



Figures and figure supplements

Sequestration and activation of plant toxins protect the western corn rootworm from enemies at multiple trophic levels

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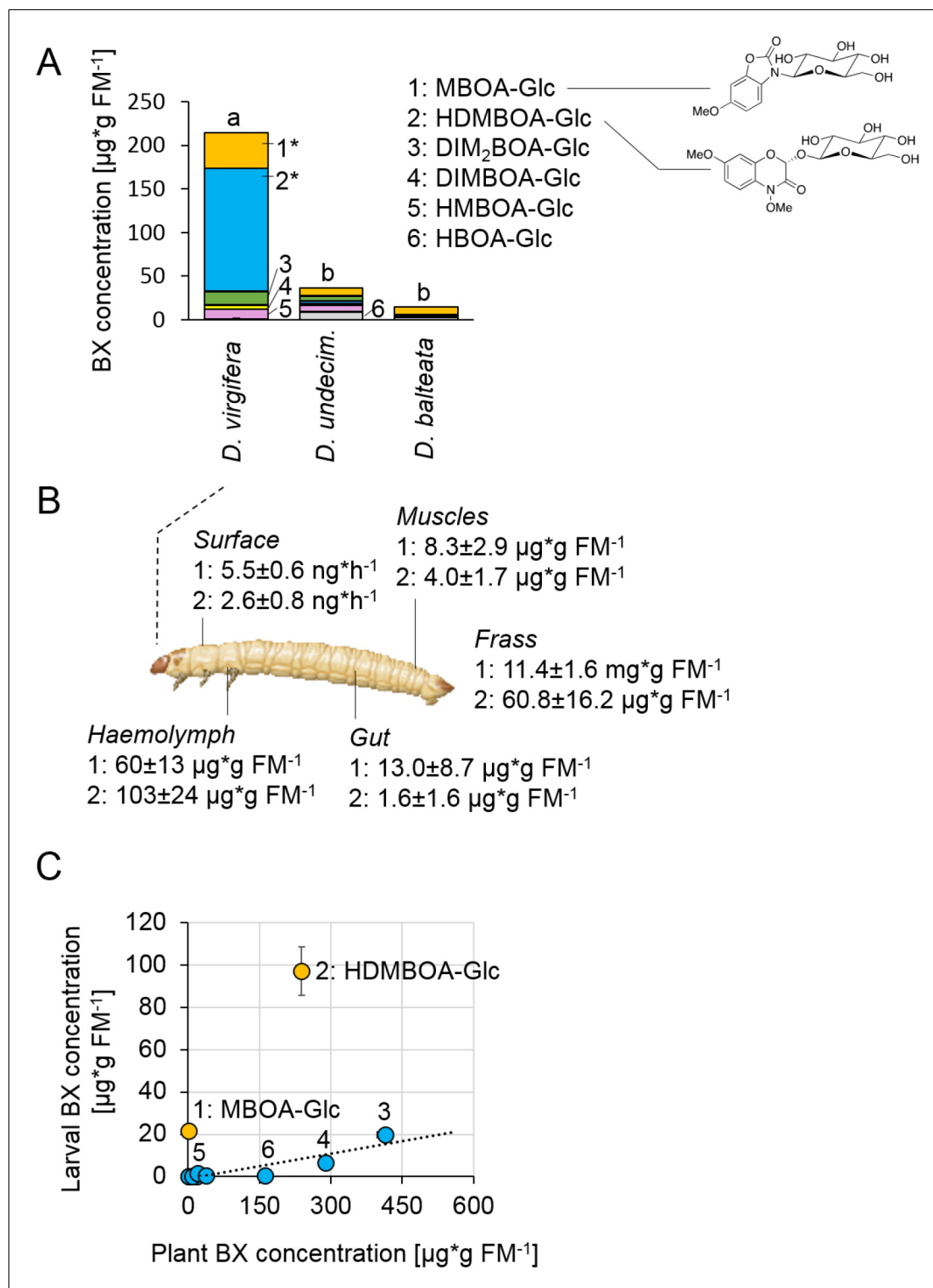


Figure 1. *Diabrotica virgifera* specifically and actively sequesters maize benzoxazinoids (BXs) (Figure 1—figure supplements 1–3). (A) BX concentration in larvae of the specialist *D. virgifera*, and the generalists *D. undecimpunctata* (*D. undecim.*) and *D. balteata*. Numbers denote the six most abundant BXs. Stars indicate significant differences between species (one-way ANOVA on transformed data (rank and square root transformations), * $p < 0.05$). (B) BX concentrations in the haemolymph, gut, muscles, exudates (surface), and frass of *D. virgifera* larvae fed on wild-type B73 plants. (C) Correlation between BX concentrations in maize B73 plants and in third instar *D. virgifera* larvae that fed on those plants since hatching. Unlabeled blue dots correspond to other types of BXs. A linear regression between plant and larval concentrations is shown ($R^2 = 0.8141$, $p = 0.004$, excl. MBOA-Glc and HDMBOA-Glc). Means \pm SE are shown. Raw data are available in Figure 1—source data 1.

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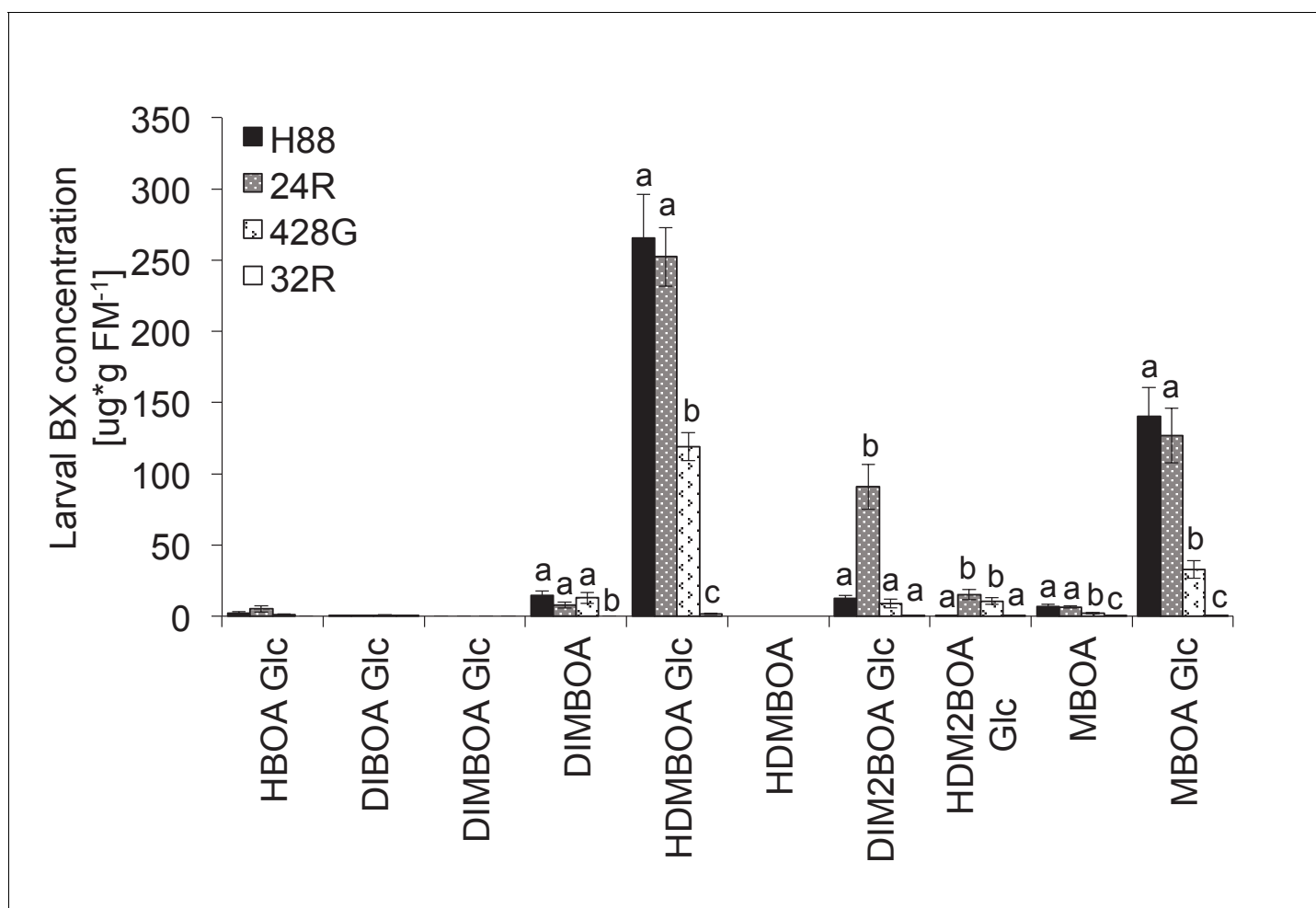


Figure 1—figure supplement 1. Benzoxazinoid levels in *Diabrotica virgifera* larvae fed on different maize lines. H88 corresponds to the wild type, 428G to the original *bx1* mutant, 24R to an *igl* mutant and 32R to a 100% BX-deficient *bx1.igl* double mutant (see [29]). Means \pm SE are shown. Letters indicate significant differences between genotypes (one-way ANOVAs, $p < 0.05$). No BXs were detected in larvae fed on 32R.

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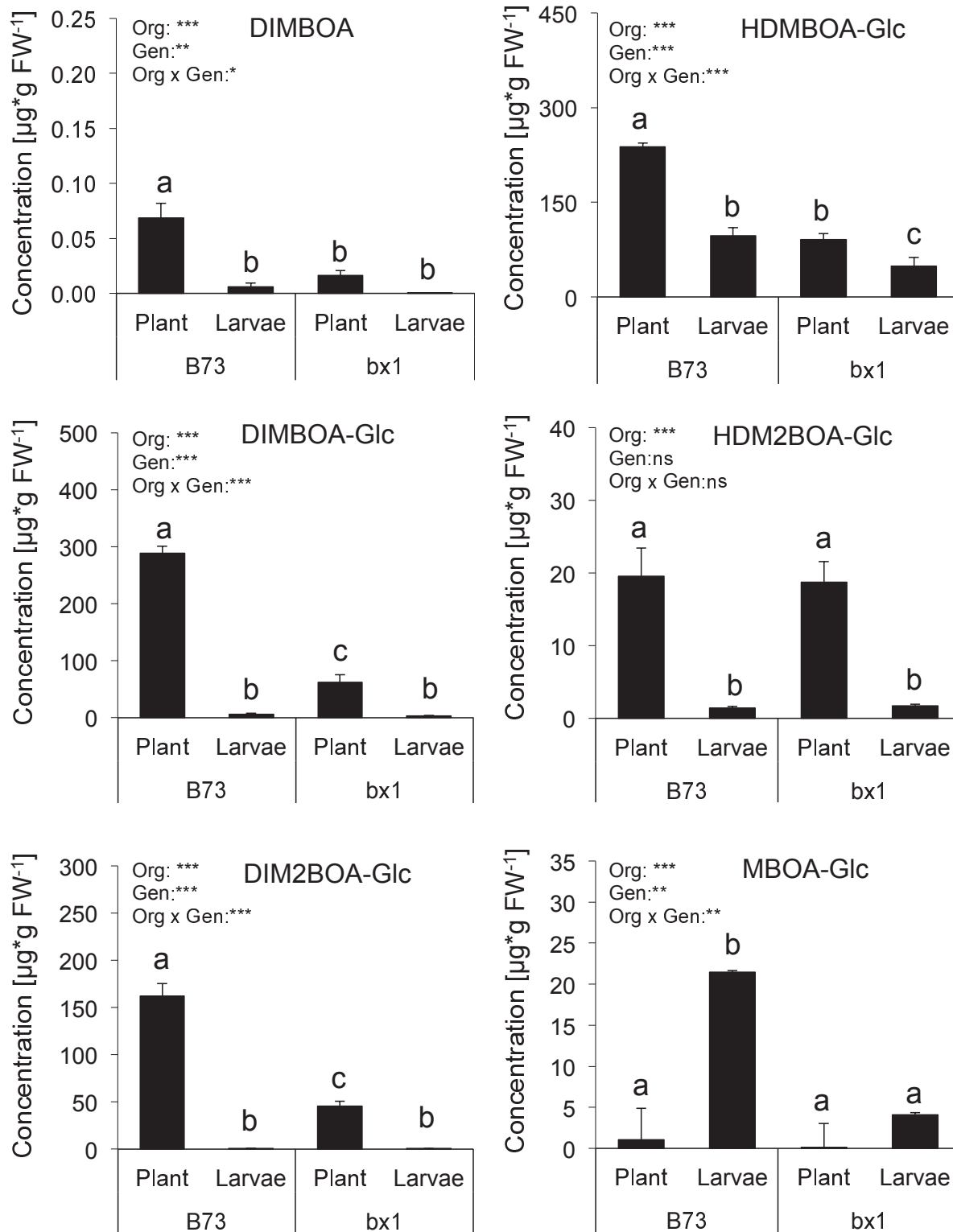


Figure 1—figure supplement 2. Benzoxazinoid levels in *Diabrotica virgifera* larvae fed wild-type (B73) and *bx1* (*bx1*:B73) mutant plants. Means \pm SE are shown. Results of two-way ANOVAs are shown (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$). Letters indicate significant differences between genotypes (post-hoc test, $p < 0.05$).

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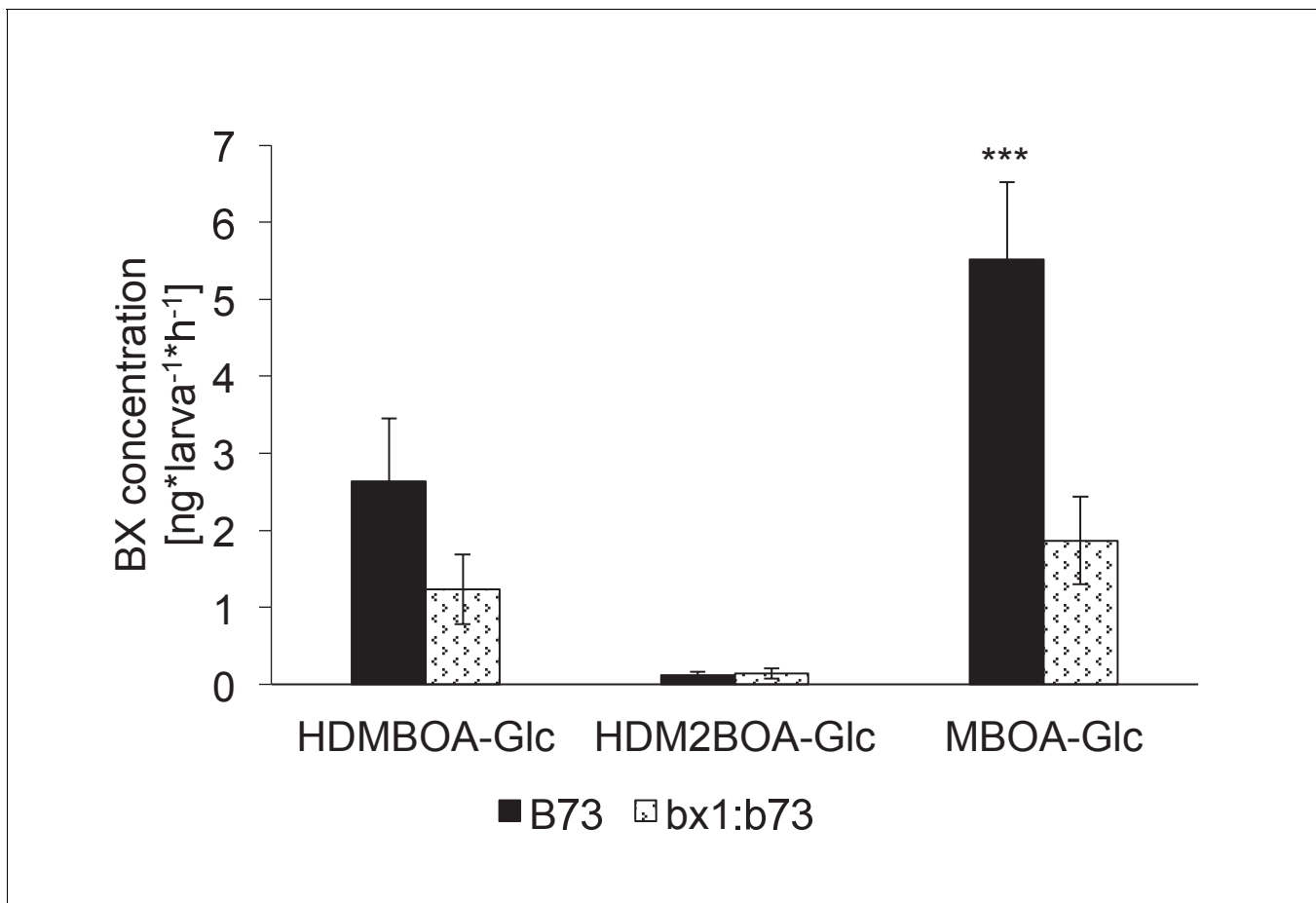


Figure 1—figure supplement 3. Benzoxazinoid levels in aqueous surface extracts of *Diabrotica virgifera* larvae fed on wild-type (B73) and *bx1* (*bx1:b73*) mutant plants. Means \pm SE are shown. Stars indicate significant differences between genotypes (Student t-test, $p < 0.001$).

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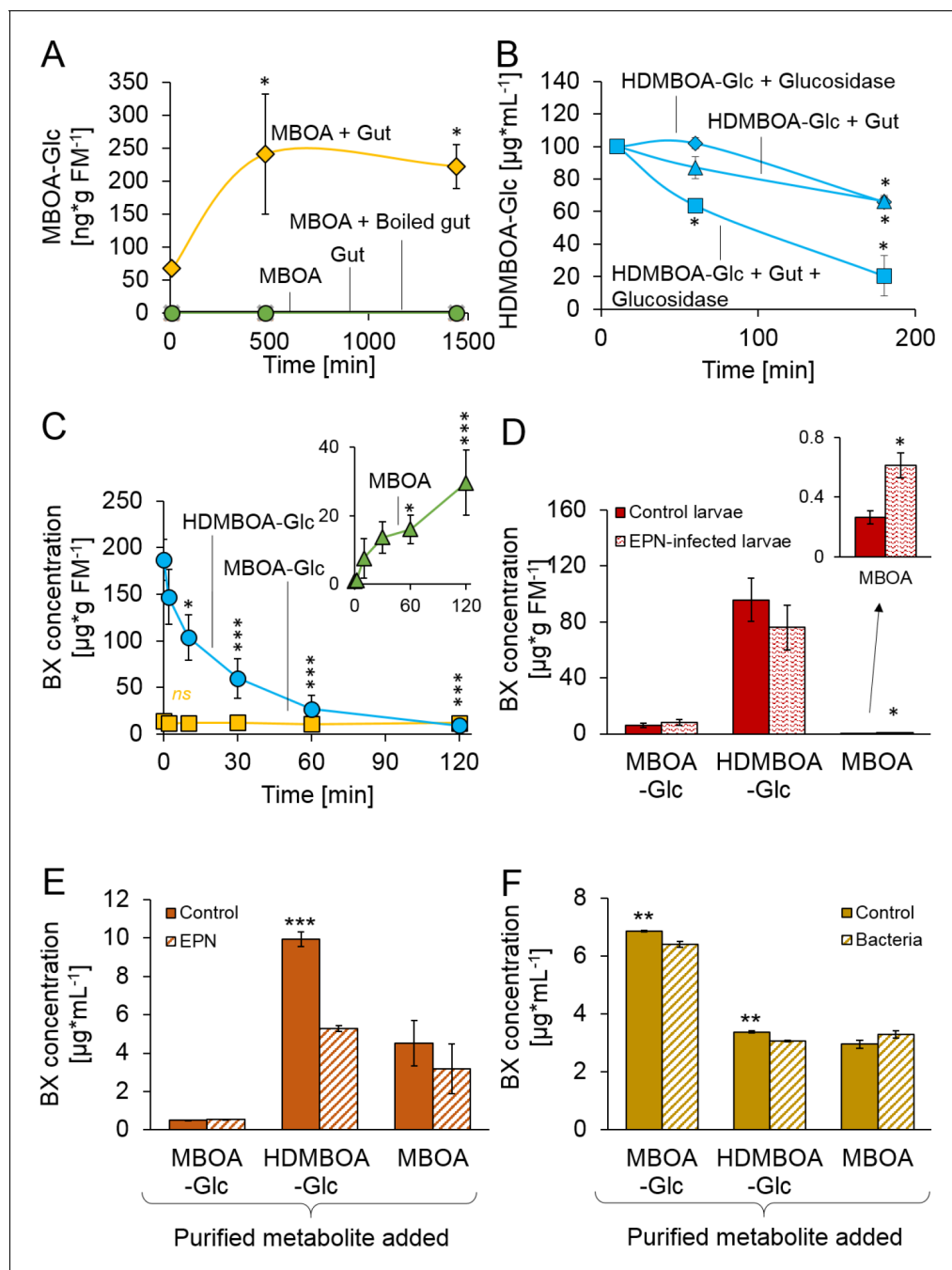


Figure 2. Stabilization and reactivation of stored benzoxazinoids (BXs) by *Diabrotica virgifera* and its natural enemies (Figure 2—figure supplement 1). (A) Stabilization of MBOA by conversion to MBOA-Glc in *D. virgifera* gut extracts. (B) HDMBOA-Glc deglycosylation in *D. virgifera* gut extracts. (C) BX reactivation in *D. virgifera* larvae upon mechanical tissue disruption. (D) BX reactivation in *D. virgifera* larvae upon exposure to the entomopathogenic nematode (EPN) *Heterorhabditis bacteriophora*. (E) BX reactivation by *H. bacteriophora* 24 hr after addition of purified metabolites. (F) BX reactivation by the EPN endosymbiotic bacterium *Photorhabdus luminescens* 24 hr after addition of purified metabolites. Means \pm SE are shown. Stars indicate significant differences between time points (repeated measures ANOVAs, A–C) or between treatments (Student's t-tests, D–F; * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$). Raw data are available in Figure 2—source data 1.

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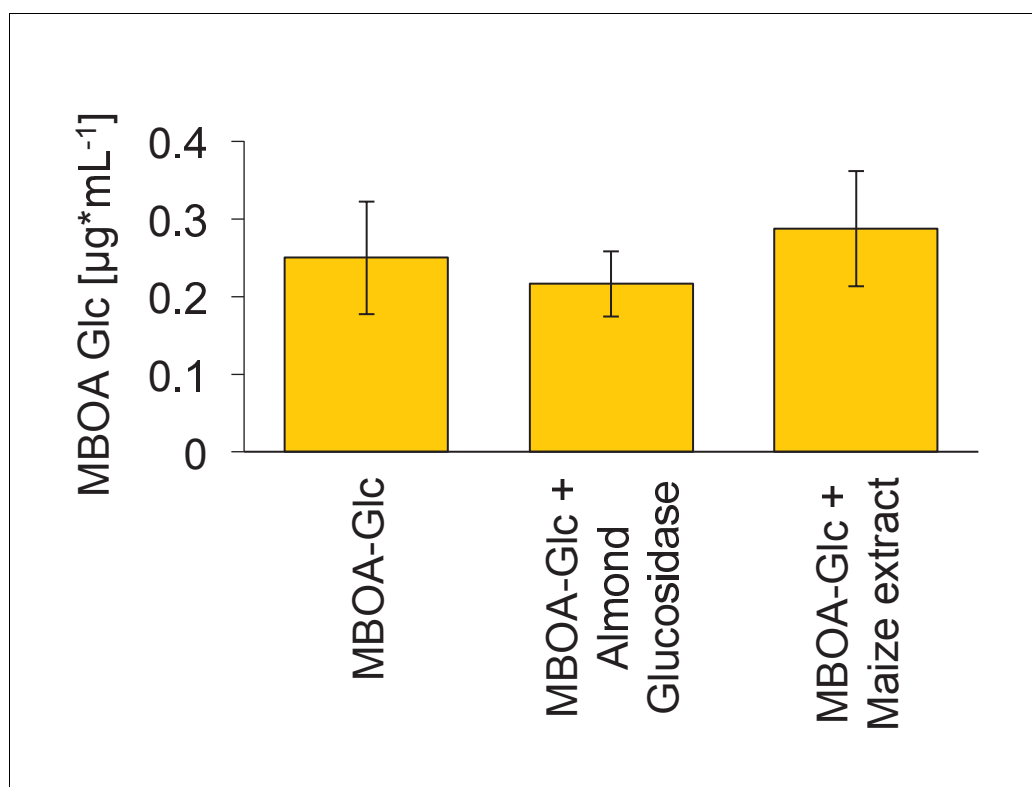


Figure 2—figure supplement 1. Degradation of MBOA-Glc by plant-derived hydrolases. MBOA-Glc concentrations upon incubation with a broad-spectrum almond glucosidase (CAS Nr. 9001-22-3) and with root extracts of a benzoxazinoid-free maize seedling (32R). Means \pm SE are shown. No significant change in MBOA-Glc was observed (one-way ANOVA).

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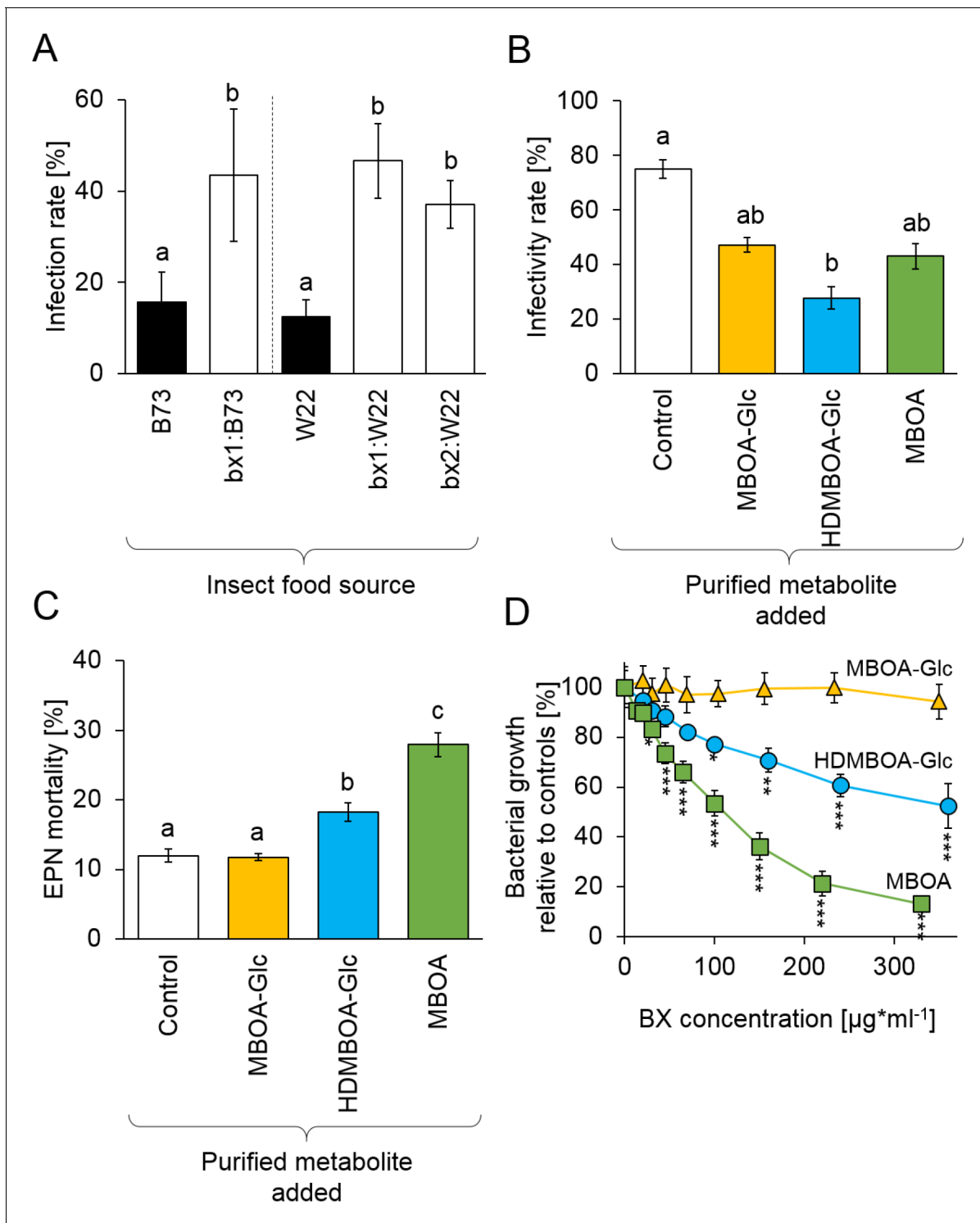


Figure 3. Benzoxazinoids (BXs) protect *Diabrotica virgifera* from its natural enemies (**Figure 3—figure supplement 1**). (A) Infection success by the entomopathogenic nematode (EPN) *Heterorhabditis bacteriophora* on *D. virgifera* larvae fed on WT (B73 and W22) or BX-deficient (bx1:B73, bx1:W22, bx2:W22) plants. (B) Effect of 7 days exposure to BXs on *H. bacteriophora* infectivity. (C) Effect of 7 days exposure to BXs on *H. bacteriophora* mortality. (D) Effect of BXs on the growth of the symbiotic entomopathogenic bacterium *Photorhabdus luminescens*. Different letters indicate significant differences between plant genotypes. Means \pm SE are shown. Stars indicate significant differences between concentrations (A-C: one-way ANOVA, D: repeated measures ANOVA, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$). Raw data are available in **Figure 3—source data 1**.

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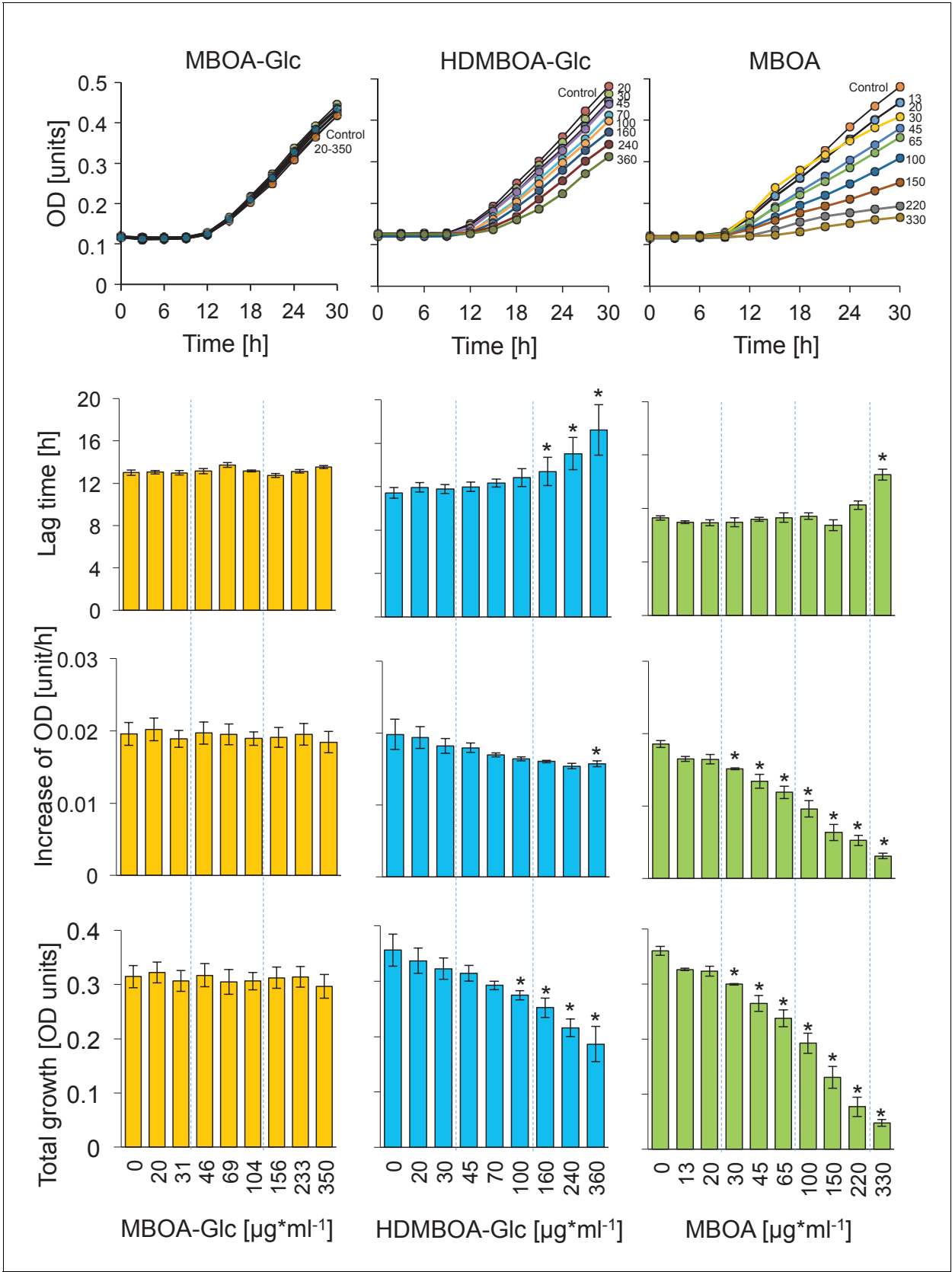


Figure 3—figure supplement 1. Growth curves and growth characteristics of *Photorhabdus luminescens* EN01 upon exposure to MBOA-Glc, HDMBOA-Glc and MBOA at different concentrations. Means \pm SE are shown. Stars indicate significant differences between genotypes (repeated measures ANOVAs, $p < 0.05$).

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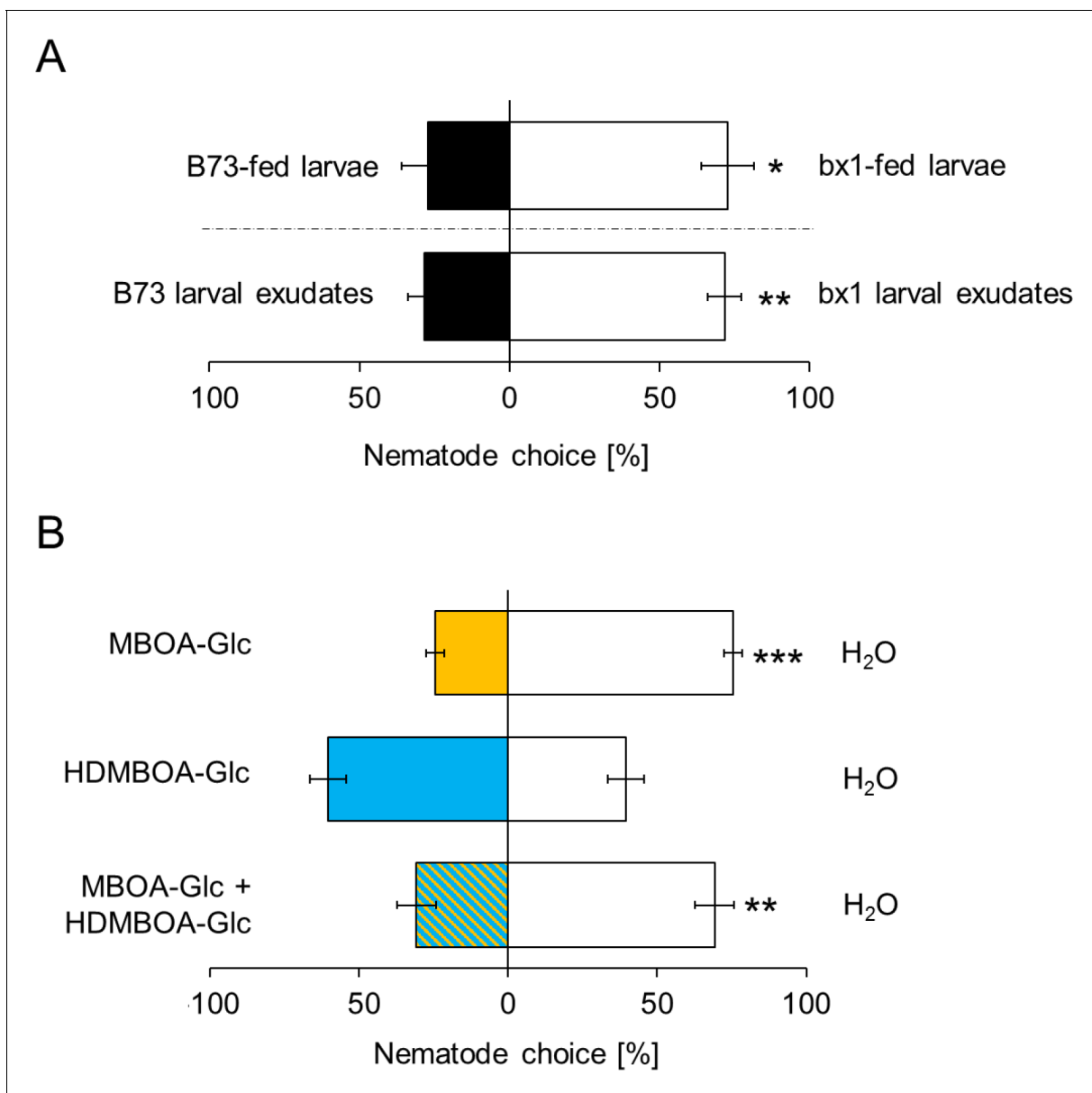


Figure 4. MBOA-Glc decreases the attractiveness of *Diabrotica virgifera* larvae. (A) Attraction of the entomopathogenic nematode (EPN) *Heterorhabditis bacteriophora* to *D. virgifera* larvae fed on wild-type (B73) and *bx1*-mutant (*bx1*:B73) (top) and aqueous surface extracts of larvae fed on wild type and *bx1*-mutant (bottom). (B) *H. bacteriophora* attraction to pure MBOA-Glc and HDMBOA-Glc at physiological concentrations. Means \pm SE are shown. Letters indicate significant differences between treatments (one sample t-tests, * p <0.05, ** p <0.01, *** p <0.001). Raw data are available in [Figure 4—source data 1](#).

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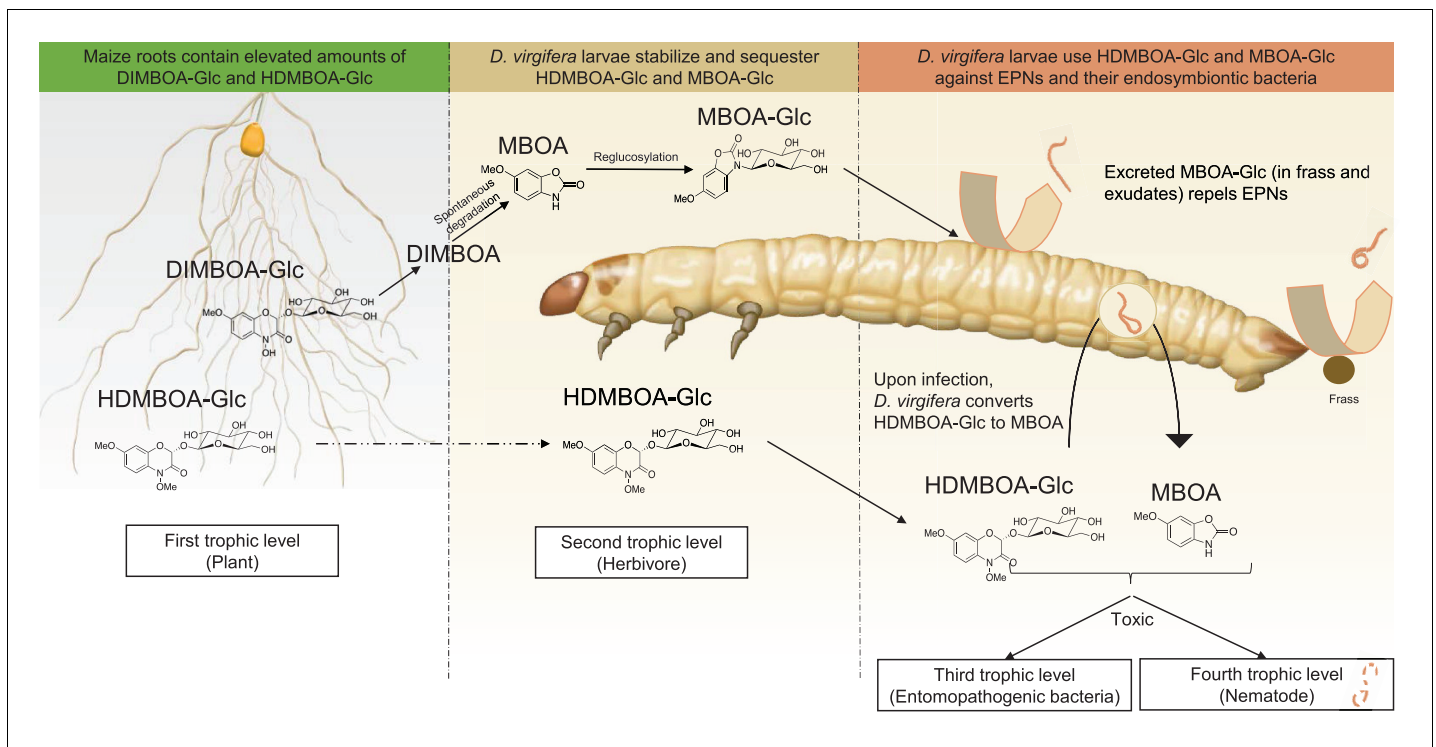


Figure 5. A model illustrating how BX sequestration and activation of plant toxins protects *Diabrotica virgifera* larvae from their enemies at multiple levels. MBOA-Glc, released in the frass and on the exoskeleton, repels infective juvenile entomopathogenic nematodes. Upon infection by nematodes and their symbiotic entomopathogenic bacteria, HDMBOA-Glc is activated to produce MBOA. Both HDMBOA-Glc and the activated MBOA reduce the growth of the symbiotic bacteria and kill EPNs.

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