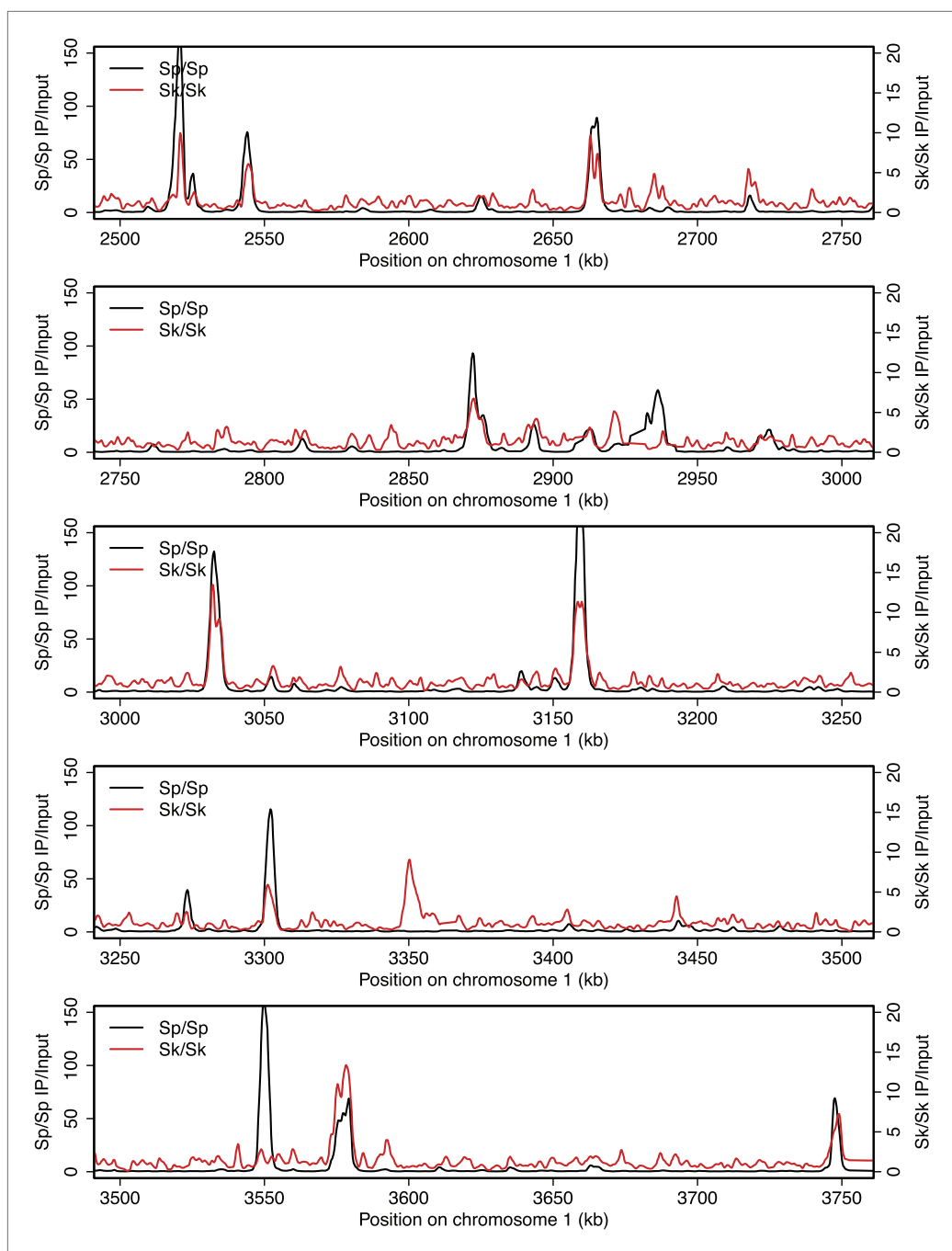
**Figure 2—figure supplement 1.** DSB hotspots in *Sk* and *Sp*.

DOI: [10.7554/eLife.02630.005](https://doi.org/10.7554/eLife.02630.005)



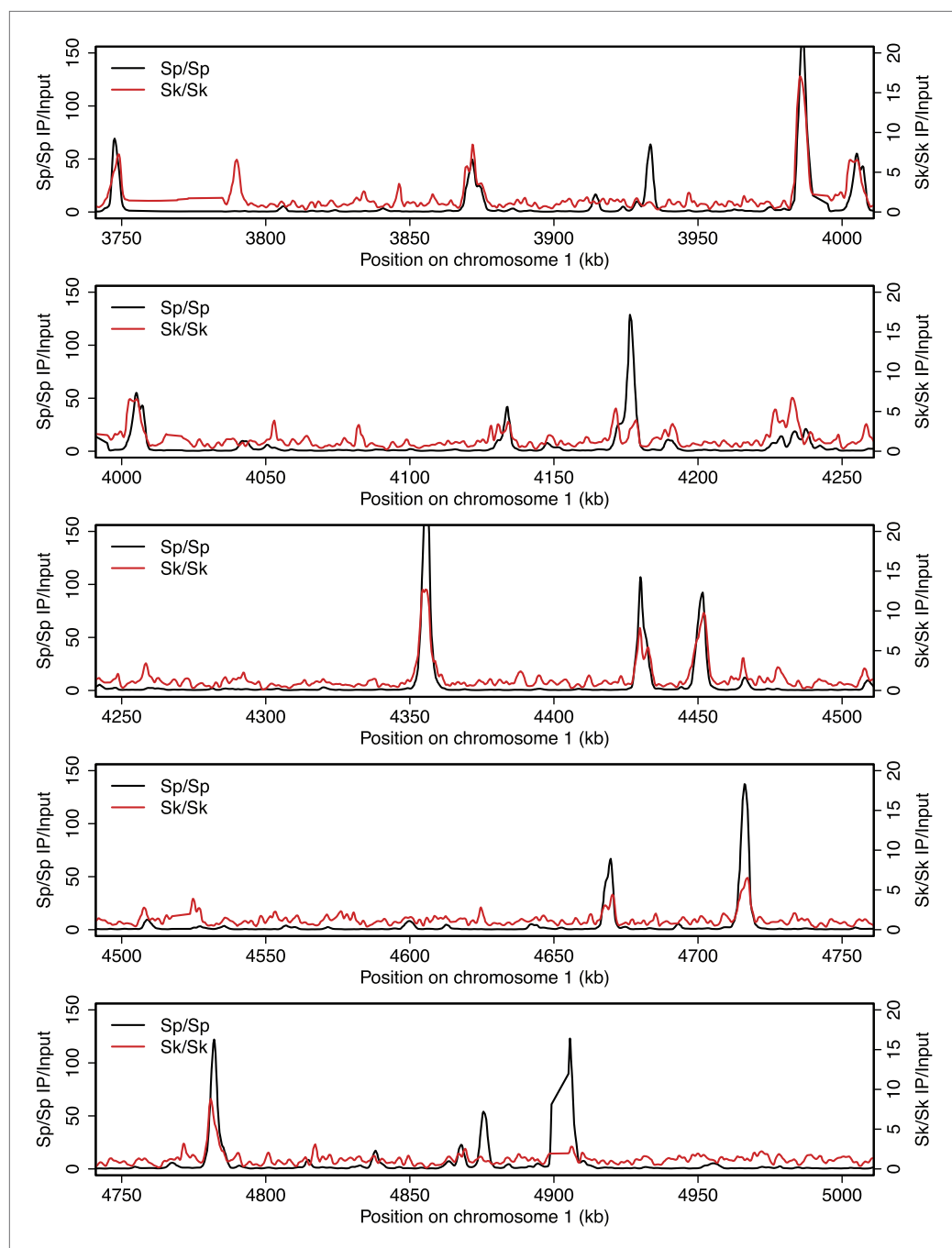
**Figure 2—figure supplement 2.** DSB hotspots in *Sk* and *Sp*.

DOI: [10.7554/eLife.02630.006](https://doi.org/10.7554/eLife.02630.006)



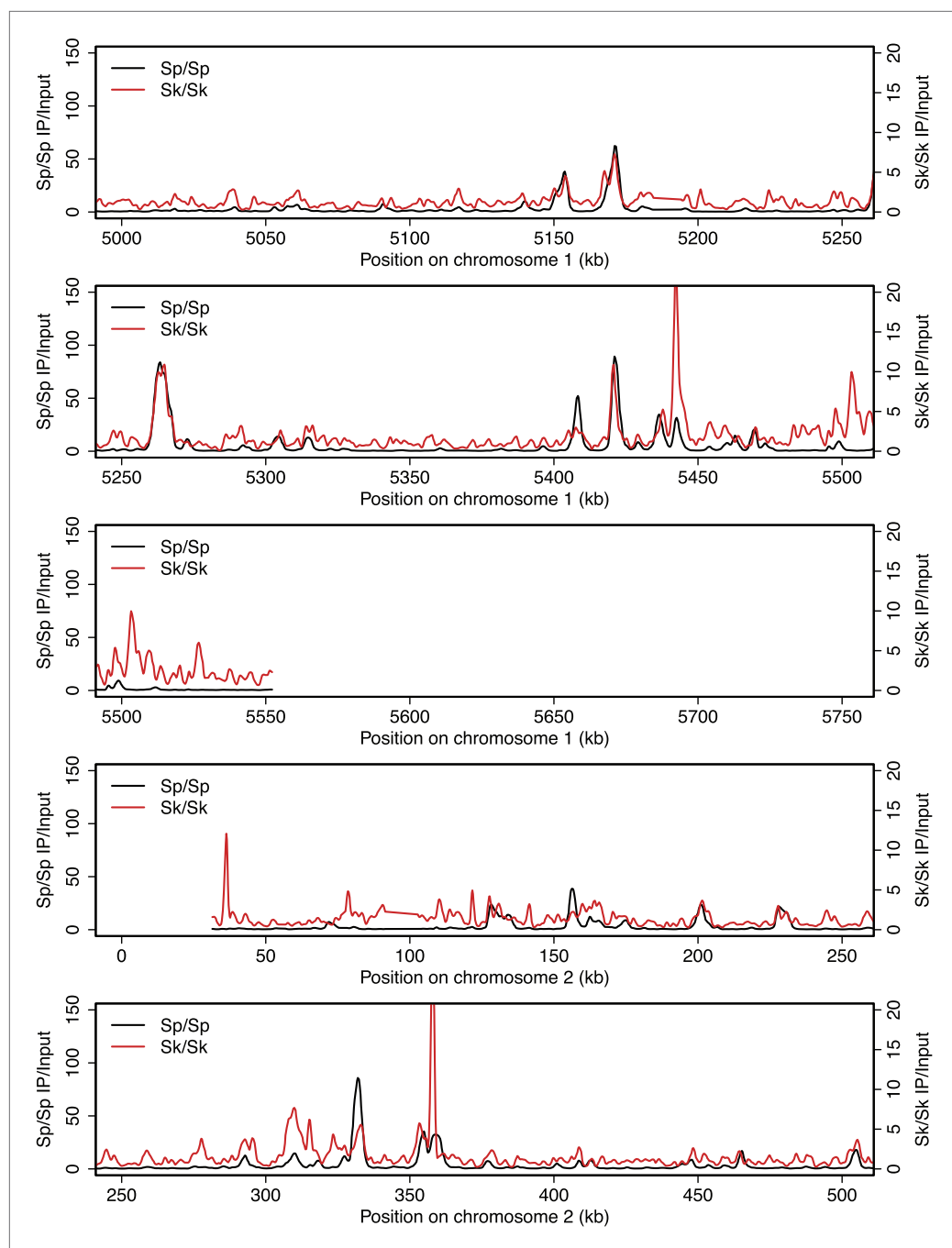
**Figure 2—figure supplement 3.** DSB hotspots in *Sk* and *Sp*.

DOI: [10.7554/eLife.02630.007](https://doi.org/10.7554/eLife.02630.007)



**Figure 2—figure supplement 4.** DSB hotspots in *Sk* and *Sp*.

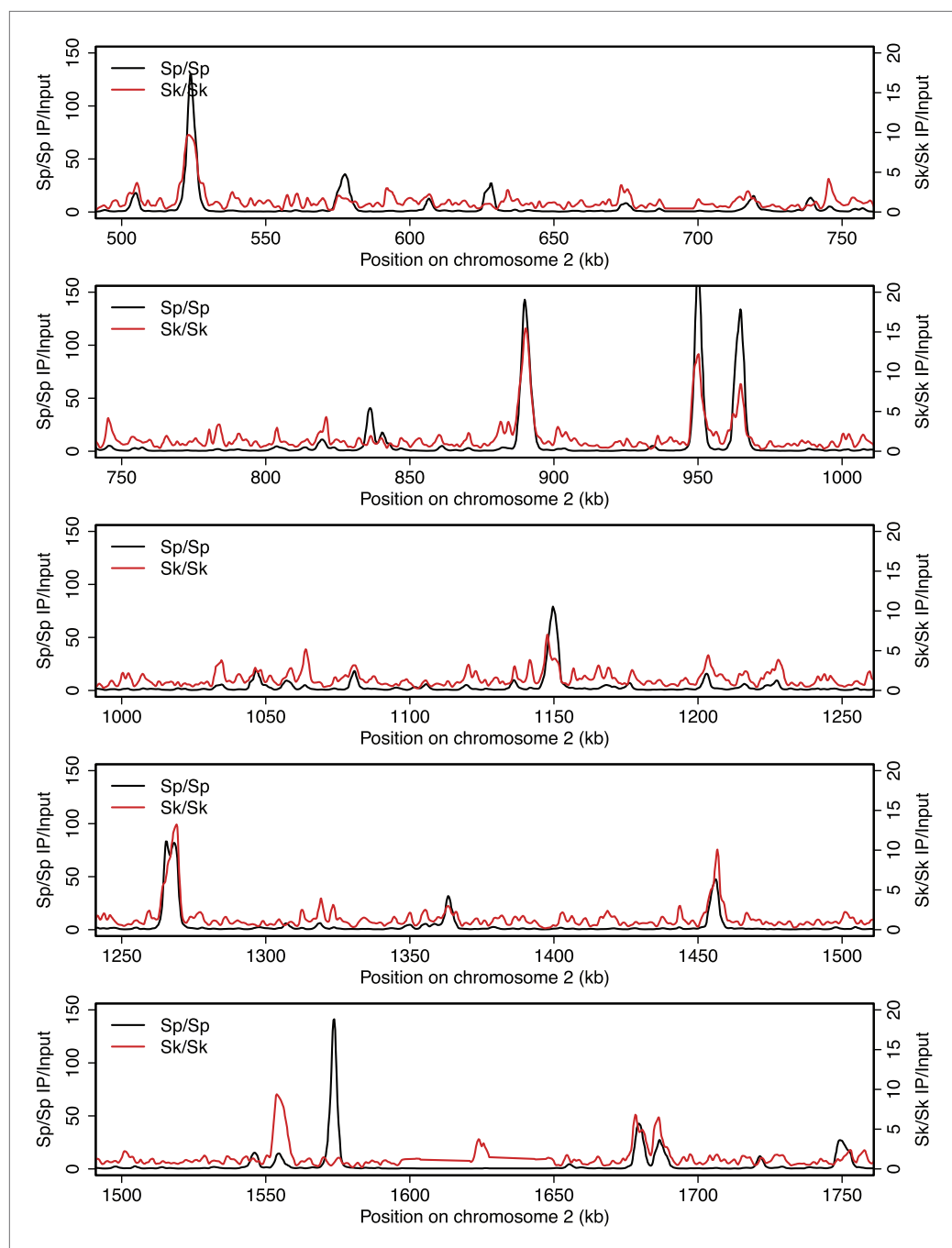
DOI: [10.7554/eLife.02630.008](https://doi.org/10.7554/eLife.02630.008)



**Figure 2—figure supplement 5.** DSB hotspots in *Sk* and *Sp*.

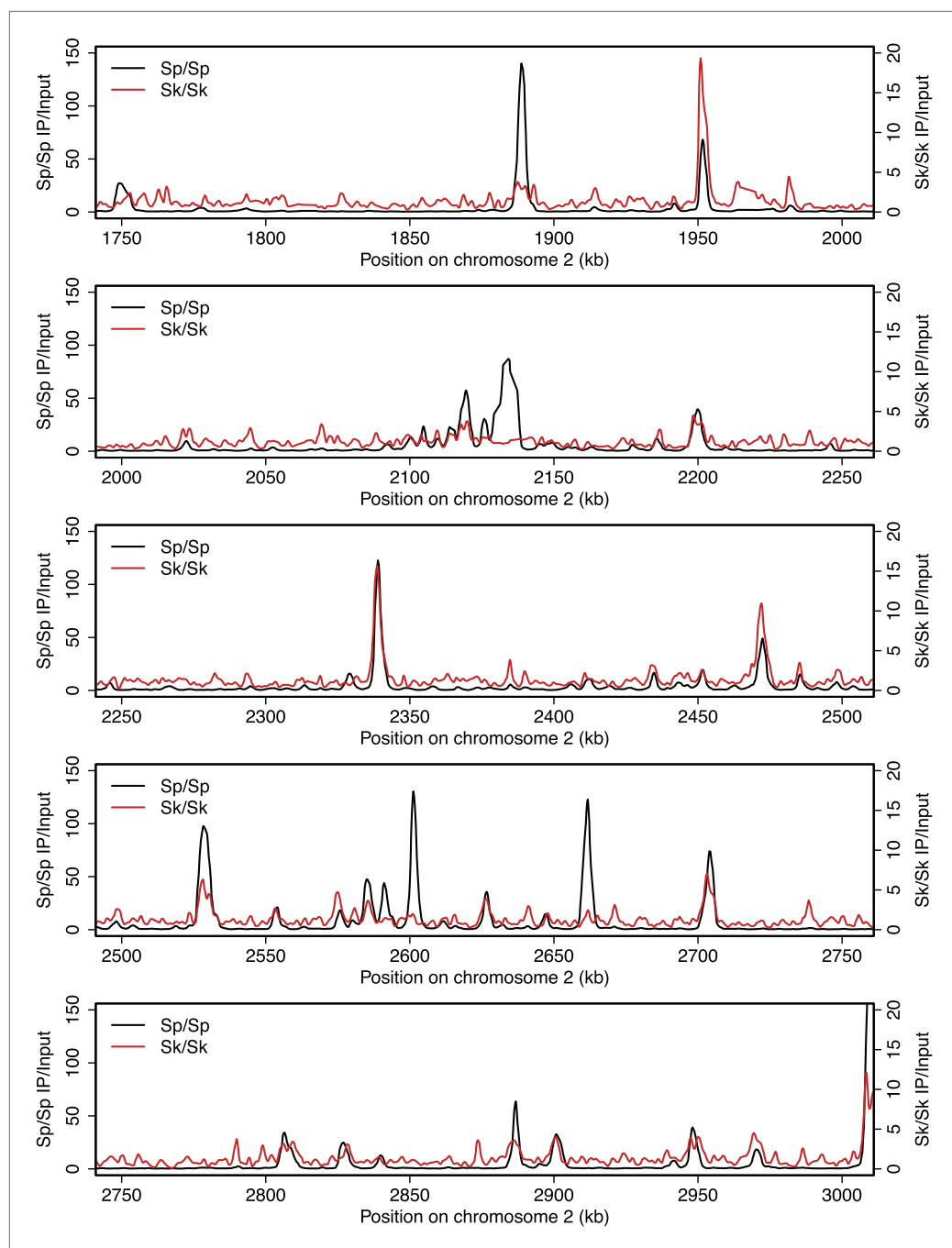
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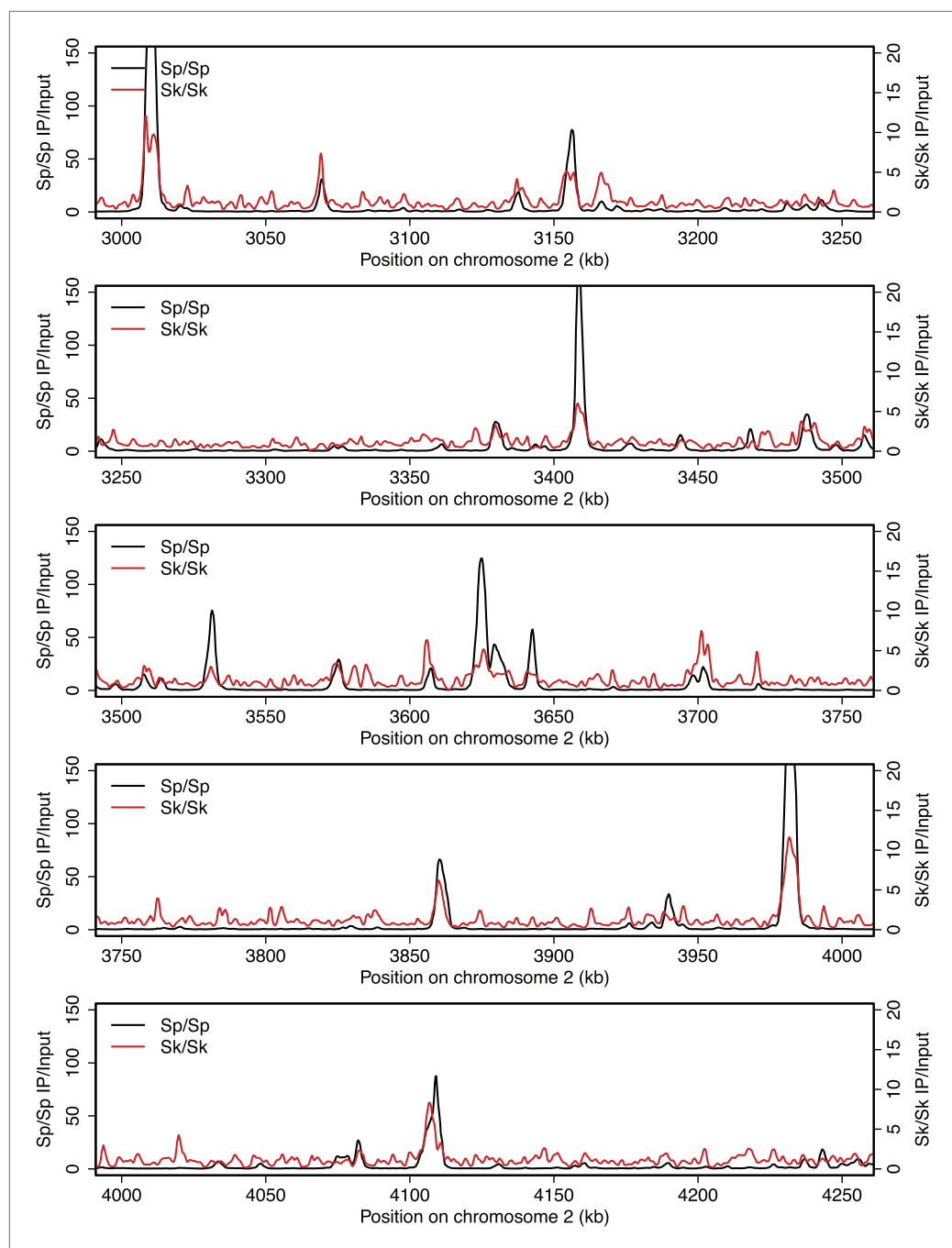
**Figure 2—figure supplement 6.** DSB hotspots in *Sk* and *Sp*.

DOI: [10.7554/eLife.02630.010](https://doi.org/10.7554/eLife.02630.010)



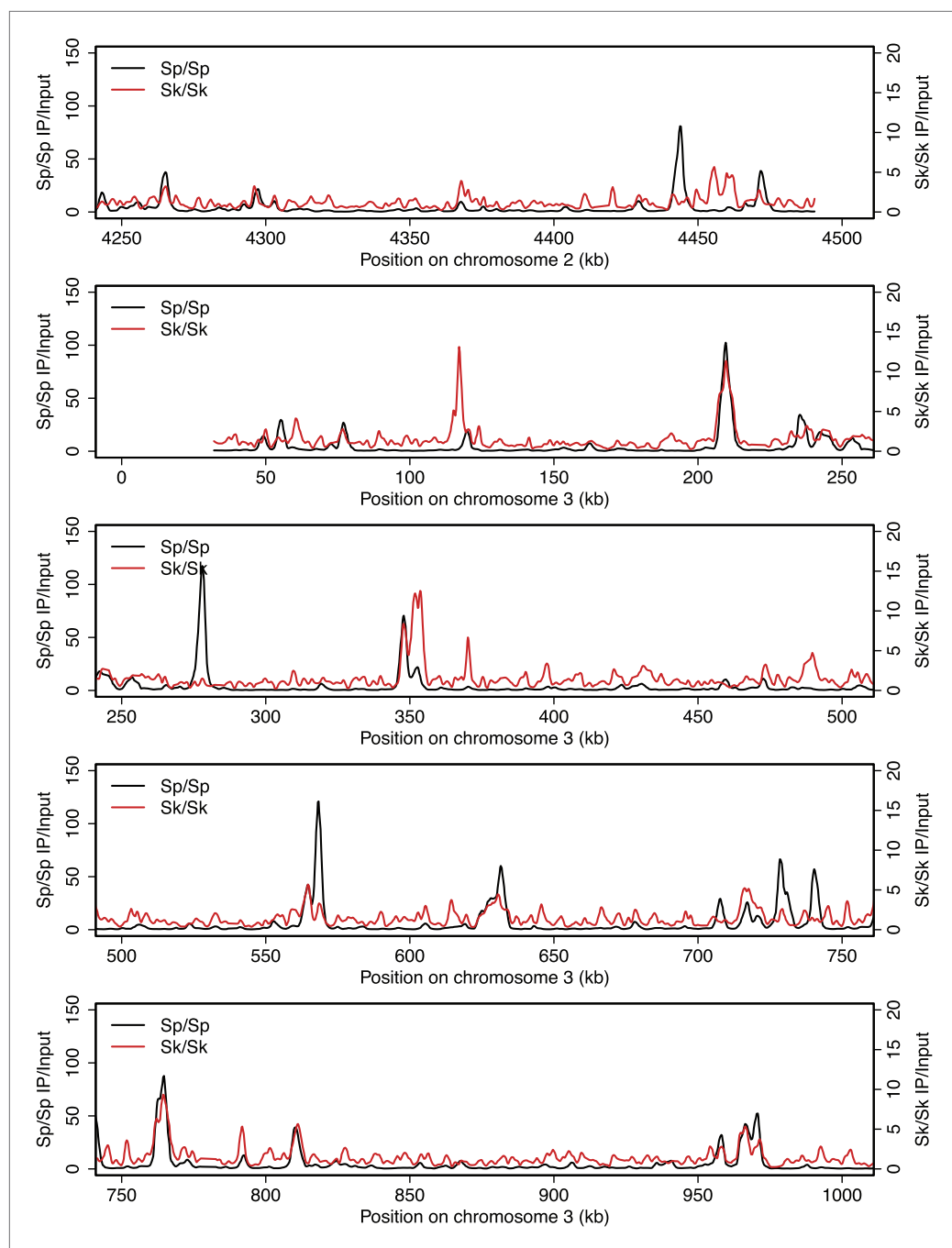
**Figure 2—figure supplement 7.** DSB hotspots in *Sk* and *Sp*.

DOI: [10.7554/eLife.02630.011](https://doi.org/10.7554/eLife.02630.011)



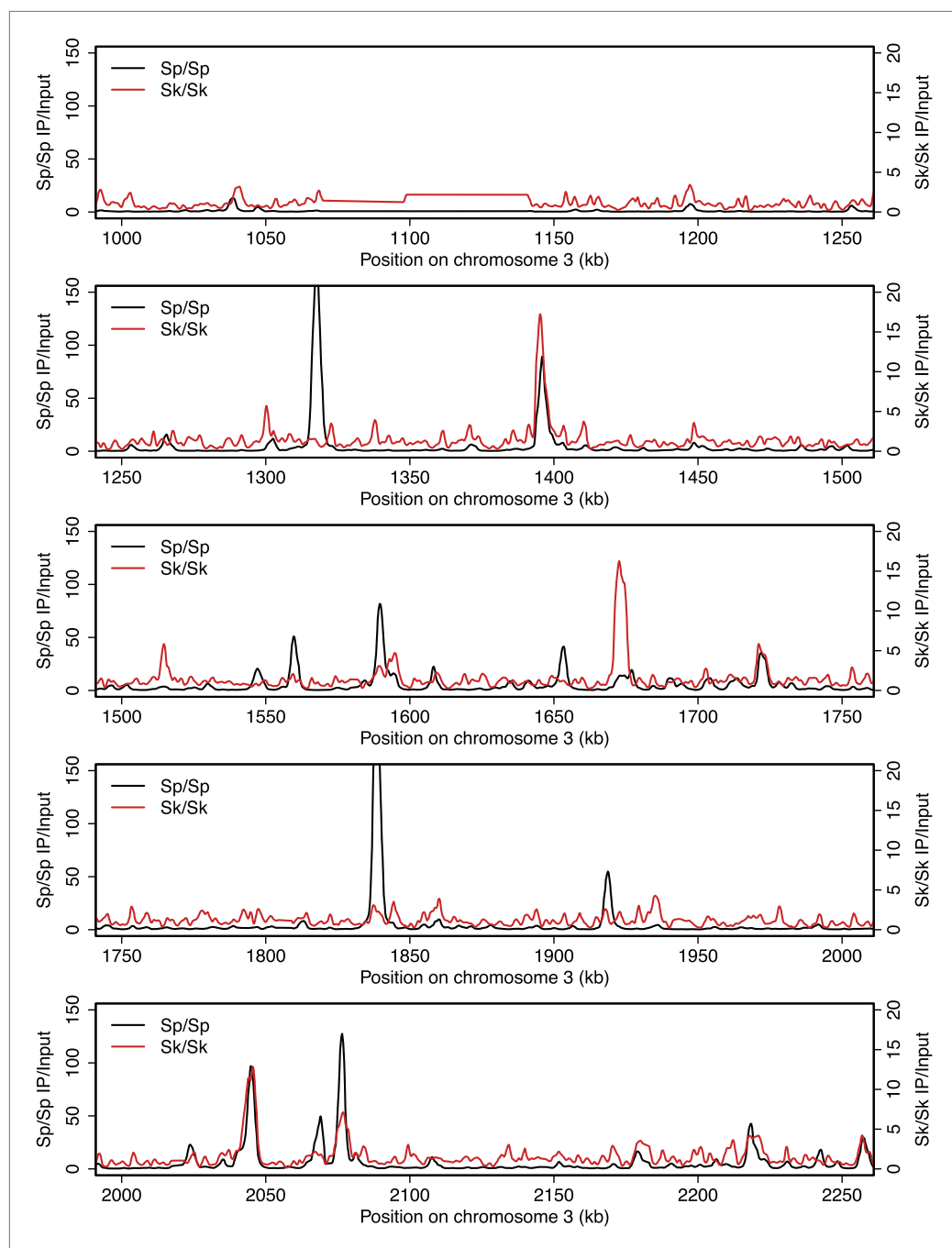
**Figure 2—figure supplement 8.** DSB hotspots in *Sk* and *Sp*.

DOI: [10.7554/eLife.02630.012](https://doi.org/10.7554/eLife.02630.012)



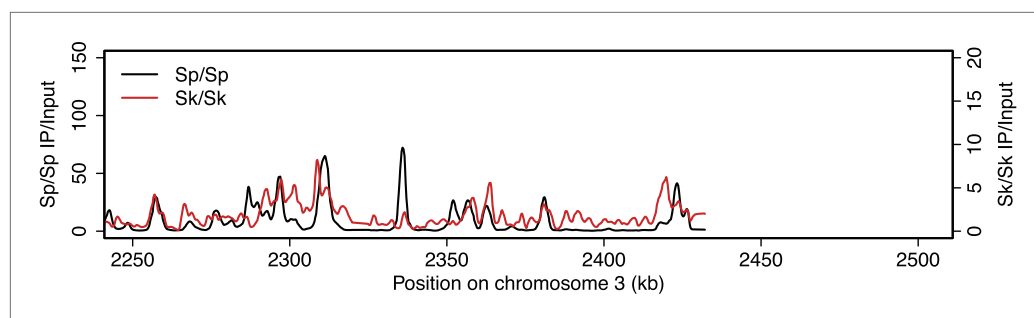
**Figure 2—figure supplement 9.** DSB hotspots in *Sk* and *Sp*.

DOI: [10.7554/eLife.02630.013](https://doi.org/10.7554/eLife.02630.013)



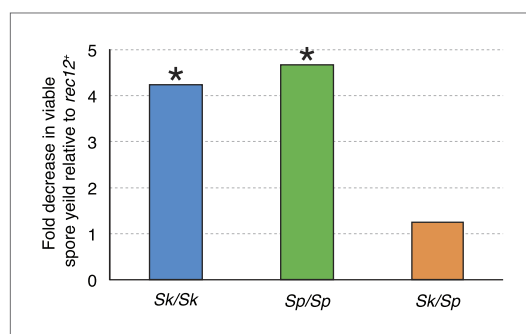
**Figure 2—figure supplement 10.** DSB hotspots in *Sk* and *Sp*.

DOI: [10.7554/eLife.02630.014](https://doi.org/10.7554/eLife.02630.014)



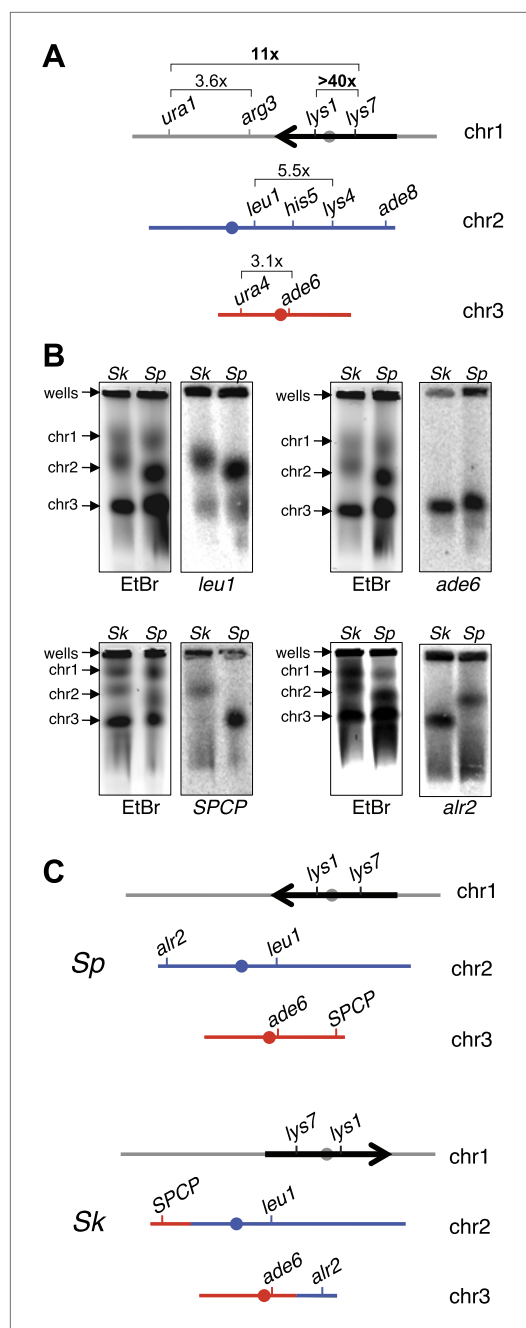
**Figure 2—figure supplement 11.** DSB hotspots in *Sk* and *Sp*.

DOI: [10.7554/eLife.02630.015](https://doi.org/10.7554/eLife.02630.015)



**Figure 3.** Recombination does not alter *Sk/Sp* hybrid fertility. The average *rec12*<sup>+</sup> viable spore yield of each diploid was divided by that of the corresponding *rec12Δ* mutant. For the pure species diploids, the viable spore yield was significantly lower in the absence of *Rec12* ( $n \geq 5$  experiments for each genotype; \*  $t$  test  $p < 0.05$ ). The viable spore yield of *Sk/Sp* hybrids, on the other hand, was not significantly different between *rec12*<sup>+</sup> and *rec12Δ* *Sk/Sp* hybrids ( $p = 0.42$ ). This indicates that recombination likely hurts fertility just as much as it promotes fertility in *Sk/Sp* hybrid meiosis. These data are shown in a different format in **Figure 5A**.

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**Figure 4.** Chromosome rearrangements limit the recovery of recombinant progeny in *Sk/Sp* hybrids. **(A)** A cartoon illustrating the fold decrease in recombinant frequencies in *Sk/Sp* spores compared to that in *Sp/Sp* spores. The detailed recombination data are in **Figure 4—figure supplements 2 and 3**. The *Sp* karyotype is depicted with a grey/black chromosome 1, blue chromosome 2, and red chromosome 3. The backwards black arrow indicates an inversion in *Sp* relative to the ancestral karyotype. **(B)** Pulsed-field gels separating *Sp* and *Sk* chromosomes and Southern blots of the gels probed with DNA from the indicated loci revealed a reciprocal translocation that includes several Figure 4. Continued on next page

Figure 4. Continued

essential genes including *alr2* and *SPCP1E11.08* (abbreviated *SPCP*). The EtBr-stained gels are on the left and the Southern blots are on the right in each pair. (C) A cartoon summary of the karyotype differences between *Sp* and *Sk*. The arrow indicates the location of the inversion in *Sp*. A few landmark loci are shown.

DOI: 10.7554/eLife.02630.017

Chromosome	Interval	Recombinants	Total	<i>Sk/Sk</i> distance (cM)	<i>Sp/Sp</i> distance (cM)	Fold Change
2	<i>lys4 – his4</i>	3	129	2.3	12	5.2
2	<i>his4 – ade8</i>	42	132	50.6	94	1.9
3	<i>ade6 – ura4</i>	72	157	unlinked	190	NA

Figure 4—figure supplement 1. Recombination frequencies in *Sk*.

DOI: 10.7554/eLife.02630.018

Chr	Interval	Recombinants	Total	<i>Sk/Sp</i> distance (cM)	<i>Sp/Sp</i> distance (cM)	Fold decrease
1	<i>ura1 – arg3</i>	95	257	67	240	3.6
1	<i>ura1 – lys7</i>	32	95	56	600	11
1	<i>ura1 – lys1</i>	53	257	54	480	8.9
1	<i>arg3 – lys1</i>	33	257	27	230	8.5
1	<i>arg3 – lys7</i>	52	212	34	360	11
1	<i>lys1 – lys7</i>	0	95	0	120	>40
2	<i>leu1 – his5</i>	12	201	6.1	46	7.5
2	<i>his5 – lys4</i>	57	201	42	200	4.8
2	<i>leu1 – lys4</i>	59	201	44	240	5.5
2	<i>lys4 – his4</i>	17	307	5.9	12	2
2	<i>his4 – ade8</i>	43	307	16	94	5.9
2	<i>lys4 – ade8</i>	48	307	19	100	5.3
3	<i>ade6 – ura4</i>	51	144	62	190	3.1
2 and 3	<i>leu1 – ade6</i>	20	101	25	NA	NA

Figure 4—figure supplement 2. Recombination frequencies in *Sk/Sp* hybrids are low relative to *Sp*.

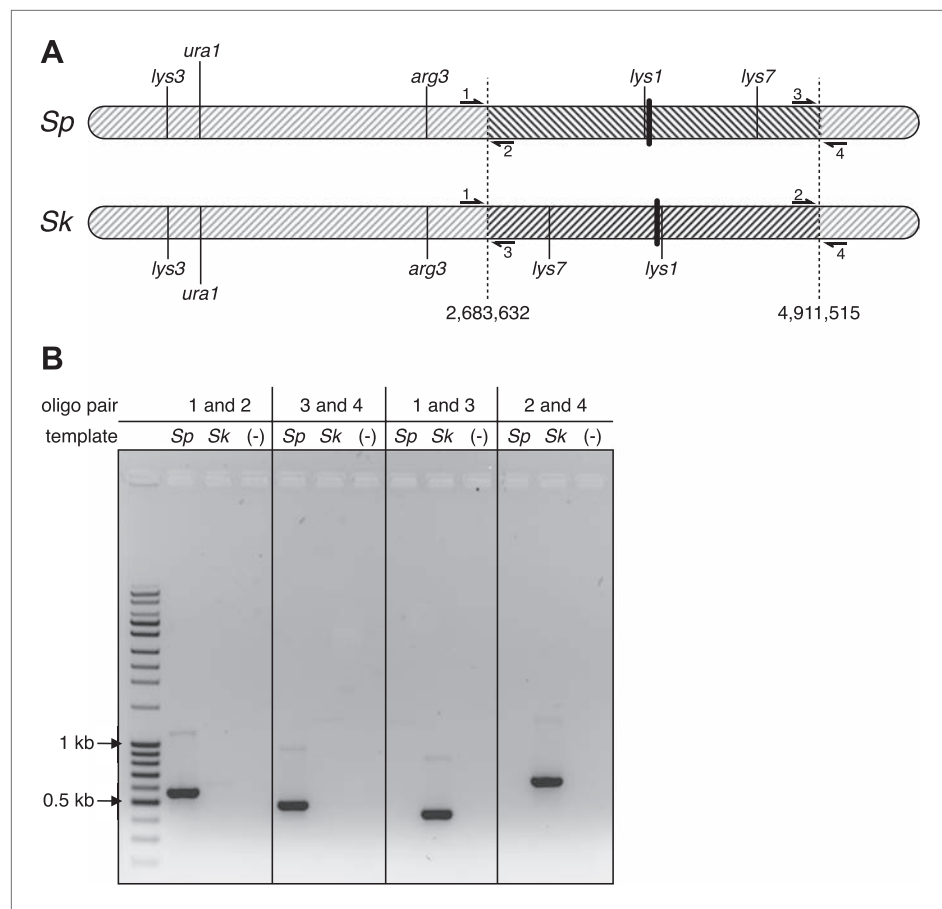
DOI: 10.7554/eLife.02630.019



strains crossed	Chr.	Parental <i>Sp</i>		Parental <i>Sk</i>		Recombinant 1		Recombinant 2		Hybrid distance (cM)	<i>Sp</i> Distance (cM)
		Genotype	#	Genotype	#	Genotype	#	Genotype	#		
SZY142x210	1	<i>arg3- lys7-</i>	51	<i>arg3+ lys7+</i>	109	<i>arg3- lys7+</i>	36	<i>arg3+ lys7-</i>	16	34	360
SZY142x180	1	<i>ura1- arg3-</i>	64	<i>ura1+ arg3+</i>	98	<i>ura1- arg3+</i>	52	<i>ura1+ arg3-</i>	43	67	240
SZY142x180*	1	<i>ura1- lys7-</i>	63	<i>ura1+ lys7+</i>	0	<i>ura1- lys7+</i>	0	<i>ura1+ lys7-</i>	32	56	600
SZY142x180*	1	<i>lys1+ lys7-</i>	95	<i>lys1- lys7+</i>	NA	<i>lys1+ lys7+</i>	0	<i>lys1- lys7-</i>	NA	0	120
SZY142x180	1	<i>ura1- lys1+</i>	63	<i>ura1+ lys1-</i>	109	<i>ura1- lys1-</i>	53	<i>ura1+ lys1+</i>	32	54	480
SZY142x180	1	<i>arg3- lys1+</i>	74	<i>arg3+ lys1-</i>	129	<i>arg3- lys1-</i>	33	<i>arg3+ lys1+</i>	21	27	230
SZY128x71	2	<i>leu1- his5-</i>	12	<i>leu1+ his5+</i>	177	<i>leu1- his5+</i>	1	<i>leu1+ his5-</i>	11	6.1	46
SZY128x71	2	<i>his5- lys4+</i>	15	<i>his5+ lys4-</i>	129	<i>his5- lys4-</i>	8	<i>his5+ lys4+</i>	49	42	200
SZY128x71	2	<i>leu1- lys4+</i>	9	<i>leu1+ lys4-</i>	133	<i>leu1- lys4-</i>	4	<i>leu1+ lys4+</i>	55	44	240
SZY127x94	2	<i>lys4+ his4+</i>	104	<i>lys4- his4-</i>	186	<i>lys4+ his4-</i>	8	<i>lys4- his4+</i>	9	5.9	12
SZY127x94	2	<i>his4+ ade8-</i>	91	<i>his4- ade8+</i>	173	<i>his4+ ade8+</i>	22	<i>his4- ade8-</i>	21	16	94
SZY127x94	2	<i>lys4+ ade8-</i>	88	<i>lys4- ade8+</i>	171	<i>lys4+ ade8+</i>	24	<i>lys4- ade8-</i>	24	19	110
SZY297x480	3	<i>ade6+ ura4+</i>	22	<i>ade6- ura4-</i>	71	<i>ade6+ ura4-</i>	4	<i>ade6- ura4+</i>	47	62	190

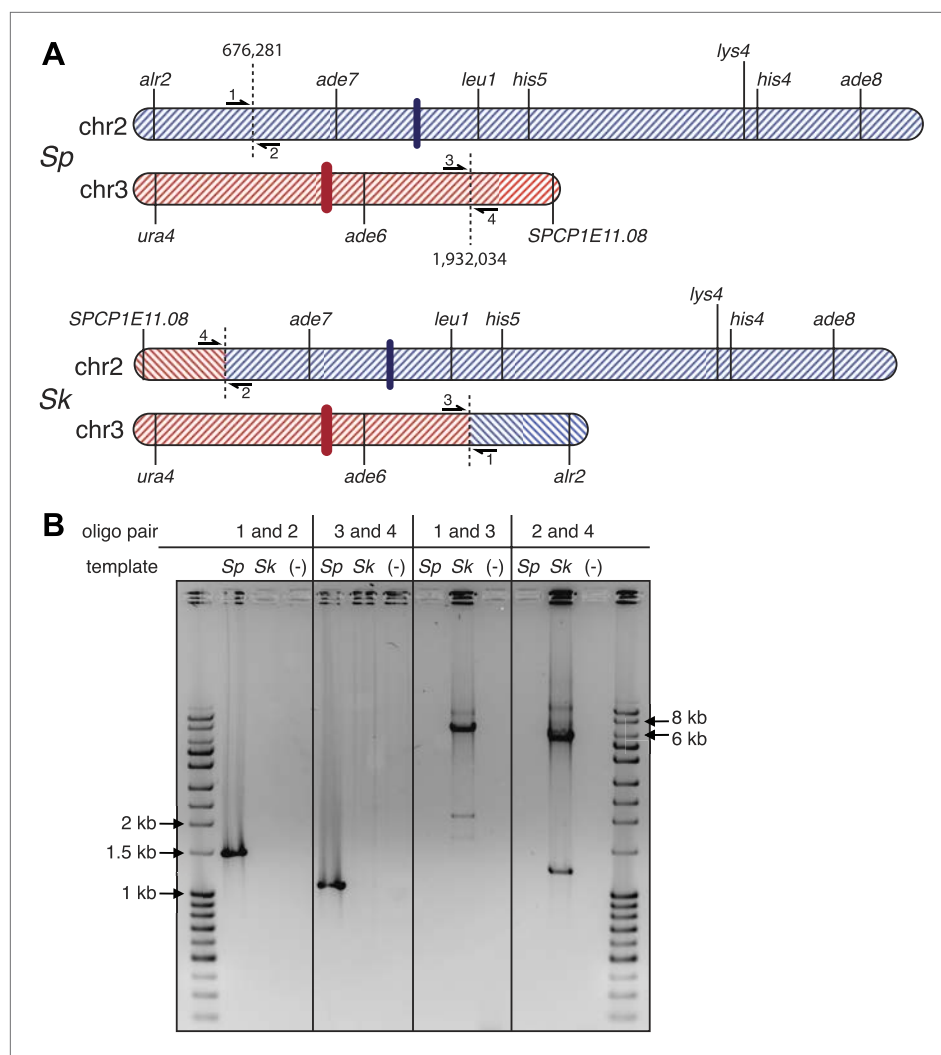
**Figure 4—figure supplement 3.** *Sk* alleles are underrepresented in the progeny of *Sk/Sp* hybrids.

DOI: [10.7554/eLife.02630.020](https://doi.org/10.7554/eLife.02630.020)



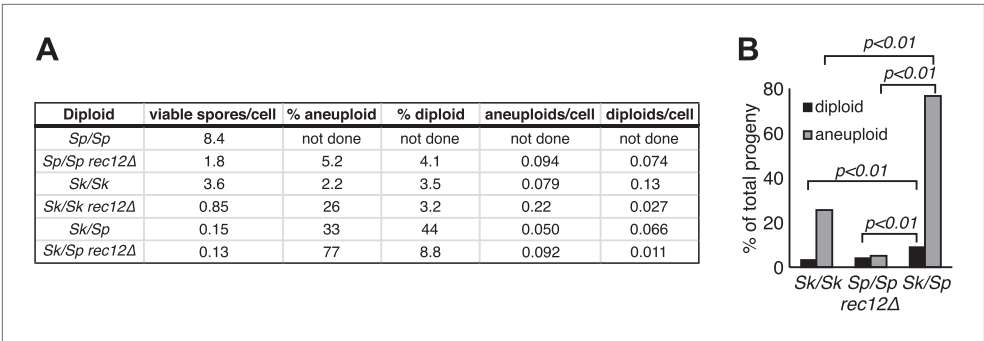
**Figure 4—figure supplement 4.** *Sp* has an inversion on chromosome 1 relative to *Sk*.

DOI: [10.7554/eLife.02630.021](https://doi.org/10.7554/eLife.02630.021)

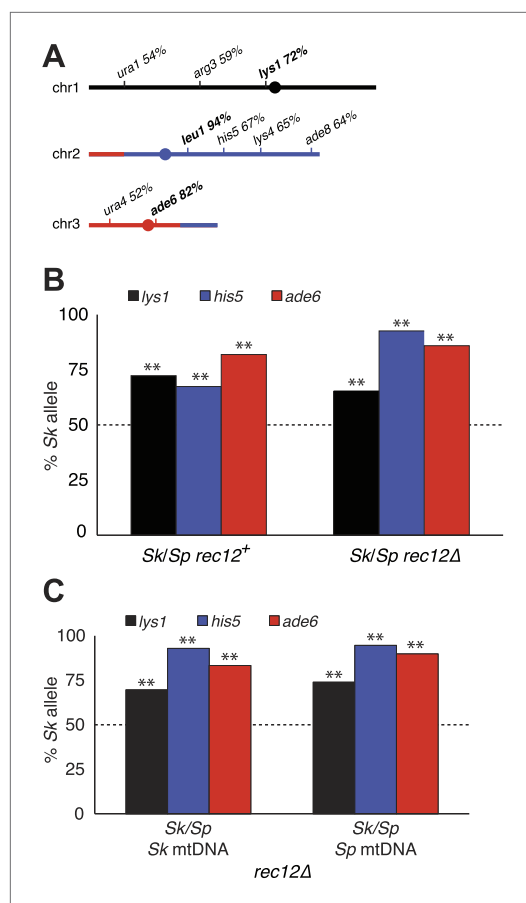


**Figure 4—figure supplement 5.** *Sk* has a reciprocal translocation between chromosomes 2 and 3.

DOI: [10.7554/eLife.02630.022](https://doi.org/10.7554/eLife.02630.022)



**Figure 5.** Increased aneuploidy amongst viable *Sk/Sp* gametes is recombination-independent. **(A)** We calculated both viable spore yield (viable spores/cell) as well as the fraction of viable spores that are aneuploid or diploid ('Materials and methods'). In the absence of *Rec12*, the relative frequencies of aneuploids and diploids are elevated in all cases. However, there is significantly more aneuploidy and diploidy of viable spores produced by *rec12Δ Sk/Sp* hybrids than by *rec12Δ* pure species controls. This shows the phenotype is not caused solely by recombination defects. In addition, *Sk/Sp* diploids do not generate more aneuploids or diploids relative to the number of cells induced to undergo meiosis compared to pure-species controls. Some of these data are presented in a different format in **Figures 1H** and **Figure 3**. **(B)** A bar graph illustrating the fraction of the viable spores produced by the indicated *rec12Δ* diploids that are aneuploid or diploid (G-test,  $n > 300$  for each). DOI: [10.7554/eLife.02630.023](https://doi.org/10.7554/eLife.02630.023)



**Figure 6.** Alleles on all three *Sk* chromosomes show drive (independent of mitochondrial DNA type). **(A)** *Sk* alleles were inherited by significantly more than 50% of the viable spores produced by *Sk/Sp* hybrids, except *ura1* and *ura4* (G-test  $p < 0.01$ ;  $n > 100$  for each). The markers nearest to the meiotic drive loci (i.e., those showing the greatest bias towards *Sk* inheritance) are shown in boldface. The color scheme is the same as that in **Figure 4**. The data underlying these numbers are shown in **Figure 4—figure supplement 3**, and **Figure 6—figure supplement 1**. **(B)** The *Sk* alleles of *lys1*, *his5* and *ade6* show significant drive both in the presence and absence of recombination (\*\* $p < 0.01$ ,  $n > 300$  for *lys1* and *his5*,  $n > 80$  for *ade6*). The amount of *his5* drive is greater in the absence to *Rec12* due to enhanced linkage with the driving locus. The data underlying this graph are shown in **Figure 6—figure supplement 1**. **(C)** Incompatibilities between the *Sk* mitochondrial DNA and *Sp* nuclear genes are not responsible for the drive phenotype because we observed the same drive in *rec12Δ* *Sk/Sp* hybrids with either *Sk* or *Sp*-derived mitochondrial DNA (\*\* $p < 0.01$ ,  $n > 200$  for *lys1* and *his5*,  $n > 50$  for *ade6*). The data underlying this graph are shown in **Figure 6—figure supplement 4**.

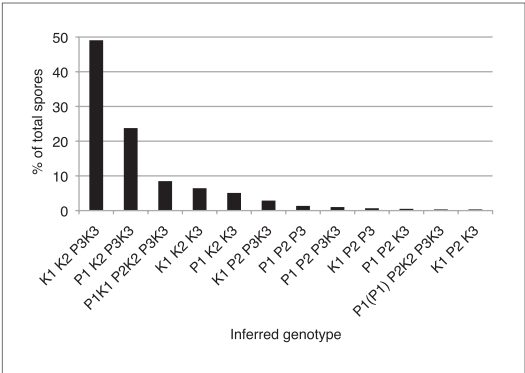
DOI: 10.7554/eLife.02630.024

locus	<i>Sp/Sp rec12*</i> (n=328)		<i>Sp/Sp rec12Δ</i> (n=368)		<i>Sk/Sk rec12*</i> (n=227)		<i>Sk/Sk rec12Δ</i> (n=342)		<i>Sk/Sp rec12*</i> (n=621)		<i>Sk/Sp rec12Δ</i> (n=589)	
	observed	%	observed	%	observed	%	observed	%	observed	%	observed	%
<i>his5*</i>	149	45.4	174	49.9	113	51.6	158	47.7	236	67.4	40	7.4
<i>his5</i>	179	54.6	175	51.1	106	48.4	173	52.3	114	32.6	497	92.6
<i>lys1*</i>	159	48.5	168	48.1	111	50.7	173	52.3	97	27.7	350	65.2
<i>lys1</i>	169	51.5	181	51.9	108	49.3	158	47.7	253	72.3	187	34.8
<i>ade6*</i>	178	54.3	153	46.4	94	43.9	129	53.1	26	18.1	12	14.1
<i>ade6</i>	150	45.7	177	53.6	120	56.1	114	46.9	118	81.9	73	85.9
diploid		ND	19	5.2	8	3.5	11	3.2	271	43.6	52	8.8
aneuploid		ND	19	5.2	5	2.2	88	25.7	204	32.9	452	76.7
VS <sub>Y</sub>	8.4		1.8		3.6		0.85		0.15		0.13	

**Figure 6—figure supplement 1.** Summary of *Sk/Sp* hybrid and pure species diploid meiotic phenotypes and distribution of alleles in their progeny.  
DOI: 10.7554/eLife.02630.025

locus	aneuploids		haploids	
	<i>Sk/Sp rec12*</i> (n=204)	<i>Sk/Sp rec12*</i> (n=146)	<i>Sk/Sp rec12*</i> (n=146)	<i>Sk/Sp rec12*</i> (n=146)
	observed	%	observed	%
<i>his5*</i>	134	65.7	102	69.9
<i>his5</i>	70	34.3	44	30.1
<i>lys1*</i>	59	28.9	38	26.0
<i>lys1</i>	145	71.1	108	74.0

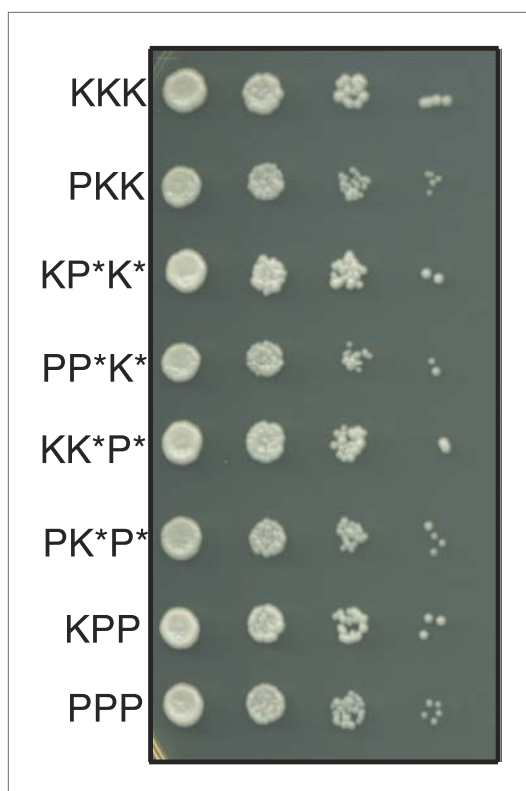
**Figure 6—figure supplement 2.** Biased transmission favoring *Sk* alleles on chromosomes 1 and 2 is observed in aneuploid and haploid spores.  
DOI: 10.7554/eLife.02630.026



**Figure 6—figure supplement 3.** Distribution of progeny from *rec12Δ Sk/Sp* hybrid meiosis.  
DOI: 10.7554/eLife.02630.027

locus	<i>Sk/Sp rec12Δ</i> (n=339)		<i>Sk/Sp rec12Δ</i> (n=341)	
	<i>Sk</i> mtDNA		<i>Sp</i> mtDNA	
	observed	%	observed	%
<i>his5</i> <sup>+</sup>	21	7.1	16	5.4
<i>his5</i> <sup>-</sup>	275	92.9	283	94.6
<i>lys1</i> <sup>+</sup>	206	69.6	221	73.9
<i>lys1</i> <sup>-</sup>	90	30.4	78	26.1
<i>ade6</i> <sup>+</sup>	9	16.7	6	10.2
<i>ade6</i> <sup>-</sup>	45	83.3	53	89.8

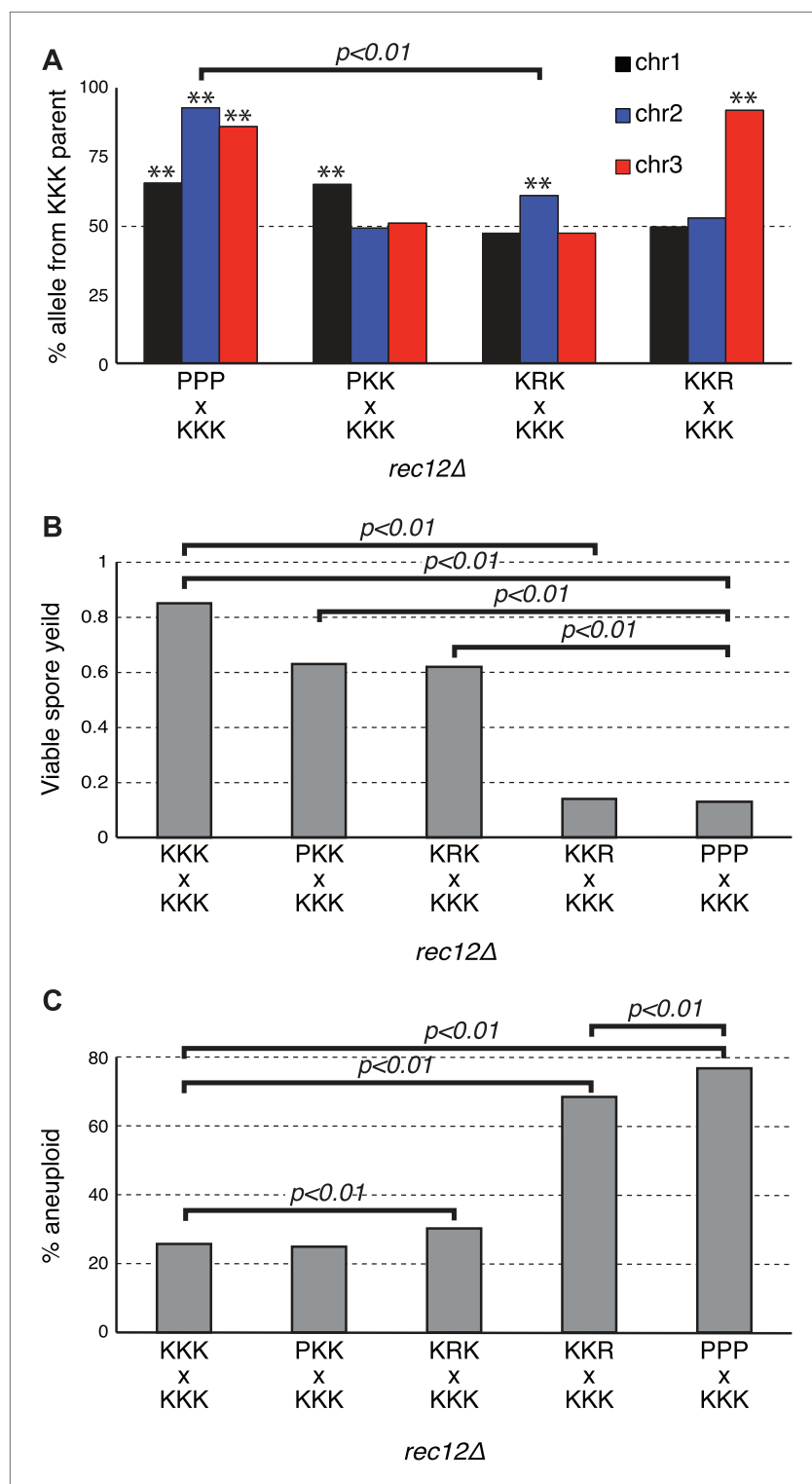
**Figure 6—figure supplement 4.** Meiotic drive in *Sk/Sp* hybrids is independent of mitochondrial DNA.  
DOI: [10.7554/eLife.02630.028](https://doi.org/10.7554/eLife.02630.028)



**Figure 7.** The haploid progeny of *Sk/Sp* hybrids have similar growth rates. The progeny of *rec12Δ Sk/Sp* hybrids with the indicated chromosomes were diluted and grown on rich YEA medium. KKK indicates the *Sk* parental genotype, whereas PPP indicates the *Sp* parental genotype. The strains were genotyped using *lys1*, *his5* and *ade6* alleles on chromosomes 1, 2, and 3, respectively. Strains that inherit intact chromosomes 2 and 3 from different species are non-viable because they lack many essential genes. However, we do rarely recover viable recombinant strains that have alleles from *Sk* chromosome 2 and *Sp* chromosome 3 (and vice versa). Potentially recombinant chromosomes are denoted with an \*. All haploid strains recovered have growth rates similar to that of the parental species, suggesting mitotic growth defects do not underlie differential recovery of the genotypes.

DOI: [10.7554/eLife.02630.029](https://doi.org/10.7554/eLife.02630.029)

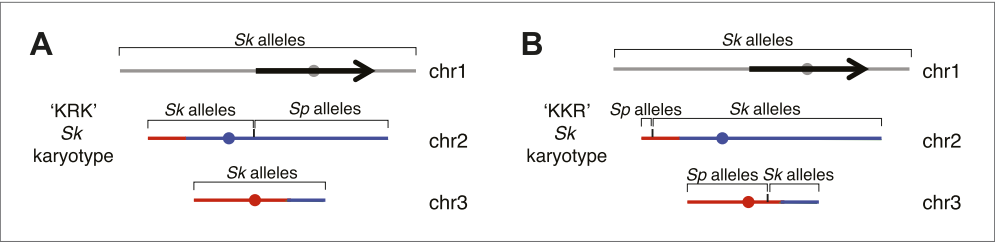




**Figure 8.** Sk drive alleles are autonomous and contribute to hybrid infertility. Aneuploidy is largely caused by heterozygosity of Sk and Sp DNA on chromosome 3. **(A)** Comparison of meiotic drive phenotypes between *rec12Δ* diploids generated by mating Sk to Sp or to haploid strains obtained from Sk/Sp hybrids. 'R' indicates a recombinant chromosome (**Figure 8—figure supplement 1**), which is compatible with all Sk chromosomes but does not contain a meiotic drive allele. All Sk chromosomes can drive autonomously (\*\* indicates drive; G-test  $p < 0.01$ ). However, the drive of Sk chr2 is lower in the KRK/KKK diploid than in pure PPP/KKK hybrids (G-test;  $n > 500$  for chromosomes 1 and 2,  $n > 80$  for chromosome 3 in each cross). The PPP/KKK data are also shown in **Figure 6B**. *Figure 8. Continued on next page*

Figure 8. Continued

(B) Fertility defects of hybrids parallel the amount of drive observed amongst the viable spores (see A, p-values obtained from t-tests, averages of at least five experiments are shown). This is consistent with drive causing spore death. (C) The high aneuploidy amongst the viable progeny of *Sk*/*Sp* hybrids is largely due to heterozygosity of one or more loci on chromosome 3 (G-test,  $n > 500$  for each cross). The PPP/KKK and KKK/KKK viable spore yield and aneuploid data are also shown in **Figure 5**. The data underlying these graphs are summarized in **Figure 8—figure supplement 2**.  
DOI: 10.7554/eLife.02630.030



**Figure 8—figure supplement 1.** Genotype of recombinant strains used in **Figure 8**.  
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locus	PPP x KKK <i>rec12Δ</i> n=589		PKK x KKK <i>rec12Δ</i> n=684		KRK x KKK <i>rec12Δ</i> n=708		KKR x KKK <i>rec12Δ</i> n=801	
	observed	%	observed	%	observed	%	observed	%
<i>lys1</i> <sup>+</sup>	350	65.2	411	65.1	364	52.7	384	50.5
<i>lys1</i> <sup>-</sup>	187	34.8	220	34.9	326	47.3	377	49.5
<i>his5</i> <sup>+</sup>	40	7.4	321	50.9	270	39.2	400	52.6
<i>his5</i> <sup>-</sup>	497	92.6	310	49.1	420	60.8	361	47.4
<i>ade6</i> <sup>+</sup>	12	14.1	226	49.0	224	47.4	19	8.9
<i>ade6</i> <sup>-</sup>	73	85.9	235	51.0	252	52.6	194	91.1
diploid	52	8.8	26	3.8	12	1.7	33	4.1
aneuploid	452	76.7	170	24.9	214	30.2	548	68.4
VS <sub>Y</sub>	0.13		0.63		0.62		0.14	

**Figure 8—figure supplement 2.** Summary of meiotic phenotypes for *Sk*/*Sp* hybrids and diploids with one heterozygous chromosome and the distribution of alleles in their viable progeny.  
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