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Disruption of theta-timescale spiking impairs learning but spares hippocampal replay

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eLife Assessment

This **important** study employs a closed-loop, theta-phase-specific optogenetic manipulation of medial septal parvalbumin-expressing neurons in rats and reports that disrupting theta-timescale coordination impairs performance of challenging aspects of spatial behaviors, while sparing hippocampal replay and spatial coding in hippocampal place cells. The findings are expected to advance theoretical understanding of learning and memory operations and to provide practical implications for the application of similar optogenetic approaches. The experiments were viewed as technically rigorous, but the strength of evidence provided in the current version of the manuscript was viewed as **incomplete**, mostly due to limited analyses and the descriptions of some of the experimental protocols.

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Abstract

The ability to rapidly learn and retrieve salient information about new environments is critical for survival. In mammals, the hippocampus plays a crucial role in that learning. Specialized features of hippocampal population coding, including network-level theta oscillatory activity, location-specific firing of principal cells, and reactivation of experience during immobility (replay), have been implicated in rapid storage and retrieval of spatial information. Disruptions of theta and replay jointly, or replay alone, are sufficient to impair learning; however, the specific contribution of theta-associated temporal structure during locomotion remains unknown. In this study, we manipulated hippocampal spiking activity in rats specifically during locomotion by optogenetically activating septal parvalbumin-expressing GABAergic neurons. We developed a closed-loop, theta phase-specific stimulation protocol that reliably reduced theta power shortly after stimulation onset. This manipulation preserved the place codes of individual cells but disrupted the fine temporal structure of endogenous spatio-temporal representations (for example, theta sequences) at the pairwise and population level. Theta disruption during locomotion was also sufficient to cause pronounced deficits in learning the more cognitively challenging component of a spatial alternation task, even though disruption was applied on only ~66% of trials. Notably, network effects accompanying theta disruption were restricted to

locomotor periods; we did not observe changes in replay rate, length, or content during immobility. Together, these results demonstrate the importance of the precise temporal microstructure of locomotion-associated spatial representations in the hippocampus for learning.

Introduction

Survival in novel environments is contingent on the acquisition of spatial knowledge and updating behavioral actions based on learned spatial relationships. This learning process engages numerous brain regions and circuits, with the hippocampus playing a critical role in memory-guided spatial navigation, particularly during initial learning in novel environments. Historically, foundational insights into the hippocampus' role were gained from experiments involving its complete removal or inactivation (S. M. Kim and Frank 2009 [↗](#); Morris et al. 1982 [↗](#); Scoville and Milner 1957 [↗](#); J. J. Kim and Fanselow 1992 [↗](#)). These and many other studies established a critical role for the hippocampus in learning but could not explain why disrupting the hippocampus had such profound effects.

Many efforts to address that question have focused on the population activity patterns thought to mediate hippocampal function. Individual hippocampal pyramidal neurons are active in specific locations ('place cells' firing in 'place fields') (O'Keefe 1976 [↗](#)). Place firing during movement is also closely associated with ~8 Hz theta oscillations: pyramidal cells tend to fire most during the descending phases of theta and least during the ascending phases (Mizuseki et al. 2009 [↗](#)). Furthermore, at a population level, each cycle of the theta rhythm typically engages a sequence of place cell firing (a 'theta sequence') (Skaggs et al. 1996 [↗](#); Foster and Wilson 2007 [↗](#); Burgess, Recce and O'Keefe 1994 [↗](#)). This sequence relates theta to the content of spatial representations, such that positions just behind and at the current position of the animal are represented along the descending phases of theta, while alternative "non-local" positions, such as those further ahead, are regularly expressed during ascending phases (Foster and Wilson 2007 [↗](#); Kay et al. 2020 [↗](#); Joshi et al. 2023 [↗](#); Gupta et al. 2012 [↗](#)).

Subsequently, during periods of awake immobility and sleep, the hippocampus frequently reactivates patterns associated with awake experience (Diba and Buzsáki 2007 [↗](#); Joo and Frank 2018 [↗](#); G. Buzsáki, Leung, and Vanderwolf 1983 [↗](#); Panoz-Brown et al. 2018 [↗](#); Karlsson and Frank 2009 [↗](#)). Such reactivation events, which often occur during a bout of high frequency oscillatory activity called sharp-wave ripples (SWRs), can include sequential replay of multiple place cells that together represent coherent spatial trajectories through an environment (Diba and Buzsáki 2007 [↗](#); Pfeiffer and Foster 2013 [↗](#); Kay et al. 2016 [↗](#); Karlsson and Frank 2009 [↗](#); Davidson, Kloosterman, and Wilson 2009 [↗](#)).

Theta- and SWR-related sequences are well suited to contribute to learning and memory-guided spatial navigation, both through the potential induction of plasticity in downstream structures as well as through the activation of spatially remote representations that could support flexible decision-making (G. Buzsáki 1989 [↗](#); Hasselmo 2025 [↗](#); Skaggs et al. 1996 [↗](#); Foster and Wilson 2007 [↗](#); Jadhav et al. 2012 [↗](#); Gillespie et al. 2021 [↗](#); Denovellis et al. 2021 [↗](#); Johnson and Redish 2007 [↗](#); Kay et al. 2020 [↗](#); Pastalkova et al. 2008 [↗](#); Comrie et al. 2024 [↗](#)). At the same time, the precise role of these sequences in learning and decision-making remains unclear.

Previous work has established that manipulations targeting local field potential (LFP) patterns related to these sequences can disrupt navigational behaviors. Studies related to theta have used manipulations of neurons in the medial septum (MS), a brain region critical for learning and regulating the hippocampal theta rhythm. (Mizumori et al. 1990 [↗](#); J. C. Robinson, Wilmot, and Hasselmo 2023 [↗](#)). Specifically, disruptions of normal theta oscillatory activity, either through cooling of the MS and reducing theta frequency (Petersen and Buzsáki 2020 [↗](#)), entrainment of the endogenous LFP during movement at a frequency well above the normal ~8 Hz (Zutshi et al. 2018 [↗](#); Quirk et al. 2021 [↗](#)), or scrambling of theta (Etter et al. 2023 [↗](#)), impairs performance of a previously learned task.

Disrupting SWR activity or elongating SWR duration during awake immobility also affects behavior, impairing or improving learning in a spatial alternation task respectively (Jadhav et al. 2012 [↗](#); Fernández-Ruiz et al. 2019 [↗](#)). Effects on theta sequences were not reported in these studies, but these results are broadly consistent with a role for replay and reactivation in learning. Most recently, optogenetic activation of medial entorhinal inhibitory neurons at gamma frequencies during locomotion was shown to disrupt theta sequences, subsequent sequential replay, and the ability of rats to learn to navigate efficiently among rewarded locations in a maze (Liu et al. 2023 [↗](#)). The study suggested that theta sequences are necessary to establish replay sequences, raising the possibility that disruptions of theta may affect learning through an impact on replay.

Whether it is possible to disrupt theta sequences without affecting replay, and whether such a disruption would be sufficient to impair learning, remains unknown. Addressing this question is of particular interest because hippocampal activity in the theta state, during locomotion, has been hypothesized to directly support plasticity for learning and memory (G. Buzsáki 1989 [↗](#); Skaggs et al. 1996 [↗](#); Jensen and Lisman 1996 [↗](#); Wang et al. 2015 [↗](#); Mizuseki et al. 2009 [↗](#); O'Keefe and Recce 1993 [↗](#); Hasselmo, Bodelón, and Wyble 2002 [↗](#); Comrie, Frank, and Kay 2022 [↗](#); Foster and Wilson 2007 [↗](#)). An understanding of how individual physiological features, such as these locomotion-associated theta dynamics, contribute to learning may also enable identification of therapeutic strategies for improving memory in the setting of hippocampal dysfunction (Gillespie et al. 2024 [↗](#); Atherton, Dupret, and Mellor 2015 [↗](#); Fernández-Ruiz et al. 2019 [↗](#); Martorell et al. 2019 [↗](#)).

We therefore developed a closed-loop approach to reliably manipulate hippocampal theta and associated activity during locomotion. Our approach leveraged a specific MS parvalbumin-expressing GABAergic projection, which in turn targets GABAergic neurons in the hippocampal formation (Zutshi et al. 2018 [↗](#); Quirk et al. 2021 [↗](#); Joshi et al. 2017 [↗](#); Wang et al. 2015 [↗](#); Fuhrmann et al. 2015 [↗](#); J. Robinson et al. 2016 [↗](#); Kloc et al. 2020 [↗](#); Mouchati et al. 2020 [↗](#); Lepperød et al. 2021 [↗](#); Etter et al. 2023 [↗](#); Király et al. 2023 [↗](#); Gemzik, Donahue, and Griffin 2021 [↗](#); Yong, Chang, and Brandon 2022 [↗](#); J. C. Robinson et al. 2023 [↗](#)). Using this approach, we were able to selectively suppress theta power and disrupt the sequential firing of place cells without affecting average place field activity. This manipulation, when applied in ~2/3rds of trials, was sufficient to profoundly impair learning when choices depended on memory for specific past experiences. Learning of simpler spatial trajectories remained intact, as did the expression of replay sequences. These results establish that manipulation of theta oscillatory activity and the associated temporal order of spiking is sufficient to impair learning.

Results

Medial septal (MS) parvalbumin (PV)-expressing neurons coordinate theta rhythmic activity across the hippocampal formation (Petsche, Stumpf, and Gogolak 1962 [↗](#); Freund and Antal 1988 [↗](#); Tim James Viney et al. 2018 [↗](#); Joshi et al. 2017 [↗](#)). Previous optogenetic manipulations in mice (Bender et al. 2015 [↗](#); Zutshi et al. 2018 [↗](#); Quirk et al. 2021 [↗](#); Etter et al. 2023 [↗](#)) have used optogenetic manipulations of this circuit to entrain hippocampal local field potential at experimenter-requested frequencies between 4 and 20 Hz. We similarly used a viral strategy to express a light-activated channel (ChR2) in PV+ neurons within the MS of PV-Cre transgenic rats. We also implanted an optic fiber targeting the MS as well as either silicon electrodes or multi-tetrode arrays targeting the hippocampus in $n = 10$ rats.

Each of these animals received optical stimulation restricted to periods of locomotion on either a linear track, a W-shaped track, or both (see **methods**). The stimulation pattern was governed by a protocol that either activated MS PV neurons at a wide range of requested frequencies (“rhythmic stimulation”) or at specific phases of the ongoing theta rhythm (“phase-specific stimulation”) recorded in the hippocampus (Figure 1A [↗](#)). To enable us to assess whether the effects of the manipulation were restricted to stimulation intervals, each behavioral epoch was divided into

three approximately equal periods, an initial and final stimulation interval (henceforth “*stimulation-on*”) and a middle no-stimulation control interval (henceforth “*stimulation-off*”), during which animals ran for rewards without optical stimulation (Figure 1B [↗](#)).

We confirmed that the PV cells transfected in the MS projected to the hippocampus as expected (Figure 1C [↗](#)). Histological analysis also revealed accurate targeting of both virus and optic fiber to the MS in $n=6$ animals (hereafter the “*targeted*” group) and inaccurate targeting of either virus or optic fiber in $n=4$ animals (hereafter the “*control*” group). Thus, with this experimental approach, for each experiment, we had both within-animal controls (*stimulation-on* versus *stimulation-off* intervals) and across-animal controls (*targeted* versus *control*). In a subset of animals ($n=3$), we also determined the laser power and pulse durations required to reliably entrain hippocampal LFP during locomotion and rest (see **Methods**). For each analysis, we included all animals in which the relevant manipulation was tested and that met the predefined inclusion criteria (see **Methods**).

Reliable and reversible manipulation of hippocampal theta oscillations on linear track

In targeted but not control animals, rhythmic optogenetic stimulation of MS PV neurons reliably entrained hippocampal LFP oscillations during *stimulation-on* intervals without any detectable effect on endogenous theta activity during *stimulation-off* intervals (Figure 1D [↗](#), example of 10 Hz stimulation). Correspondingly, the power spectrum of the LFP during *stimulation-on* intervals (Figure 1E [↗](#)) showed a marked peak at the targeted frequency. To quantify the degree of entrainment, we computed an *entrainment score*, which was defined as the excess LFP power observed at the requested frequency during stimulation periods compared to control periods (see **Methods**). We found a robust entrainment of the LFP at requested frequencies for targeted animals and no detectable change in control animals (Figure 1F [↗](#), example of 10 Hz). These observations were consistent across stimulation frequencies (Supplementary Figure 1 [↗](#)). These experiments replicate results from previous work in transgenic mice (Zutshi et al. 2018 [↗](#); Quirk et al. 2021 [↗](#); Etter et al. 2023 [↗](#)) and helped us optimize the optic fiber, laser power, and pulse widths required for effectively driving MS neurons in transgenic rats (see **Methods**).

Closed-loop theta phase-specific manipulation results in a rapid reduction of theta power on linear track

Spiking activity in the hippocampus is closely associated with ongoing theta oscillations, with pyramidal cells tending to fire least during the ascending phases of theta and most during the descending phases (Mizuseki et al. 2009 [↗](#)). This temporal structure is coordinated by a network of local and long-range GABAergic inputs from cortical and subcortical regions (Joshi et al. 2017 [↗](#); Varga, Golshani, and Soltesz 2012 [↗](#); Viney et al. 2018 [↗](#); Salib et al. 2019 [↗](#); Király et al. 2023 [↗](#)), each precisely timed and phase-coupled to endogenous theta oscillatory activity. A subset of rhythmic MS PV neurons, in particular, project to hippocampal axo-axonic cells (Tim J. Viney et al. 2013 [↗](#); Joshi et al. 2017 [↗](#)), and thus activation of MS PV neurons should have a disinhibitory effect on hippocampal pyramidal cells. We therefore reasoned that closed-loop activation of MS PV neurons during the ascending phase of ongoing theta (Supplementary Figure 2A [↗](#)) would introduce additional pyramidal cell spikes and disrupt normal hippocampal theta dynamics.

Consistent with this prediction, we found optogenetically activating MS PV neurons at the ascending phase of theta resulted in a marked reduction of theta power beginning approximately 200 ms after initial stimulation onset (Figure 1G [↗](#)). To quantify this relationship, we computed a *suppression score*, defined as the decrease in LFP power observed at the peak frequency during *stimulation-off* periods compared to that observed during *stimulation-on* periods (see **methods**). This measure confirmed the prominent reduction in theta power (Figure 1H [↗](#)) in targeted animals compared to control animals (Figure 1I [↗](#)). As in the rhythmic stimulation condition, these changes were restricted to stimulation times, and endogenous theta dynamics were unaffected during *stimulation-off* run periods.

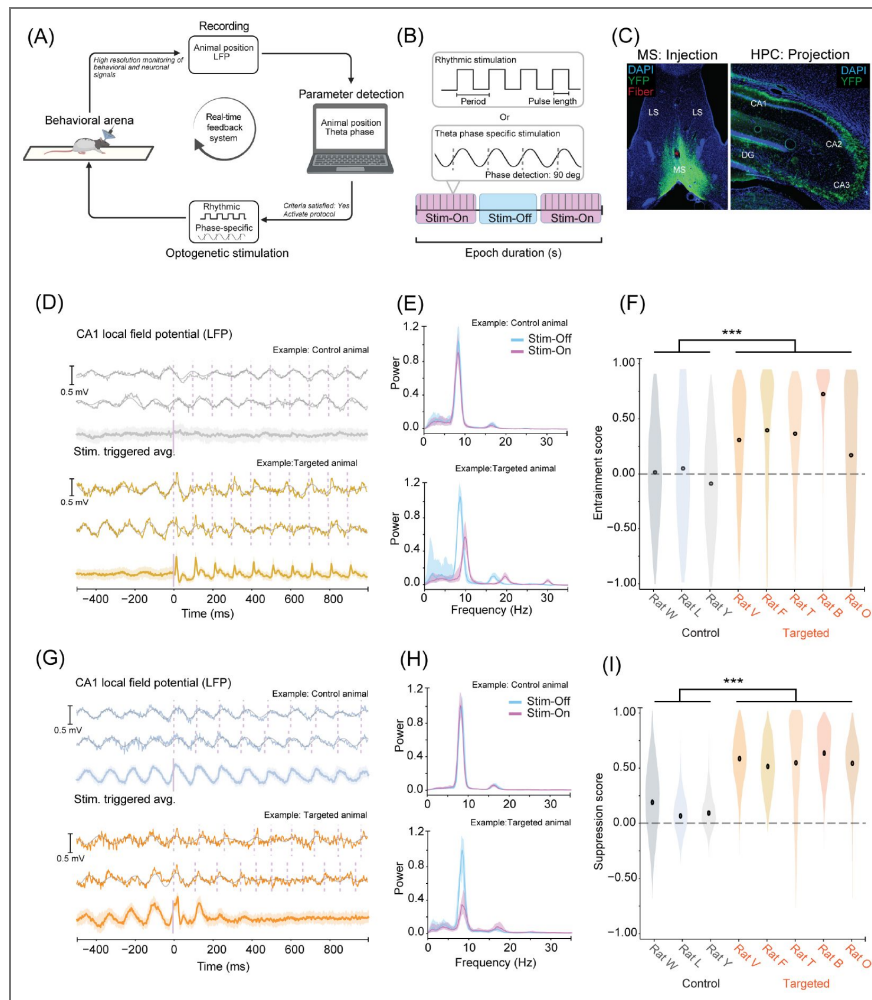


Figure 1. Reliable and reversible manipulation of hippocampal theta oscillations on a linear track.

A, Schematic of experimental approach. Rats implanted with electrodes targeted to hippocampal region CA1 and a tapered optic fiber targeted to the dorso-ventral extent of the medial septum. Optogenetic stimulation was applied only when the rats were locomoting on a behavioural arena (a linear track shown), using a real-time closed-loop system. **B**, Two types of optogenetic stimulation protocols, either rhythmic stimulation (defined by period and pulse length), or a theta phase-specific stimulation (90 degree targeting shown), was applied in each session (upper grey boxes). A day consisted of interleaved run and rest epochs of 15–20 minutes each. Run epochs included ‘test’ intervals (purple, “stimulation-on”), where the optogenetic stimulation would be applied during locomotion, and “control” intervals (blue, “stimulation-off”), where no optogenetic manipulation was applied during locomotion. **C**, Virus (AAV5 Ef1a DIO-hChR2(H134R), labeled with eYFP) targeted the medial septum (left) and long-range projecting axon fibers in the hippocampus (right). **D**, Individual examples (upper two) and epoch-averaged (lower) hippocampal CA1 LFP traces during 10 Hz rhythmic stimulation in one control animal (grey) and one targeted animal (orange) triggered by stimulation onset. Error bars show 95% confidence intervals (CI). **E**, Power spectral density of hippocampal LFP during 10Hz stimulations during stimulation-on and stimulation-off intervals in one control animal (upper) and one targeted animal (lower). **F**, Entrainment scores distributions across sessions for $n = 3$ control animals (grey) and $n = 5$ targeted animals (orange). Targeted animals show significant entrainment compared to controls (mixed effect linear model (animal, transfection state), effect of transfection; $p = 0.0015$, targeted $n = 19816$ interval pairs, control $n = 8517$ interval pairs). **G**, Individual examples (upper two) and epoch-averaged (lower) hippocampal CA1 LFP traces during theta phase-specific stimulation in one control animal (grey) and one targeted animal (orange) over time since stimulation onset. Error bars show 95% CI. **H**, Power spectral density of hippocampal LFP during theta-phase specific manipulation during stimulation-on and stimulation-off intervals in one control animal (upper) and one targeted animal (lower). **I**, Suppression score distributions across sessions for $n = 3$ control animals (grey) and $n = 5$ targeted animals (orange). Targeted animals show significant suppression compared to controls (mixed effect LM (animal, transfection state), effect of transfection $p = 2.6 \times 10^{-27}$, targeted $n = 34268$ intervals, control $n = 14250$ intervals). Note, one control animal (Rat I) lacked sufficient runs, and one targeted animal (Rat D) was not tested at the highest laser power during locomotion. Hence these animals could not be included in the entrainment and suppression score calculations.

This manipulation did not affect running velocity between *stimulation-on* intervals compared to *stimulation-off* intervals (*t*-test between the distribution of average velocities per-trial within each animal group; targeted animals: $p = 0.8$; control animals: $p = 0.5$). Thus, this theta-phase-specific manipulation selectively suppresses the extracellularly measured theta rhythm while leaving gross locomotor behavior intact.

Theta disruption alters temporal spiking of pyramidal cells, but preserves place fields

The disruption of theta oscillations could potentially affect both temporal and spatial properties of individual hippocampal pyramidal cells. Previous studies have yielded varied results, with some dissociating spatial and temporal properties through specific optogenetic manipulations (Liu et al. 2023 [↗](#)), and others reporting mixed effects (Etter et al. 2023 [↗](#)). To examine the effects on temporal and spatial coding we first verified that putative pyramidal cells in control animals were unaffected by the light pulses while cells in targeted animals showed short latency (5–10 ms), robust responses (Figure 2A–C [↗](#)). This effect is consistent with the strong disinhibitory input from the medial septum to the hippocampus (Joshi et al. 2017 [↗](#)) and the expected synaptic delays by strongly myelinated GABAergic neurons (Jones, Norris, and Henderson 1999 [↗](#)).

To examine the impact of our manipulation on the sequential temporal relationships between principal cells, we constructed cross-correlation histograms for pairs of putative place neurons (i.e., units with localized place fields; **see Methods**) during both *stimulation-on* and *stimulation-off* intervals. These histograms typically exhibit strong theta modulation, with peaks at the theta timescale; the offset of these peaks from zero reflects longer timescale differences in firing related to spatial firing fields (Dragoi and Buzsáki 2006 [↗](#); Skaggs et al. 1996 [↗](#)). For each cross-correlation, we identified the highest peak value within ± 100 ms of zero. We then pooled data across all animals and run sessions and plotted the location of each cell pair's peak during *stimulation-on* versus *stimulation-off* intervals (Figure 2D [↗](#)).

These plots revealed the expected diagonal of peak offset for control animals, reflecting consistent cross-correlation structure with or without stimulation. Consistent structure was less readily visible in targeted animals, with many cell pairs showing more synchronous activity during *stimulation-on* periods ($R^2 = 0.10$, control vs. targeted $p < 10^{-4}$ for pooled data; more conservative hierarchical bootstrap $p = 0.15$, reflecting low sample count for some animals).

By contrast, we did not detect any changes in the spatial tuning of putative pyramidal neurons. Specifically, the increased spiking during *stimulation-on* intervals respected place field boundaries (Supplementary Figure 2B [↗](#)) and the place fields of single cells were not detectably affected by theta disruption (Figure 2E–F [↗](#), single cell place fields, pooled for **G**). These results are consistent with previous studies that performed MS manipulations (Zutshi et al. 2018 [↗](#); Etter et al. 2023 [↗](#)). Thus, our manipulation protocol provides an opportunity to ask specifically how temporal coding contributes to behavior.

Theta disruption impairs learning in a novel spatial environment

We next investigated the effects of disruption of locomotion-associated hippocampal temporal dynamics on learning. Specifically, we focus on the initial learning of a hippocampal-dependent spatial alternation task in a W-track environment (S. M. Kim and Frank 2009 [↗](#)). As in previous experiments on the linear track, we maintained the *stimulation-on*, *stimulation-off*, *stimulation-on* structure (Figure 1B [↗](#)) to assess the impact of stimulation on single-cell and population-level activity. In contrast to the linear track experiments, theta-disruption was begun as soon as animals were first put on the maze, allowing us to assess the importance of theta-timescale structure for learning. Here the control group was expanded to include data from our prior work (Joshi et al. 2023 [↗](#)) that used the same training protocol.

Despite the interleaved *stimulation-off* periods, targeted animals had pronounced deficits in learning the spatial alternation task. We first examined the more cognitively challenging outbound trials in which animals must remember their previous outer arm visit to make the next

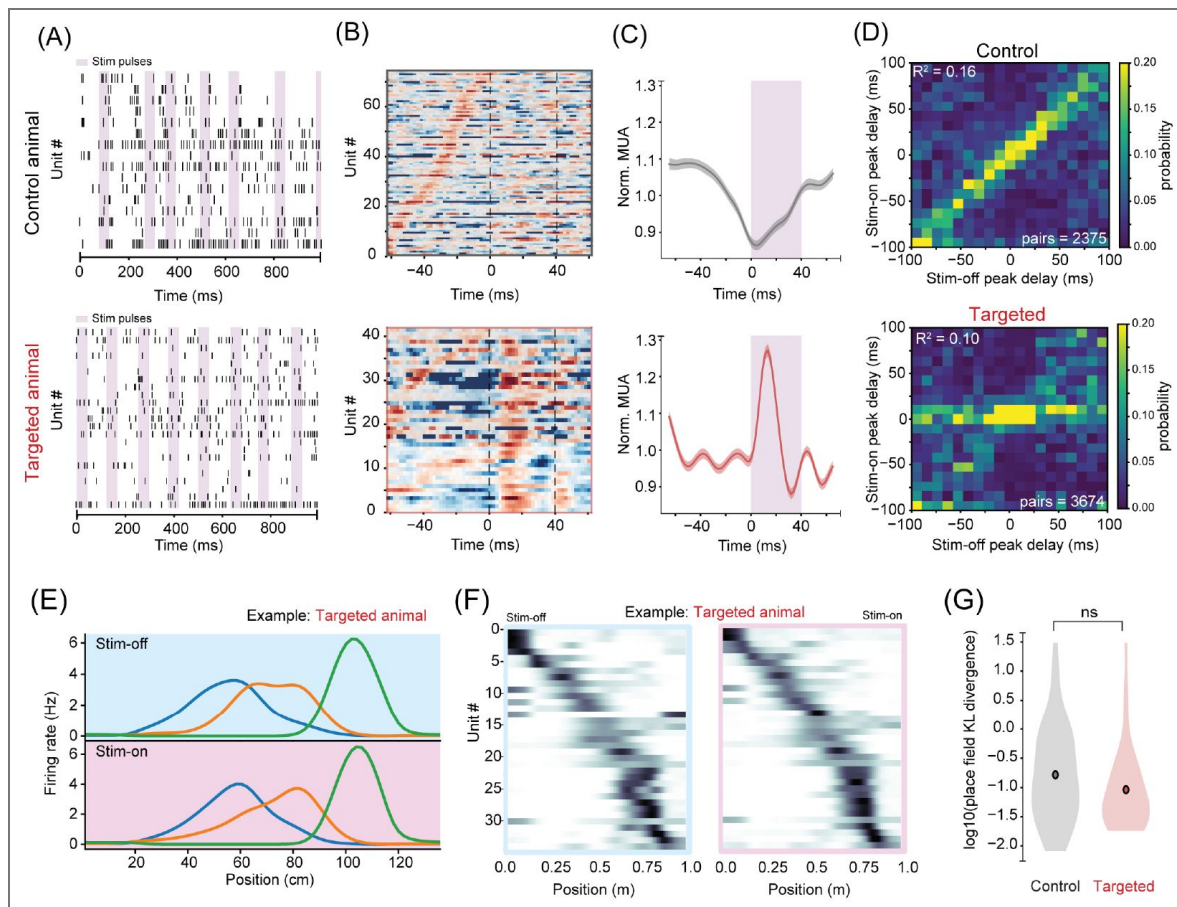


Figure 2. Theta disruption impairs temporal coding of pyramidal cells, but preserves place code on linear track.

A, Raster plots of simultaneously recorded hippocampal neurons ordered by the peak of their place fields with MS optogenetic theta phase-specific stimulation pulses highlighted (purple) in one control (top) and one targeted (bottom) animal. Note that in control rats, endogenous theta sequences are apparent. **B**, Stimulation-triggered averages of simultaneously recorded hippocampal neurons show the absence of consistent stimulation-triggered organization in control animal (top) and entrainment of spiking by optogenetic activation in targeted animal (bottom). Vertical dashed lines indicate stimulation time. Firing rates (normalized) shown in heatmap. **C**, Stimulation-triggered MUA across all control ($n = 499$ units, 4815 pulses, top) and targeted ($n = 583$ units, 5143 pulses, bottom) animals shows distinct MUA spiking structure during stimulation. In control animals, the expected relationship is observed, with spiking probability decreasing during stimulation times that occur in the ascending theta phases, whereas in targeted animals spiking probability increases ~ 10 ms after stimulation onset. Shaded regions are 95% CI of the mean. **D**, Pairwise temporal offsets between cross-correlograms computed in *stimulation-on* versus *stimulation-off* intervals in control ($n = 3674$ pairs, top) and targeted ($n = 2375$ pairs, bottom). Note, a strong diagonal indicates that the cross-correlation structure is maintained in control animals (as expected). Control vs. targeted $p < 10^{-4}$ for pooled data; hierarchical bootstrap $p = 0.15$. **E**, Examples of three place cells' spatial selectivity in targeted animals during *stimulation-off* (blue) versus *stimulation-on* (purple) intervals. Note similar spatial tuning patterns. **F**, Spatial stability across all identified place cells in one targeted animal during *stimulation-off* (left, blue box) and *stimulation-on* (right, purple box) periods in one targeted animal. Note similar spatial tuning patterns. **G**, Place field stability is measured for all units as the KL divergence (see Methods) of their spatial firing selectivity between *stimulation-on* and *stimulation-off* intervals (t -test of distributions of KL divergence between targeted ($n = 35$ units satisfying inclusion criteria across $n = 3$ animals) and control ($n = 43$ satisfying inclusion criteria across $n = 4$ animals) animals, $p = 0.8$).

correct choice. While control animals readily learned to alternate correctly, targeted animals were profoundly impaired (Figure 3A, 3B [↗](#), *t*-test performance accuracy, $p=0.01$). By contrast, we did not observe any significant differences in learning across groups on the interleaved inbound trials, where a simpler place–action association is sufficient to enable learning (Figure 3C, 3D [↗](#), *t*-test, performance accuracy, $p=0.126$).

To further characterize the selective deficit on outbound trials we calculated the probability that animals revisited the same reward port on consecutive trials (“repeats,” Figure 3E [↗](#)), which reflects a violation of the task rules and never results in reward. Control animals showed a strong bias to avoid visits to the immediately previously visited arm and thus had a distribution of repeated visits very different than random. By contrast, the targeted animals’ outbound choices were not distinguishable from random, and thus showed no overall history dependence.

This learning disruption on the W-track could not be explained by consistent changes in motivation: both targeted and control animals completed >300 trials, sufficient to measure learning (Jadhav et al. 2012 [↗](#); Joshi et al. 2023 [↗](#)). There were similarly no significant differences in average running speed or choice point occupancy during locomotion, but targeted animals did move more slowly and spend more time near the choice point on outbound trials (Figure 3F [↗](#), Video 1, Speed targeted: median: 38.5 cm/s (IQR 25.8–47.1); control: median 46.44 cm/s (IQR 32.8–55.1), linear mixed effects model: effect of transfection; $p=0.68$; interaction (transfection x trial type; $p<0.05$); occupancy targeted: median: 1.5 s (IQR 1–2.5), control: 0.7 s (IQR 1–1.7), linear mixed effects model: effect of transfection; $p=0.94$, interaction (transfection x trial type; $p<0.05$). This pattern is consistent with the targeted animals having greater choice uncertainty on outbound trials.

Disruption of theta sequences in the W-track

We then asked how our manipulation affected the structure of the spiking activity on the W-track. We first verified that, as for the linear track (Figure 2C [↗](#)), stimulation drove an increase in spiking in the targeted, but not the control group (Supplementary Figure 3A [↗](#)). We next examined the consistency of pairwise cross-correlation peaks between *stimulation-on* and *stimulation-off* periods as we did for the linear track (Figure 2D [↗](#)).

Here again we observed more consistent cross-correlation structure in control animals (linear fit $R^2 = 0.09$), and less consistent structure in targeted animals ($R^2 = 0.005$; Supplementary Figure 3B [↗](#)). These differences were highly significant when data were pooled across animals and run epochs (control vs. targeted $p < 10^{-4}$) and were also significant with the more conservative hierarchical bootstrap ($p < 10^{-3}$; Supplementary Figure 3C [↗](#)).

The presence of a persistent learning deficit on outbound trials despite *stimulation-off* periods suggested the possibility that the suppression of theta in the W-track led to persistent deficits in the temporal structure of hippocampal activity. We therefore compared the structure of activity across control and targeted animals for both *stimulation-on* and *stimulation-off* periods. We plotted, for each cell pair, the peak of the theta time-scale cross-correlation on the y-axis and the peak of the behavioral (~1 second) time-scale cross-correlation on the x-axis (see Methods).

These plots revealed a persistent deficit in the temporal organization of spiking in targeted animals present in both *stimulation-on* and *stimulation-off* periods (Supplementary Figure 3D [↗](#)). In control animals, these plots showed the expected correlation between short- and long-timescale peaks consistent with theta sequences ($R^2 = 0.077$) but this correlation was largely absent in the targeted animals ($R^2 = 0.007$; pooled comparison $p < 10^{-4}$, hierarchical bootstrap $p < 0.05$, Supplementary Figure 3E [↗](#)). Thus, disruption of theta during stimulation periods was sufficient to disrupt sequential spiking activity even after stimulation was turned off.

Finally, we also verified that, as in the linear track (Figure 2G [↗](#)), this disruption was confined to fine timescale activity: as for the linear track, place fields remained similar across *stimulation-on* and *stimulation-off* periods in both control and targeted animals (Supplementary Figure 3F [↗](#)).

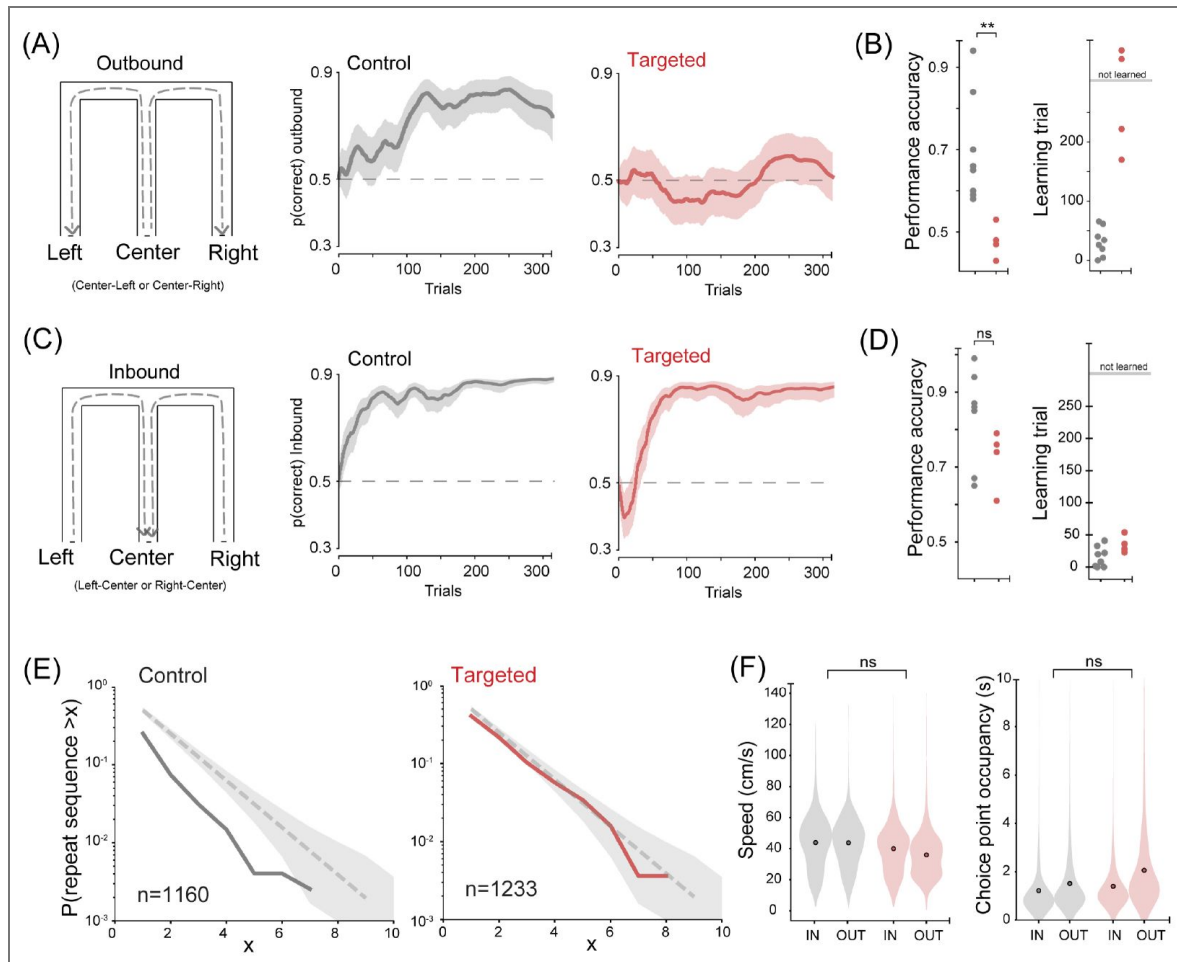


Figure 3. Theta disruption impairs learning of outbound trial structure in a novel W-track environment.

A, W-track schematic for Outbound trials (left). Learning curve for control animals ($n = 4$, middle, grey) and targeted animals ($n = 4$, right, red) on Outbound trials. **B**, Performance accuracy (left) and learning trial (right) for Outbound trials ($n = 8$ control animals, grey; $n = 4$ targeted animals, red; 4 control animals from this study, 4 control animals from Joshi et al., 2023). Performance accuracy t -test $p = 0.010$. **C**, W-track schematic for Inbound trials (left). Learning curve for control animals ($n = 4$, middle, grey) and targeted animals ($n = 4$, right, red) on Inbound trials. **D**, Performance accuracy (left) and learning trial (right) for Inbound trials ($n = 8$ control animals, grey; $n = 4$ targeted animals, red; $n = 4$ control animals from this study; $n = 4$ control animals from Joshi et al., 2023). Performance accuracy $p = 0.126$. **E**, Probability of “repeat” error trials (re-visiting a reward port on outbound trials) in control animals (grey solid line, left, $n = 1160$ trials) and targeted animals (right, red solid line, $n = 1233$ trials) compared to the 95% CI (grey shaded region) of an analytical chance distribution. Targeted animals’ choices are not different from the chance distribution. **F**, *Left*: Distribution of running speeds in control (grey) and targeted (red) animals on inbound and outbound trials (linear mixed effects model: effect of targeting, $p = 0.61$; interaction (targeting $\times$ trial type), $p = 6.52 \times 10^{-4}$) during time spent outside reward ports. *Right*: Distribution of occupancy times around the choice point (right) in control (grey) and targeted (red) animals on inbound and outbound trials (linear mixed effects model: effect of targeting, $p = 0.94$; interaction (targeting $\times$ trial type), $p = 1.39 \times 10^{-4}$).

Thus, the disruption of theta during *stimulation-on* periods leads to disruptions of the fine-timescale temporal structure throughout the W-track experiences while leaving place fields unaffected.

These findings motivated a population-level approach to better understand the impact of theta disruption on representations of position on the W-track. We used a clusterless state-space decoding algorithm to estimate, for each 2 ms bin, the probability distribution of represented locations on the track (Denovellis et al. 2021 [↗](#)).

Typical patterns of spatial representation were readily visible in control animals. There was pronounced rhythmicity reflecting theta sequences: each theta cycle typically begins with a representation close to the animal which then sweeps forward, resetting once the next theta cycle begins (Figure 4A [↗](#)) (Johnson and Redish 2007 [↗](#); Foster and Wilson 2007 [↗](#); Kay et al. 2020 [↗](#); Joshi et al. 2023 [↗](#)). We quantified the prevalence of theta-timescale position sweeps by first calculating the distance (in 2 ms bins) between the peak of the posterior of the decoded representation and the animal's actual position (ahead/behind distance). We then computed the power spectrum of this distance separately for outbound and inbound trials, given the distinct cognitive demands associated with each trial type. In the control animals, this power spectrum shows the expected pronounced peak around 8 Hz in both *stimulation-on* and *stimulation-off* periods (Figure 4B [↗](#)). We also quantified the changes in the probability of observing continuous decodes relative to stimulation times during *stimulation-on* periods. These probabilities showed the expected decrease in continuity associated with late phase theta, where the representation can jump back to a location at/or behind the actual location of the animal (Figure 4C [↗](#)). Similarly, the peri-stimulation patterns of change in entropy, which measures the concentration of the decoded distribution, showed the expected increase seen during late phase theta where spatial representations are less concentrated (Figure 4D [↗](#)).

The theta-time scale rhythmic features of the population representation of space were disrupted in targeted animals, even as the decode remained on average close to the animal (reflecting the preservation of place field structure) (Figure 4E [↗](#)). Moreover, the rhythmic features were disrupted during both *stimulation-on* and *stimulation-off* periods. The ~8 Hz peak in the power spectrum of the ahead/behind distance was much less visible on outbound trials and was absent on inbound trials (Figure 4F [↗](#)). Further, the consistent modulation of both the probability of a continuous decode state and of the entropy of the posterior distribution around stimulation times was also much less prevalent in targeted animals (Figure 4G, H [↗](#)).

We focused on the power spectrum to quantify these differences across control and targeted animals, across outbound and inbound trials, and across *stimulation-on* and *stimulation-off* periods. Consistent with the pairwise analyses (Supplementary Figure 3B-D [↗](#)), the ~8 Hz peak in the power spectrum of the ahead/behind distance was significantly larger in control animals than in targeted animals across all conditions (Figure 4I [↗](#); pooled comparison p 's $< 10^{-4}$, hierarchical bootstrap p 's < 0.05). At the same time, the maximal ahead/behind distances near the choice point, where choices must be made on outbound and inbound trials, did not differ between control and targeted animals (Supplementary Figure 3G [↗](#)). Thus, our findings indicate that the precise timing of non-local representations during theta was disrupted, but the spatial extent was not.

Finally, the differences in the power spectrum also motivated us to return to the linear track data and examine the impact of rhythmic frequency stimulation on decoded representations. To our surprise, the peak of the power spectrum of the ahead/behind distance appeared to shift systematically with the applied stimulation frequency (Supplementary Figure 4 [↗](#)). This suggests a surprising level of preserved representational movement across frequencies ranging from 6 to 12 Hz.

Theta disruption spares replay during sharp wave ripples

A previous study that used an optogenetic intervention in the Entorhinal Cortex to disrupt hippocampal theta sequences reported that doing so also disrupted the sequential structure of replay events seen during subsequent offline periods (Liu et al. 2023 [↗](#)). We therefore asked

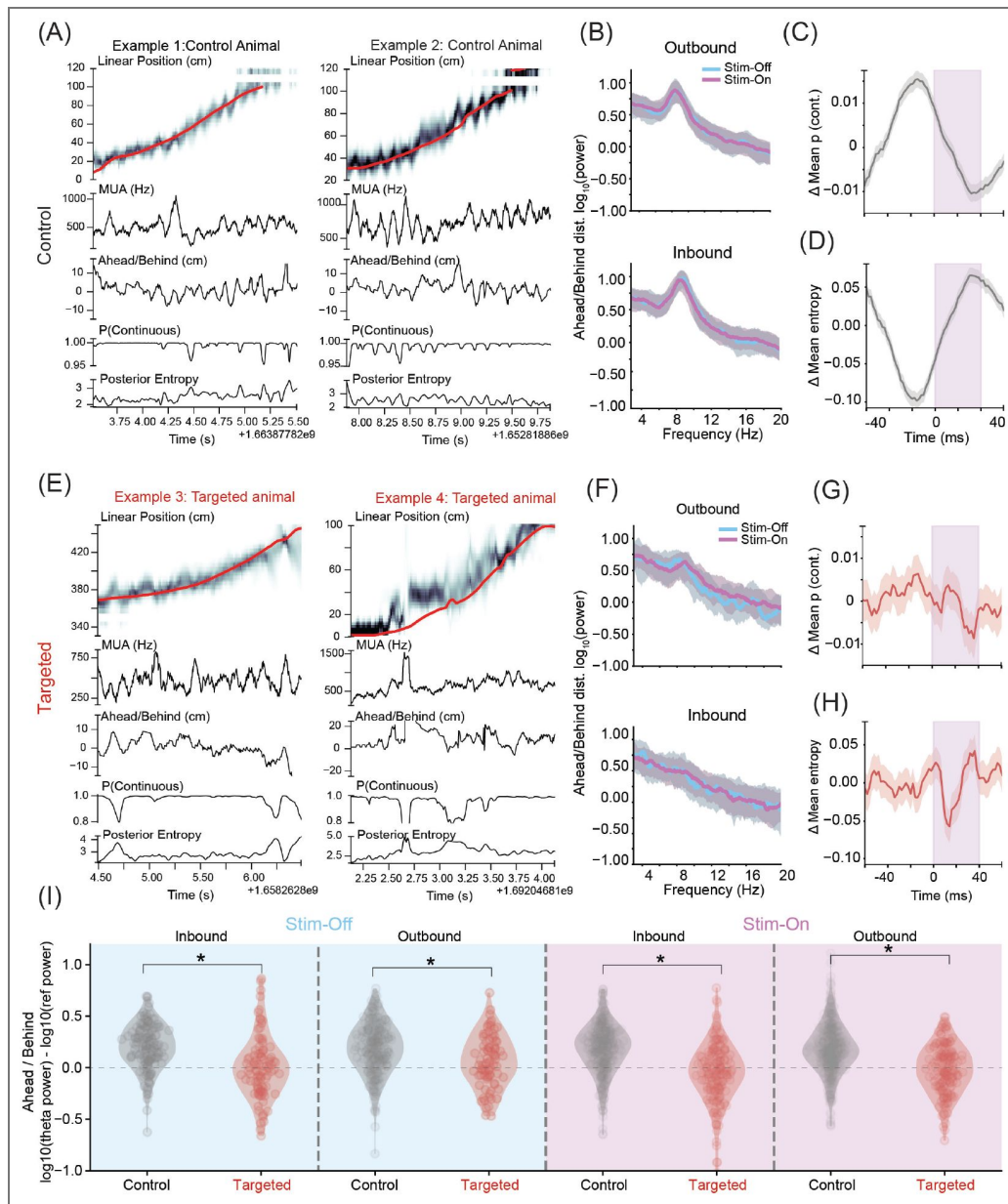


Figure 4. Theta disruption suppresses fast timescale sequential position representations.

A, Examples of decoding position from population spiking on outbound trials in control animals. From top to bottom: actual linearized position (red) and estimated position posterior (greyscale); multiunit spike rate (spikes/s); ahead/behind distance of decoded position to animal's actual position (cm); probability of a spatially continuous decoding state; and entropy of posterior over space. **B**, Power spectrum of the ahead/behind distance for *stimulation-on* and *stimulation-off* intervals across all trials in control animals, separated by trial type (top: outbound; bottom: inbound). Note the clear peak near 8 Hz reflecting the presence of robust theta sequences. **C**, Quantification of stimulus-triggered probability of the posterior to be continuous during *stimulation-on* trials for control animals. Note prominent theta-timescale structure reflecting more continuous representations in early phases of theta. **D**, Quantification of stimulus-triggered posterior entropy across all trials in control animals. Note increases in entropy toward later phases of theta corresponding to stimulation times. **E**, Examples of decoding position from population spiking on outbound trials in targeted animals; plot elements as in **A**. Note the disruption in theta-timescale structure. **F**, Power spectrum as in **B** for targeted animals. Note suppression of theta-timescale peak on outbound trials (top) and absence of obvious structure on inbound trials. **G**, Probability of continuous state as in **C** for targeted animals. Note altered distribution as compared to **C**. **H**, Entropy of posterior as in **D** for targeted animals. Note altered distribution as compared to **D**. **I**, Quantification of relative power in the power spectrum of the ahead/behind distance in the 8–12 Hz band relative to the mean of the 4–8 and 12–15 Hz bands. All comparisons were significant (all pooled data pairwise *t*-tests: $p < 10^{-4}$; all hierarchical bootstrap tests: $p < 0.05$).

whether our manipulation could provide a dissociation between theta sequences and subsequent replay, or whether both were affected.

We focused on replays seen during pauses in behavior on the track and found that despite the temporal disruption of spiking in the hippocampus during locomotion, SWRs and associated replays were not detectably affected. Replay events in both control and targeted animals showed the expected continuous progression of positions over time, along with increases in multiunit firing rates and power in the ripple band (Figure 5A). Moreover, we were able to identify clear replay events during the stimulation period within the first exposure to the track in both control and targeted animals (Figure 5A bottom row).

We then quantified SWR and replay properties based on three parameters that have been linked with hippocampal-dependent learning: SWR rate (Ego-Stengel and Wilson 2010), duration (Fernández-Ruiz et al. 2019; Vancura et al. 2023), and content (Pfeiffer and Foster 2013). There were no consistent changes in the SWR rate and length between control and targeted animals (Figure 5B). Next, we evaluated whether the content of the replay of neural activity may be altered in targeted animals. Here again, we found no evidence that theta disruption affected replay structure: sequential replays reflecting non-local spatial locations were observed during awake immobility with similar frequency across control and targeted animals (“continuous state,” Figure 5C). Finally, we measured the speed of representation movement across >50 ms long continuous periods (Figure 5D). These speeds showed the expected concentration around 10 m/s (Davidson, Kloosterman, and Wilson 2009) and were not different across control and targeted animals. Thus, our manipulation specifically disrupted the temporal order of neural activity in single CA1 cells and their sequential population representations during locomotion.

Discussion

We developed a targeted theta-phase-specific manipulation of hippocampal neural dynamics that selectively impacted sequential activity during locomotion while preserving immobility-related neural dynamics. We found that activating medial septal PV+ neurons at the ascending phase of theta during locomotion disrupted pairwise temporal properties and population-level spatial representations in the hippocampus, yet maintained the individual cell place code. This manipulation, applied during the first and final third of each run epoch, led to a profound impairment in learning the more cognitively demanding trials of a hippocampal-dependent task while sparing learning on the interleaved trials where a less cognitively demanding choice was required. Population neural activity patterns during immobility remained intact, indicating that locomotion-associated theta oscillations and immobility-associated replay mechanisms are functionally separable. Notably, despite this preservation of replay, learning deficits were as severe as those observed in a complete hippocampal lesion (S. M. Kim and Frank 2009).

Our results extend findings from prior manipulations of hippocampal neural activity via medial septal manipulation (Wang et al. 2015; Fuhrmann et al. 2015; J. Robinson et al. 2016; Zutshi et al. 2018; Kloc et al. 2020; Mouchati et al. 2020; Quirk et al. 2021; Etter et al. 2023; Gemzik, Donahue, and Griffin 2021). While previous studies measured the impact of septal manipulation on theta, these studies did not systematically examine the fine timescale structure of population representations. Here we find clear evidence that driving the septal parvalbumin neurons at requested frequencies (rhythmic stimulation condition) drives hippocampal spatiotemporal sequences at that requested frequency. This finding is broadly consistent with the results of medial septum cooling, which slows both theta and the speed of progression of spatial representations in each theta cycle (Petersen and Buzsáki 2020).

We also found a profound learning impairment for the outbound trials of our alternation task in an initially novel environment. This complements the demonstrations of impairment in performance of a previously learned task in previous studies (Zutshi et al. 2018; Quirk et al. 2021; Etter et al. 2023; Petersen and Buzsáki 2020). In our work, we extend those studies to show that SWR rate, length and content are not impacted by theta disruption.

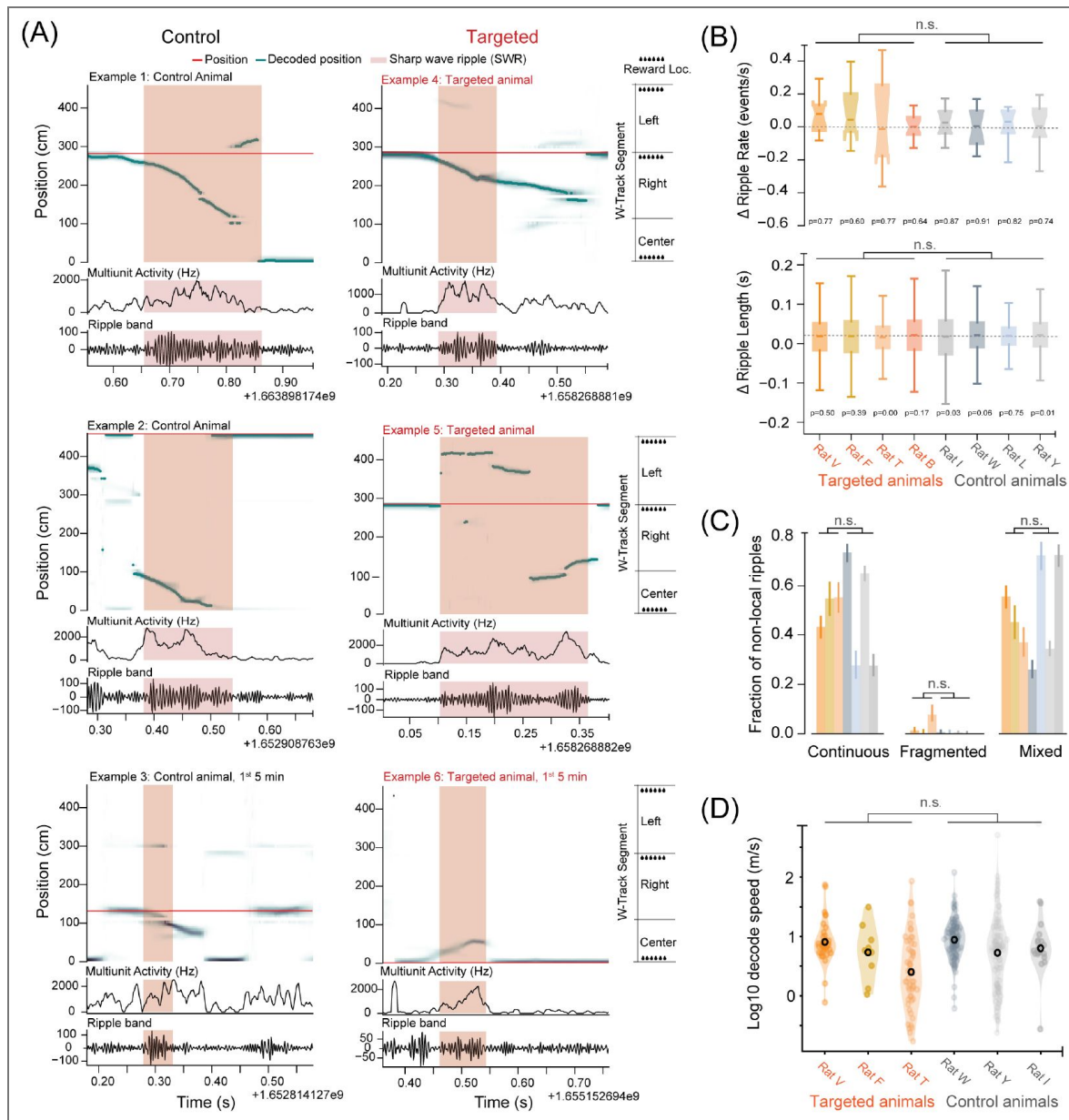


Figure 5. Theta disruption does not impact sharp-wave ripples (SWRs) or replay.

A, Examples of decoding position from population spiking in control (left) and targeted (right) animals during SWR events. Bottom row shows events occurring during the first five minutes of experience on the track (a *stimulation-on* period). Each panel: Top: actual linearized position (red) and estimated position posterior (greyscale). Middle: multiunit spike rate (spikes/s). Bottom: bandpass-filtered ripple events. SWR events highlighted in light pink. **B**, SWR rates (top) and length (bottom) do not differ significantly between *stimulation-on* and *stimulation-off* intervals in targeted (shades of orange) and control (shades of grey) animals (paired *t*-test for each animal, *p* values reported in the figure; comparison of groups not significant, mixed effects linear model $p = 0.929$ for rates, $p = 0.426$ for lengths). **C**, Fractions of non-local SWRs that are spatially continuous, fragmented, or a mix of continuous and fragmented states, in targeted (orange) and control (grey) animals. Note overlapping distributions for each ripple content category. **D**, Average movement speed of the peak of the posterior distribution for continuous decodes (duration > 50 ms, probability of the event being continuous > 90)%. Distributions of speeds were very similar across control and targeted animals (mixed effects linear model, $p = 0.253$). Note, Rat B had a 32Ch linear probe and hence could not be used for SWR-associated content analysis. Rat L did not have enough continuous replay qualifying the inclusion criteria, but relaxing the inclusion criteria yielded similar results. We noted that there were some very high speed values. A visual inspection of those events suggested that they arise from cases where the posterior distribution was bimodal, and the peak of the distribution transitioned from one mode to the other over a single timestep, thus suggesting high speed movement even though each mode of the distribution was evolving much more slowly.

The importance of precise temporal organization of spiking during theta for learning is consistent with the hypothesis that this organization facilitates synaptic plasticity (Skaggs et al. 1996 [↗](#); Lisman and Jensen 2013 [↗](#); Buzsáki 2002 [↗](#)). Our results provide strong support for this hypothesis: disruption of the precise sequential activity during theta was sufficient to profoundly impair learning on outbound trials while having no detectable effect on average place field properties or on learning on the interleaved inbound trials.

The dissociation between outbound and inbound learning mirrors patterns seen with awake SWR disruption (Jadhav et al. 2012 [↗](#)) and indicates that disrupting either pattern is sufficient to lead to a learning deficit, although theta disruption appears to generate a more profound effect. The selectivity to outbound trials also offers clues as to the computations that may be disrupted. Outbound trials require that animals remember the outer arm they came from on the previous inbound trial such that they can use that information to choose to go to the opposite outer arm. This requires associations across locations, and correctly timed theta sequences may be critical for learning and retrieving these associations. Specifically, Hasselmo et al. (Hasselmo 2025 [↗](#)) proposed that early phases of theta support information encoding, while later phases support retrieval (Hasselmo, Bodelón, and Wyble 2002 [↗](#); Hasselmo 2025 [↗](#); Mizuseki et al. 2009 [↗](#)). Consistent with this hypothesis, hippocampal theta sequences typically represent locations close to the animal during early phases of theta and can represent hypotheticals, including possible futures and alternative pasts, during later phases (Comrie, Frank, and Kay 2022 [↗](#)). The possibility that the timing of these representations is critical for normal brain function is consistent with our observation that representations of locations more distant from the animal were preserved during theta disruption, but the timing of these representations was no longer regular. Thus, the reliability and rhythmicity of septal input to the hippocampal formation are likely critical for providing temporal windows in which relevant behavior-related patterns can be organized into learning and action for outbound trials.

By contrast, a simple location-action association is sufficient for inbound trials, where animals could learn to execute a left or right turn when coming from the right or left outer arms, respectively. Thus, preserved place field activity in targeted animals could enable normal inbound learning.

Critically, the behavioral effects in targeted animals were seen even though stimulation was off during the middle third of each exposure to the W-track. Consistent with this behavioral result, sequential firing during locomotion (at both the pairwise and population level) was disrupted during *stimulation-on* periods and remained disrupted in *stimulation-off* periods. This surprising result indicates that the disruption of theta sequences during the initial experience in an environment is sufficient to have lasting effects, potentially as a result of the rapid plasticity engaged during early learning.

Understanding precisely why this temporal organization is critical will require more distributed measurements. Notably, MS targets include multiple cortical and subcortical targets (Joshi 2017 [↗](#)), and our manipulation may have disrupted precise spike timing throughout these regions. Additionally, monitoring regions beyond the temporal cortex would be informative given the broad coordination between hippocampal theta and other systems (Joshi et al. 2023 [↗](#); Eichenbaum 2017 [↗](#); Buño and Velluti 1977 [↗](#); Berg, Whitmer, and Kleinfeld 2006 [↗](#); Ledberg and Robbe 2011 [↗](#)).

Our results also demonstrate that theta sequences and replay sequences are dissociable. While our manipulation disrupted theta sequences on the track, we saw no effect on replay during periods of immobility on the track. Instead, we observed very similar patterns of activation of sequential representations during SWR events in targeted and control animals. This result contrasts with the conclusion of a previous paper (Liu et al. 2023 [↗](#)) that showed the disruption of hippocampal theta sequences and subsequent replay during rest using an optogenetic manipulation of the entorhinal cortex (EC). While our study and the previous study both employed disruptions limited to locomotion, one possible explanation for the different outcomes is the site of the disruption (MS vs. EC). Previous work has shown that the EC can influence replay (Yamamoto and Tonegawa 2017 [↗](#)), and thus, the EC manipulation may have additionally impacted EC neural activity, which

in turn may have impacted subsequent replay. That said, both our MS manipulation and the previous EC manipulation could have an impact in sites other than hippocampal area CA1 where spatial firing, theta sequences, and replay were assessed. Nevertheless, our results show that disrupting theta sequences is not sufficient to disrupt hippocampal replay, and that replay alone is not sufficient to enable learning. Replay was intact while neural dynamics during locomotion were impaired, and this manipulation was accompanied by a critical impairment in learning the more cognitively challenging outbound component of the W-track.

Our findings, and previous dissociations between precise timescale and place field properties (Petersen and Buzsáki 2020 [↗](#); Liu et al. 2023 [↗](#); Wang, Roth, and Pastalkova 2016 [↗](#)) further suggest that different circuits with different time constants are responsible for processing spatial and temporal information in the hippocampal circuit. Spatial/contextual information may arrive from regions with slower timescales (such as the cortex), making them less susceptible to sub-second brief disruptions, while precisely timed inputs from the medial septum coordinate the tightly controlled timing offsets between hippocampal neurons. We hypothesize that learning requires the intersection of these two streams of information in the hippocampal network, and is impaired by the disorganization of the precise temporal templates in which internal plans can be matched to external inputs.

Gaining a deeper understanding of the roles of these inputs will depend on manipulations that target specific patterns of organization of neural activity. This work will build on a wide array of previous studies that developed approaches to disrupt or alter pattern-specific firing (e.g., in medial septum (Wang et al. 2015 [↗](#); Fuhrmann et al. 2015 [↗](#); J. Robinson et al. 2016 [↗](#); Zutshi et al. 2018 [↗](#); Kloc et al. 2020 [↗](#); Mouchati et al. 2020 [↗](#); Quirk et al. 2021 [↗](#); Etter et al. 2023 [↗](#); Gemzik, Donahue, and Griffin 2021 [↗](#)), entorhinal cortex (Liu et al. 2023 [↗](#); Lepperød et al. 2021 [↗](#)), nucleus reuniens (Ito et al. 2015 [↗](#)), supramammillary nucleus (Farrell et al. 2021 [↗](#)), and pedunclopontine nucleus (Kaur et al. 2025 [↗](#))). We suggest that greater incorporation of open and closed-loop feedback paradigms will enable a better dissection of the roles of specific patterns of input and information flow in these circuits.

In summary, our results establish a dissociation between locomotion-related theta sequences and offline replay during learning, and indicate that disrupting neural activity patterns during locomotion is sufficient to drive a profound and specific learning deficit. This points toward a critical role of theta-sequences in building and accessing representations that enable choices that depend on integrating past and potential future locations.

Materials and Methods

Experimental model and animals

Neural activity (cellular firing and local field potential) was recorded from the CA1 region of the dorsal hippocampus while simultaneously performing optogenetic stimulation of parvalbumin (PV) neurons in the medial septum in ten PV-Cre Long Evans male rats (6–10 months old, 500–650 g) (Yu et al. 2018 [↗](#)). All experimental procedures were in accordance with the University of California San Francisco Institutional Animal Care and Use Committee and US National Institutes of Health guidelines.

Surgery, virus injection, array insertion, and electrophysiological recordings

Prior to surgical procedures, animals were maintained on full food (ad libitum) for one week. Surgery involved injection of a Cre-dependent virus (AAV5-Ef1a-DIO-hChR2(H134R)-eYFP, UNC Vector Core) into the medial septum (ML −0.15 mm, AP +0.904 mm, DV −6.94 mm) and implantation of a multi-electrode recording array in hippocampal area CA1 (AP −3.8 mm, ML +2.8 mm, DV set by tetrode adjustment). Virus (450 nL, 4×10^{12} viral particles/mL) was injected at $0.05 \mu\text{L min}^{-1}$ using a Hamilton Nanofil 10 μL syringe with a beveled 33G needle, mounted on a stereotaxic syringe pump (KD Scientific). After each injection, the needle was left in place for 5–10 minutes before

retraction to ensure diffusion. Next, we inserted a tapered optic fiber (Optogenix, Lambda-B, NA = 0.39; core/cladding diameters = 200 μm /225 μm ; emitting length = 2 mm; implant length = 6 mm; connector: 2.5 mm ceramic ferrule) following the same path as the injection needle. The optic fiber was inserted 200 μm deeper than the injection location to maximize lateral spread of the laser beam in the injection zone. After virus injection, rats were implanted either with 32-channel silicon probes ($n = 3$; NeuroNexus, 1 shank, 32 electrode sites, shank length = 6 mm, spacing = 50 μm , electrode surface area 177 μm^2) or with a 128-channel tetrode drive (v4.6). Animals were allowed 2–3 weeks for viral expression before experiments began. For the animals implanted with 32-channel silicon probes ($n = 3$), we tested the laser power and pulse durations required to entrain theta oscillations. All three animals were targeted as expected; however, for one animal, the maximum laser power tested was 50 mW, which was not sufficient to induce theta oscillatory activity during the run (only during rest periods). Hence, that animal was not included in the rhythmic stimulation analysis.

Behavioural arenas

Rats were pre-trained on a linear track (125 cm long) to alternate between reward wells on either end. Prior to training, animals were food restricted to 85% of their starting body weight. Nose pokes at the wells were detected by an infrared beam, and upon a successful visit 200 μL of sweetened milk was delivered. Pre-training included two 30 min sessions interleaved with an hour of rest in the rest box.

Linear track

Animals implanted with the recording drives/probes were made to run on the linear track for sweetened milk reward while the electrophysiological signals were recorded using SpikeGadgets hardware and software (v.1.8.0). The animals implanted with the 128 Ch tetrode drives had to undergo a series of tetrode adjusting steps over 10–14 days to lower the tetrodes to the hippocampal cell layer before we started behavioral protocols. The recording setup involved multiple independently moving pulley systems running on carbon fiber tracks hanging from the ceiling, which supported the cables running from the animal's head allowing the animals to move freely on the 2-D linear track plane. The connection from the drive was passed on through a rotating commutator before connecting to the computer allowing the animals to freely rotate their heads without putting rotational strain on the cables. The medial septal parvalbumin neurons were stimulated using an optic fiber from the laser connected to the optic fiber ferrule on the head of the animal. The animals underwent a stimulation with spatial constraint where the laser was ON only when the animals were away from the reward wells, thereby stimulating primarily during running times. The stimulation protocol (described below) involved pulses of varying width (pulse width values), intervals (interval values) and numbers (number of pulses).

W-Track

After linear track running sessions, animals ran on a W-shaped track to test spatial learning and memory-guided decision making. The track involved 3 arms with the sidearms connected to the base of the center arm forming a W-shape. The rule for this task was that the animals were rewarded when they alternate between the two side arms of the track while visiting the center arm in between each alternation. 0.2 mL sweetened milk was administered at the end of each arm for every correct visit. Electrophysiological data from dCA1 was recorded while the animals underwent a closed loop stimulation of the medial septal parvalbumin neurons. The closed loop stimulation involved both spatial constraint where the laser was ON only when the animals were away from the reward wells and theta phase constraint where the stimulation was tuned to the ascending phase of theta (90 degrees).

Optogenetic manipulation setup

Optogenetic stimulation was achieved through an Omicron LuxX Plus laser (488 nm, 100 mW maximum output) connected to implanted optic fiber cannulae. The cannulae were purchased from Optogenix (Lambda-B, numerical aperture: 0.39, core/cladding diameter: 200 μm /225 μm ,

emitting length: 2 mm, implant length: 6 mm, ceramic ferrule connector: 2.5 mm). Laser power was initially titrated, beginning at 5 mW and increasing gradually until reaching 77 mW, which was used for all subsequent experiments.

In the probe-only animals ($n = 3$), we tested the ability to entrain hippocampal LFP during the interleaving rest box periods, by varying laser power (from 5 mW to 77 mW at source) and pulse width (1–60 ms). We found that with a pulse width of 1–5 ms at laser powers greater than 25 mW, we could not completely entrain hippocampal LFP in the rest box (when the animals are predominantly immobile), while higher laser powers (77 mW) and longer pulse widths (> 20 ms) were required for entrainment on the linear track (when the animals are engaged in the movement) (Supplementary Figure 5 [↗](#)). Thus, for the rhythmic and phase-specific manipulation, we used 77 mW laser power at source and pulse widths greater than 20 ms. Note that while this is a high overall light power, the use of a tapered fiber substantially lowers the power density in the tissue as compared to a blunt fiber.

Optogenetic stimulation protocols

Optogenetic stimulation protocols were delivered using a custom adaptation of the FSGui module of SpikeGadgets Trodes software. Publicly available code can be found at https://bitbucket.org/mkarlsson/trodes/branch/fsgui_theta2 [↗](#). This module enables two relevant modes of stimulation: rhythmic stimulation at a certain frequency, and theta-phase specific stimulation at the requested phase of the recorded theta rhythm. To limit stimulation to behavioral periods associated with locomotion and the associated hippocampal theta rhythm, we restricted use of these stimulation modes to periods in which the animal occupied a user-defined spatial area of the maze. Spatial filters were set to only permit stimulation when animals were locomoting on the maze, and to prevent stimulation when animals occupied reward port locations.

Open loop stimulation was delivered using the following parameters: Spatial lockout period in samples (set to 7500); Pulses per pulse (set to one for data presented in this paper); Period (ms): 50, 80, 100, 125, 165, 250; Pulse length (ms): 1, 5, 27, 40 (we resorted to using > 20 ms as it provided us with the best entrainment response as compared to 1 and 5 ms).

Closed loop theta phase-specific stimulation parameters enabled specification of a target theta phase. Briefly, one channel of 30 kHz of LFP data recorded from corpus callosum was downsampled to 1500 Hz. A second-order Butterworth filter with a short group delay was applied to the LFP data in the 4–12 Hz frequency band (similar to (Siegle and Wilson 2014 [↗](#))). Upward- and downward-going zero-crossings were detected in the filtered data, and were used to estimate the period of the prior cycle. The period of the prior cycle and zero crossing times were then used to estimate the sample at which the incoming data would reach the target theta phase. A lockout period was specified to prevent excess stimulus triggers. This enabled stimulation at the target phase (Sup. Figure 3 [↗](#)). The closed loop stimulation parameters used in these experiments were: Target theta phase (90 degrees); Spatial lockout period in samples (7500); Theta lockout period (2400 samples); Pulses per pulse train (1); Pulse length (ms): either 27 ($n = 3$ probe animals) or 40 ($n = 5$ animals).

Behavior analysis

A state space algorithm (Smith et al. 2004 [↗](#)) was used to estimate the probability of a correct choice and accompanying 90% confidence interval on inbound, outbound, or all trials. The “learning trial” was defined as the first trial on which the lower bound of the 90% confidence interval was above chance (50%) (Smith et al. 2004 [↗](#)). For each animal, we compute related behavioral metrics: the learning trial, the slope of the regression of performance on evenly spaced numbers from 0 to 1 equal to the total number of trials, and the number of trials performed correctly. We also computed average learning curves across animals within each group, and for each trial type, by finding the mean fraction of trials performed correctly in consecutive spans of 20 trials. We show these averages and accompanying 95% confidence intervals.

Data analysis

Codebase and statistical approach

We prepared this dataset in a commonly accessible and reusable format, meeting the Neurodata Without Borders (NWB) standard (Rübel et al., 2023). This data was then ingested into the spyglass framework (Lee KY, Denovellis EL, et al., 2024), which allowed for centralized data management, ensuring all intermediary steps from the raw NWB file to various analysis files were logged. Complete code for data processing and figure generation is available at: https://github.com/LorenFrankLab/ms_stim_analysis.

Across all experiments, an epoch is defined as a single period of time in the task arena, and is typically ~20 min. At most, one setting of the optogenetic stimulation protocol was run per epoch. Within each epoch, the ~5–7 min “*stimulation-off*” period during the middle of the epoch served as a control for within-epoch comparison of the effect of the driving stimulus. For most analysis, we restrict the data to the trial intervals, defined as the time between leaving a reward port to entering the next. Reward port occupancy was determined as the tracked head position being within 20 cm of the end of the track. For analysis of features specific to animal movement, these intervals were further restricted to times where the animal head speed exceeded 10 cm s⁻¹.

Bootstrap confidence intervals were performed by resampling the observed distribution of events 1000 times and calculating the summary statistics for each. For hierarchical bootstrapping, both the animals within treatment groups and the events within animals were resampled for each calculation. *t*-test statistics were run using the scipy implementation. Mixed linear regression models were run using the statsmodels python package.

LFP analysis

LFP data was generated from the original 30 kHz electrophysiology recordings by filtering the data with a 0–400 Hz low-pass filter and downsampling to 1 kHz. Results were generated by analyzing the data from a reference electrode above the CA1 layer. Artifacts in the data were defined as periods where the amplitude of the filtered data exceeded 2 mV and were excluded from analysis.

LFP power spectrums were calculated from intervals when animals were outside the reward ports and traveling with a speed of at least 10 cm s⁻¹. Power density spectrums and related scores described below were calculated for each continuous interval meeting these criteria using the scipy implementation of Welch’s method. These were then weighted by duration of the interval and averaged to produce the results shown. [Full analysis: func:

```
Analysis/lfp_analysis/opto_spectrum_analysis()
```

Entrainment score

Entrainment scores were calculated for open-loop fixed-frequency stimulations based on the power of the LFP spectrum at the target frequency during stimulus on (p_{test}) and stimulus off (p_{control}) intervals. [Full analysis func:

```
Analysis/lfp_analysis/calculate_entrainment_score().]
```

$$E = \frac{p_{\text{test}} - p_{\text{control}}}{p_{\text{test}} + p_{\text{control}}}.$$

Suppression score

Suppression scores were calculated by first identifying the frequency with maximum power in the theta band ($4 < f < 12$ Hz) in the control power spectrum, then measuring power at that frequency in the control ($p_{\text{peak_control}}$) and test ($p_{\text{peak_test}}$) intervals. [Full analysis func:

```
Analysis/lfp_analysis/calculate_suppression_score().]
```

$$S = \frac{p_{\text{peak_control}} - p_{\text{peak_test}}}{p_{\text{peak_control}} + p_{\text{peak_test}}}.$$

Spike Sorting

Spike sorting was performed in the `spyglass.spikesorting.v0` pipeline. Raw recording files were referenced and bandpass filtered from 600–6000 Hz. Artifacts in the recording were identified as periods where at least 75% of the filtered channel recordings exceeded 2000 μV with 500 ms padding on each side of the event. These were excluded from sorting. Spike sorting was run on each epoch through the `spikeinterface` package using `mountainsort4` sorter with; signal whitening, `adjacency_radius` 100, `clip_size` 30, `detect_threshold` 3, and `detect_interval` 10. After sorting we computed quality metrics on the identified units using `spikeinterface`. We then excluded units with a nearest neighbors noise overlap greater than 0.03, a peak offset greater than 2, or an inter-spike interval violation greater than 0.0025.

Event-triggered firing rates for individual cells

For each sorted unit, spike events were binned based on time delay to the nearest stimulus onset. We then calculated the KL divergence between this distribution of spike delays and a null uniform distribution in the same interval. To determine if this represented a significant divergence from the null distribution, we performed circular shuffling of the stimulus times by offsetting each stimulus onset time by an independently chosen value from a range of $\pm$ the stimulus period. The KL divergence between the shuffled delay distribution and the uniform distribution was calculated. This process was repeated 1000 times to form a null distribution of KL divergences for each unit. A unit was determined to be significantly modulated if the observed KL divergence was above the 99.5 percentile of the null KL distribution.

Event triggered MUA analysis

Multiunit firing rates were calculated by summing the total number of firing events from sorted units within 2 ms time windows, and then smoothing with a gaussian filter with standard deviation of 5 ms. The mean of this value at each timepoint across trials was estimated using bootstrap sampling techniques described above.

Place fields

Place fields were calculated by fitting a `SortedSpikesDetector` decoding model from the `non_local_detector` package ([GitHub](#))¹. Positions were first projected to the linear track skeleton to allow for 1D decoding. Decoding was performed using the packaged sorted spikes kernel density estimation algorithm with sampling rate of 500 Hz, position standard deviation of 6 cm, and discrete position binning of 2 cm. For comparisons of the place coding of cells between the optogenetic test and control intervals, we fit separate models using each of these segments as the encoding interval.

Unit place field coverage

For much of our analysis, we wanted to restrict to units which displayed spatially-localized distributions of firing activity. To do so, we calculated the highest posterior density of the place field distribution to determine the minimum spatial area required to capture 50% of this firing distribution. Thresholding an upper limit for this value allowed us to select cells with well-localized firing patterns.

Temporal offset of cross-correlation

Cross correlograms were calculated for intervals when the animal was continuously running outside of the reward port (speed $> 10 \text{ cm s}^{-1}$ for at least 500 ms). Histograms for all spike pairs were calculated over 1 ms time bins.

To calculate fast, theta-timescale delay times, the cross correlograms were then smoothed with a gaussian kernel with standard deviation $\sigma = 3 \text{ ms}$. Cells pairs were included for further analyses if they had at least 50 pairs of spike events co-occurring within a $\pm 100 \text{ ms}$ window. To define the delay of peak cross-correlation times, we used the `scipy` function `find_peaks` with a distance equivalent to 80 ms and selected the delay peak with the largest magnitude.

For slow, positional timescale delay times, the cross correlograms were smoothed with a gaussian kernel with standard deviation $\sigma = 500$ ms. Cell pairs were included if they had at least 100 pairs of spike events co-occurring within a ± 1000 ms window. Peaks were identified with a distance equivalent of 2 s, and the peak closest to zero taken as the delay time.

Place Field Similarity

Consistency of place field coding under the stimulus protocol was quantified by the KL divergence between the place distribution of each neuron's firing during the control versus the test interval. Units were included in the KL-based place-field stability analyses only if they satisfied both of the following: **(i)** Spike-count criterion: at least 100 spikes in each interval type used for the comparison—*stimulation-off*, *stimulation-on*, and the *stimulus* sub-interval. **(ii)** Coverage criterion: the 50% highest posterior density (HPD) region of the unit's spatial firing distribution was contained within 40 cm on the linear track or within 100 cm on the W-track.

Outbound repeat probability

Outbound repeat sequences are defined as the number of visits to the same outer arm without visiting the alternate arm within an epoch. Note that these sequences are typically interleaved with rewarded visits to the center arm. We show the distribution of sequences of length greater than x in Figure 3E. This distribution under random choice of outer arm at each trial is given as

$$P(\text{sequence} > x) = 1 - \sum_{i=1}^x \left(\frac{1}{2}\right)^i.$$

We sampled this random binomial choice process for the observed number of trials 1000 times for each animal group to get the 95% confidence interval of a random choice process in each case. Values for long repeat sequences below this interval indicate learning of the outbound task.

Clusterless Decoding

Clusterless waveform features were extracted from the same filtered recording used for spike sorting. Event times were identified using `spikeinterface.DetectPeakByChannel`, with an amplitude threshold of 100 μV and a local exclusion radius of 100 μm . Waveforms of each event were featurized by the peak amplitude within ± 500 μs of the event on each channel of the tetrode on which the event was detected. Clusterless position decoding was performed using the `ClusterlessDetector` algorithm from the `non_local_detector` package (GitHub). Positions were first projected to the linear track skeleton to enable 1D decoding. Decoding was then performed with the package's clusterless kernel density estimation algorithm, using a sampling rate of 500 Hz, waveform feature standard deviation of 24 μV , position standard deviation of 6 cm, and discrete position binning of 2 cm.

As some animals had fewer detectable cells on later days of experiments, we restricted our subsequent decoding analysis to positions on the track with sufficient place cell coverage for reliable decoding. We defined this as track regions that fell in the 50% credible interval of at least two well-localized cells' firing fields (95% of the units place field distribution localized to < 50 cm of the track, see Unit place field coverage). From this we defined time intervals where the animal was physically located in these positions to use in decoding analysis below.

Ahead-Behind Distance

Orientation of the animals during movement was estimated using the arc tangent of the animal's velocity. This value is reasonably well-defined during run intervals analyzed here. Distances between the position and decode along the track path are calculated using the `get_ahead_behind_distance` function in the `non_local_detector` package.

Decoding Stimulus Response

Analysis was restricted to stimuli that occurred while the animal was running outside of the reward ports (speed > 10 cm s^{-1}). Furthermore we excluded stimulus times with poor decoding (more than 10 ms of the ± 50 ms window surrounding a stimulus containing decodes > 50 cm from the animal's position).

Decode Coverage

Decode coverage is calculated as the area of the linearized track (in cm) spanned by the 95% credible interval of the position posterior from the decoding results.

Posterior entropy

Entropy of the posterior distribution over discrete position bins is defined as:

$$S = - \sum_x p(x) \log [p(x)].$$

Ahead-behind distance

see above. Values are clipped to a range of 20 cm around the animal to prevent infrequent large events from skewing distribution.

Multi Unit Activity

Rate of spike events from all sorted neurons is calculated in 500 μ s time bins and smoothed with a gaussian kernel of $\sigma = 1$ ms.

Decoding Distance Power Spectrum

Intervals were identified where the animal was continuously running ($>10 \text{ cm s}^{-1}$) for at least 1 s outside of the reward port. Power spectrums were calculated for each of these intervals using the scipy implementation of Welch's method with a 1 s window. The mean was taken as an average weighted by the length of each interval.

Decoding Distance Power Spectrum Theta Peak

For each spectrum calculated above, the average power in the theta range (6–10 Hz) was normalized by the average power in a reference frequency range (union of 2–6 Hz and 10–14 Hz). The theta peak of the spectrum was defined as the log of this ratio.

Decoding Ahead/Behind Maximum Distance

Intervals were identified where the animal was within a 20 cm radius of the choice point center. For each stimulus within these intervals we identified the maximum decode to animal distance with ± 65 ms of stimulus onset.

Sharp Wave Ripples

Sharp wave ripples were identified using the `spyglass.ripple.v1` pipeline. LFP data was first bandpass filtered in the range 150–250 Hz. Ripple times were then identified using the z-score threshold method (Kay et al. 2016 [DOI](#)) implemented in `ripple_detection` (https://github.com/Eden-Kramer-Lab/ripple_detection [DOI](#)). The algorithm was run with a speed threshold of 4 cm s^{-1} , z-score threshold of 2, smoothing $\sigma = 4$ ms, and minimum ripple duration of 15 ms.

Ripple rates were calculated on a per-epoch basis as the number of ripple events during the optogenetic test and control intervals divided by the duration of stationary times in the corresponding interval.

Sharp Wave Ripple Content

As sharp wave ripples frequently decode to non-local positions, we cannot simply filter times when the animal's physical location is in a region of the track with sufficient place field coverage. For this analysis, we exclude epochs where at least half of the timepoints cannot be reliably decoded.

A ripple event was classified as non-local if the average decode to animal distance during the interval was greater than 20 cm. Each of these non-local ripples was further classified as continuous/fragmented if the decoding transition state probability was $>80\%$ weight toward the corresponding state for at least 80% of the ripple duration. It was otherwise labeled as a 'mixed' ripple event.

Supplementary Information

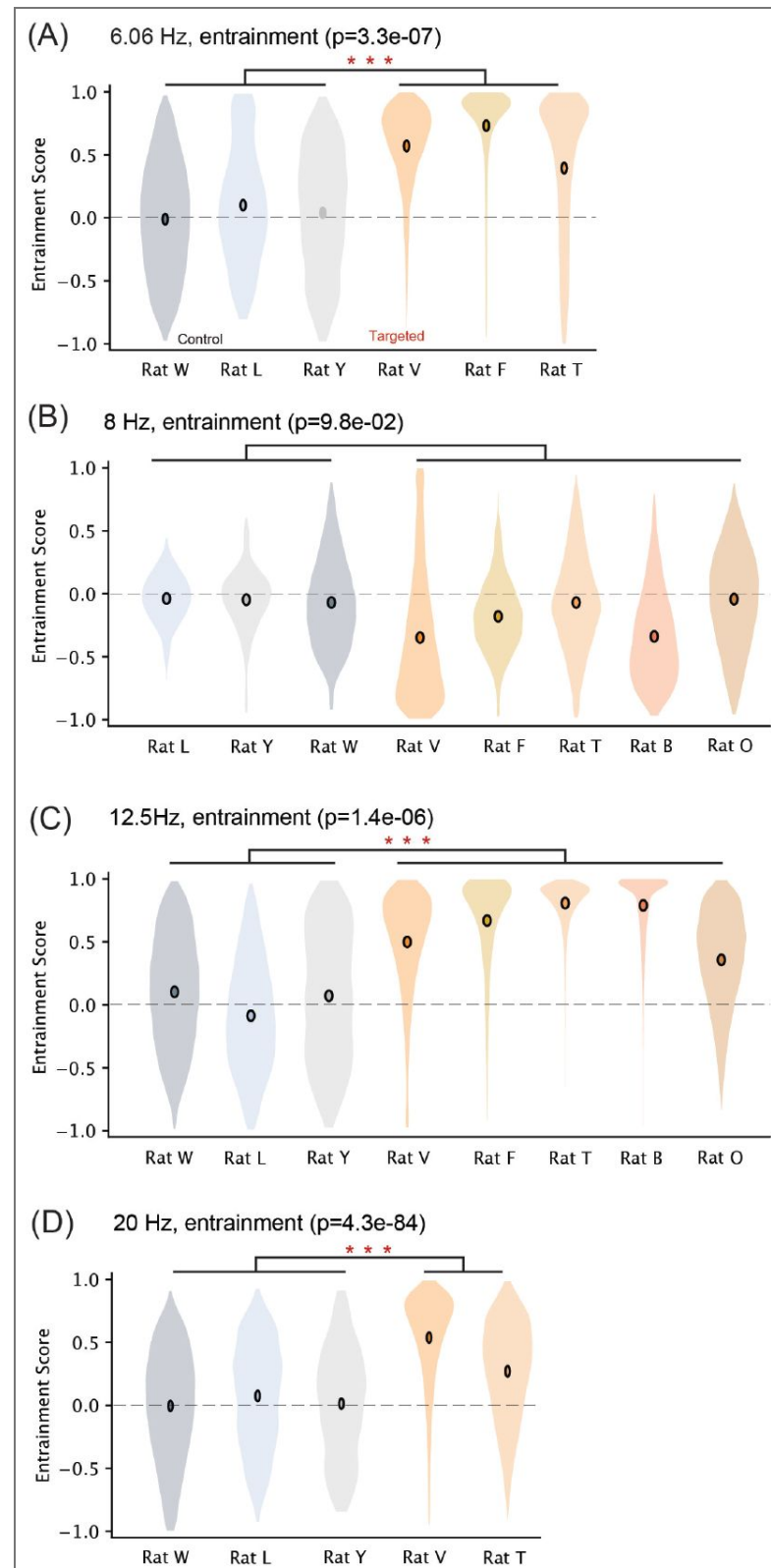


Figure S1. Entrainment of theta at different stimulation frequencies. **A**, Entrainment scores distributions across sessions for control ($n = 3$, grey) and targeted ($n = 3$, grey) animals at 6 Hz entrainment frequency. Targeted animals show significant entrainment compared to controls (mixed effects linear model (animal, targeted state), effect of targeting; $p = 3.3 \times 10^{-7}$, control $n = 17492$ interval pairs, targeted $n = 21040$ interval pairs). **B**, Entrainment

scores distributions across sessions for control ($n = 3$, grey) and targeted ($n = 3$, grey) animals at 6 Hz entrainment frequency. Targeted animals show significant entrainment compared to controls (mixed effects linear model (animal, targeted state), effect of targeting; $p = 3.3 \times 10^{-7}$, control $n = 17492$ interval pairs, targeted $n = 21040$ interval pairs). **B**, Entrainment scores distributions across sessions for control ($n = 3$, grey) and targeted ($n = 5$, orange) animals at 8 Hz entrainment frequency (control $n = 8012$ interval pairs, targeted $n = 34184$ interval pairs). **C**, Entrainment scores distributions across sessions for targeted ($n = 5$, orange) and control ($n = 3$, grey) animals at 12.5 Hz entrainment frequency. Targeted animals show significant entrainment compared to controls (mixed effects linear model (animal, targeted state), effect of targeting; $p = 1.4 \times 10^{-6}$, control $n = 20152$ interval pairs, targeted $n = 30655$ interval pairs). **D**, Entrainment scores distributions across sessions for control ($n = 3$, grey) and targeted ($n = 2$, orange) animals at 20 Hz entrainment frequency. Targeted animals show significant entrainment compared to controls (mixed effects linear model (animal, targeted state), effect of targeting; $p = 4.3 \times 10^{-84}$, control $n = 10315$ interval pairs, targeted $n = 12863$ interval pairs).

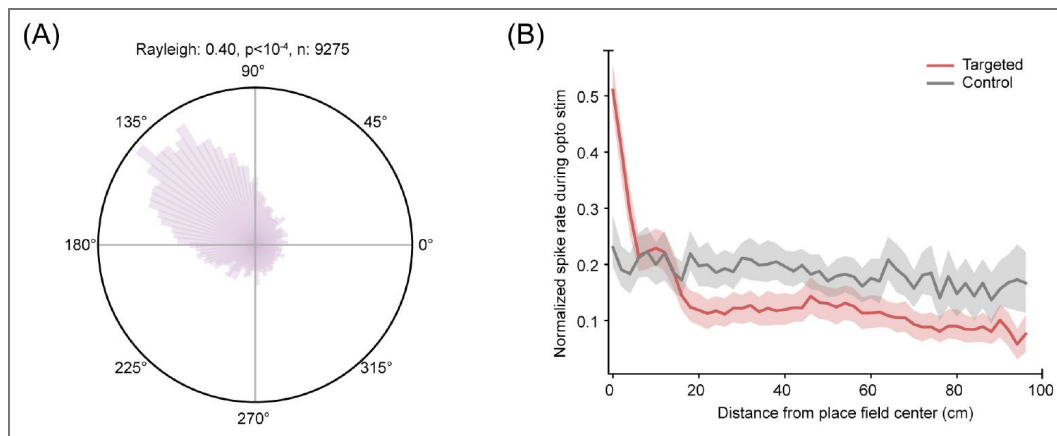


Figure S2. Accuracy and effects of theta phase-specific stimulation.

A, Stimulus-triggered phase histogram for all linear track targeted animals (mean vector length = 0.4; $p < 0.005$; target phase = 90°; pulse length > 20 ms; number of pulses = 9275). **B**, Normalized spike rate during optical stimulation in targeted (red, $n = 307$ neurons) and control (grey, $n = 295$ neurons) animals during theta phase-specific stimulation (theta phase = 90°). For each neuron, spikes were collected within each stimulus event and binned by distance of the animal from the neuron's place field peak defined from whole-epoch data. Shaded regions are the 95% CI with neuron resampling (see Methods). Note that extra spikes from stimulation are strongly localized to the place field center in targeted animals.

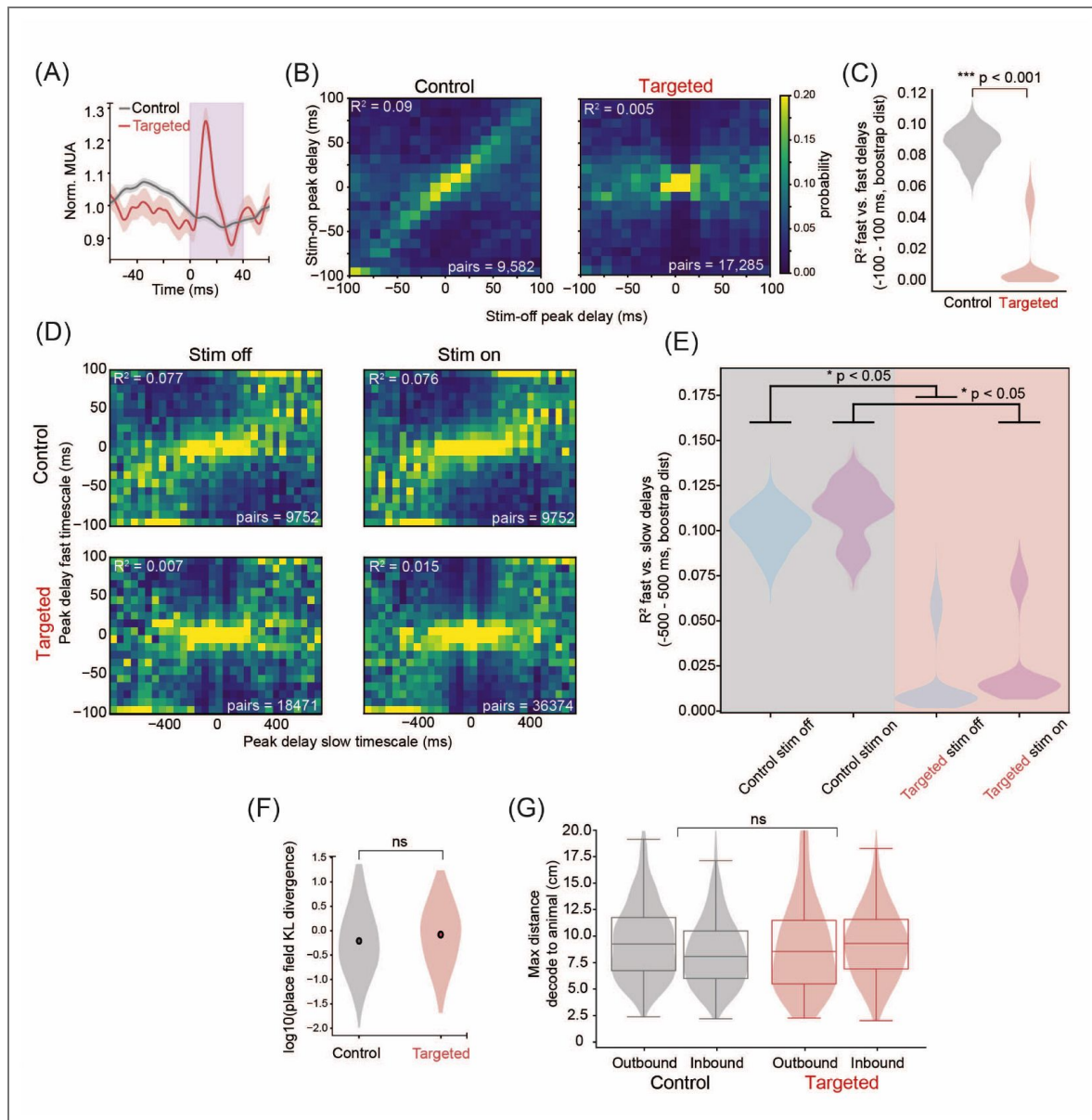


Figure S3. Disruption of short-timescale sequential spiking with preserved place fields in targeted animals.

A, Quantification of normalized multi-unit activity triggered by onset of stimulation pulses shows an increase in spiking ~10 ms after stimulation onset on the W-track (as in the linear track) in targeted animals. As expected, control animals show a decrease in spiking (stimulation is phase-specific to the ascending phase of theta). Shaded areas are the 95% CI of the mean normalized firing rates in each condition. **B**, Pairwise temporal offsets between cross-correlograms of putative pyramidal neurons computed in *stimulation-on* versus *stimulation-off* intervals in control ($n = 9582$ pairs, left) and targeted animals ($n = 17285$ pairs, right). Note, a strong diagonal indicates that the cross-correlation structure is maintained in control animals (as expected). This structure is much less apparent in transfected animals during both *stimulation-on* and *stimulation-off* periods. **C**, Hierarchical bootstrap comparison of R^2 values of linear fit, $p = 8.6 \times 10^{-4}$. **D**, Density plots of short- vs. long-timescale peaks for *stimulation-on* and *stimulation-off* periods for all cell pairs pooled across animals and run epochs. Top row: control animals; bottom row: targeted animals. The larger number of pairs for *stimulation-on* periods in targeted animals reflects a longer experimental period and stimulation driving larger numbers of short-timescale coincident firing events in cell pairs where too few events were detected for inclusion in the *stimulation-off* periods. **E**, Hierarchical bootstrap comparisons of R^2 values across conditions demonstrating significant differences during both *stimulation-on* and *stimulation-off* periods. **F**, Place field stability for all units across control and targeted animals, measured as the KL divergence of their spatial firing selectivity between *stimulation-on* and *stimulation-off* intervals (t-test of distributions of KL divergence between targeted and control animals, $p = 0.5$). **G**, Quantification of the spatial extent of non-local representations at the choice point (maximum distance of the posterior ± 65 ms around the stimulus) during inbound (IN) and outbound (OUT) task phases on the W-track for targeted (red) and control (grey) animals. Note that both targeted and control animals have a similar extent of non-local spatial representations.

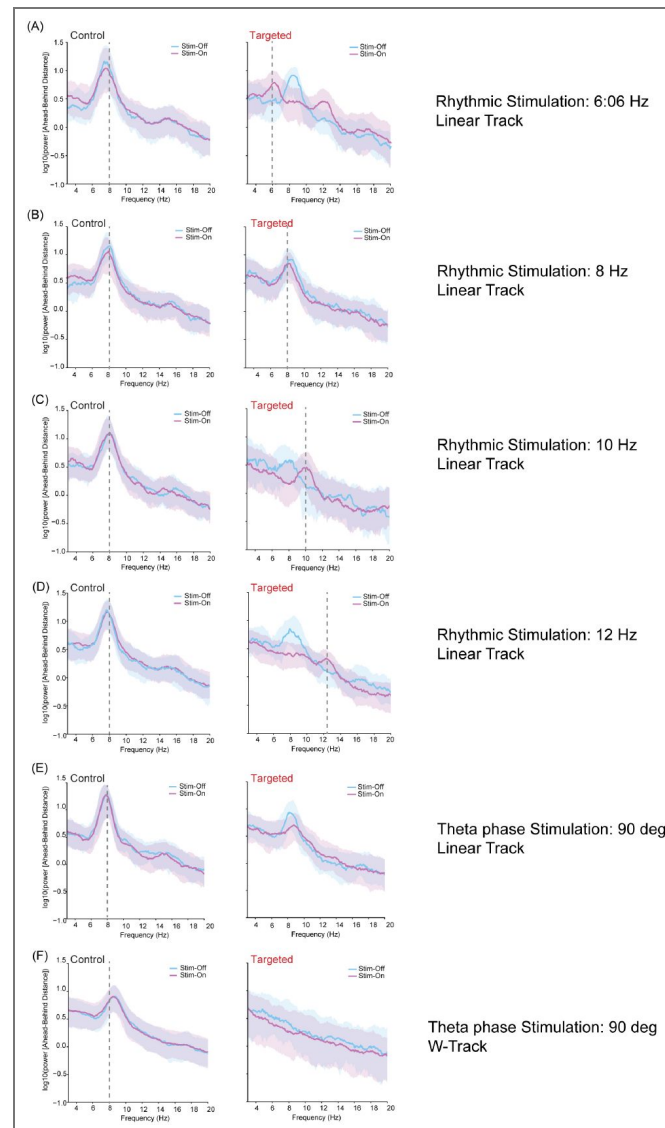


Figure S4. Effects of rhythmic vs. phase-specific stimulation on decoded representations.

A, Power spectrum of the decode-to-animal distance trace for control (left) and targeted (right) animals in *stimulation-on* (purple) versus *stimulation-off* (blue) trials during rhythmic stimulation condition on the linear track. Note that the posterior rhythmically represents current and non-local positions at the targeted frequency of 6 Hz. Control animals continue to represent positions at 8 Hz despite the stimulation. This rules out that light flickering has an impact on the hippocampal representation. **B**, Power spectrum of the decode-to-animal distance trace for control (left) and targeted (right) animals in *stimulation-on* (purple) versus *stimulation-off* (blue) trials during rhythmic stimulation condition on the linear track. Note that the posterior rhythmically represents current and non-local positions at the targeted frequency of 8 Hz. **C**, Power spectrum of the decode-to-animal distance trace for control (left) and targeted (right) animals in *stimulation-on* (purple) versus *stimulation-off* (blue) trials during rhythmic stimulation condition on the linear track. Note that the posterior rhythmically represents current and non-local positions at the targeted frequency of 10 Hz. **D**, Power spectrum of the decode-to-animal distance trace for control (left) and targeted (right) animals in *stimulation-on* (purple) versus *stimulation-off* (blue) trials during rhythmic stimulation condition on the linear track. Note that the posterior rhythmically represents current and non-local positions at the targeted frequency of 12.5 Hz. **E**, Power spectrum of the decode-to-animal distance trace for control (left) and targeted (right) animals in *stimulation-on* (purple) versus *stimulation-off* (blue) trials during theta-phase-specific stimulation targeted at the ascending phase of theta (90°) on the linear track. Note that the posterior rhythmicity is markedly reduced in targeted animals compared to control animals. **F**, Power spectrum of the decode-to-animal distance trace for control (left) and targeted (right) animals in *stimulation-on* (purple) versus *stimulation-off* (blue) trials during theta-phase-specific stimulation targeted at the ascending phase of theta (90°) on the W-track. Note that the posterior rhythmicity is markedly reduced compared to control animals. We also note that the posterior rhythmicity does not recover during control periods on the W-track.

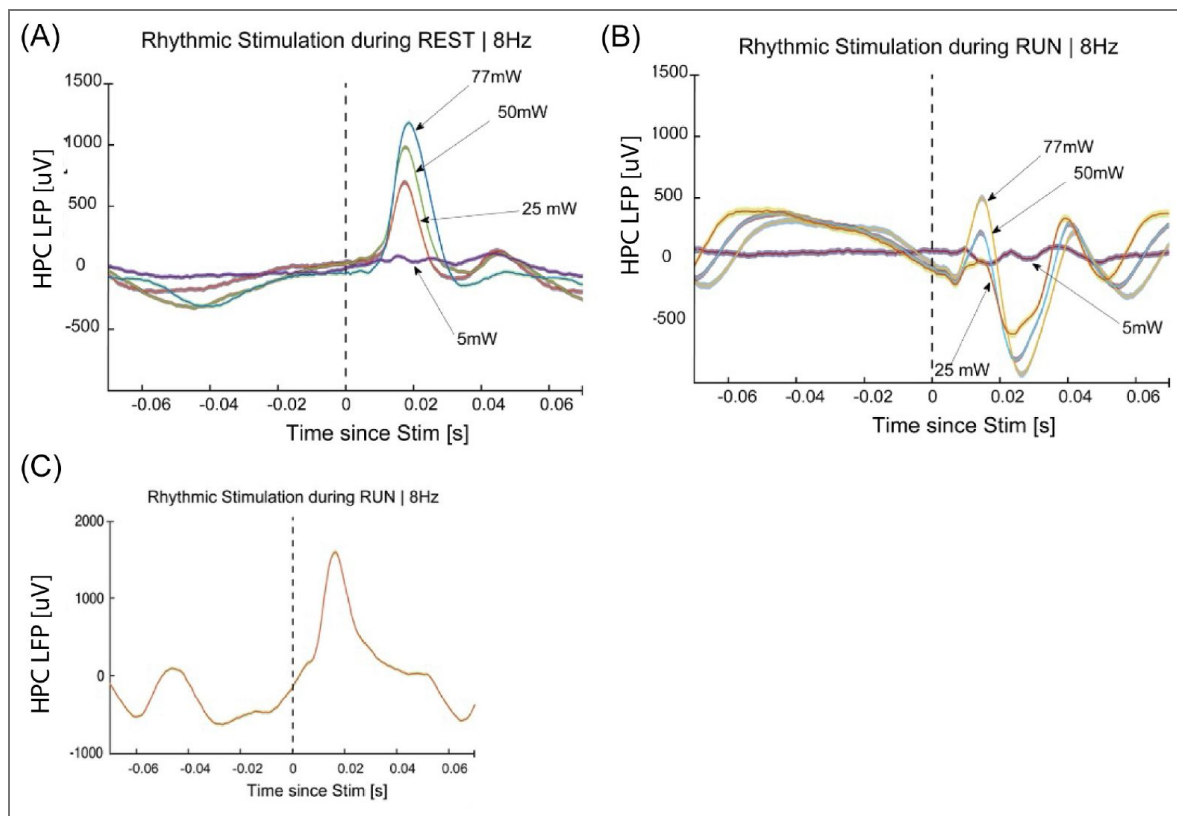


Figure S5. Laser-power dependence of LFP entrainment.

A, Stimulus-triggered LFP in one targeted animal shows a laser power-dependent response during REST periods at 5, 25, 50, and 77 mW laser powers and 1 ms pulse width. **B**, Stimulus-triggered LFP in one targeted animal shows a laser power-dependent response during RUN periods at 5, 25, 50, and 77 mW laser powers and 1 ms pulse width. **C**, Stimulus-triggered LFP in one targeted animal at 77 mW laser power and 40 ms pulse width shows complete entrainment during RUN periods.

Data availability

Data will be made publicly available via the Distributed Archives for Neurophysiology Data Integration (DANDI) archive upon publication, and a complete and fully reproducible pipeline for all analyses will also be shared. Code repositories developed in this work are cited throughout the corresponding methods sections for replicability. We prepared this dataset in a commonly accessible and reusable format, meeting the Neurodata Without Borders (NWB) standard (Rübel et al., 2023 [\[link\]](#)). In this process, we developed an NWB extension to store metadata for optogenetics experiments, such as virus, fiber, and stimulation protocol details. This extension is designed for public use and is available at <https://github.com/rly/ndx-optogenetics> [\[link\]](#). We built upon this extension in our lab's custom extension (<https://github.com/LorenFrankLab/ndx-franklab-novela> [\[link\]](#)) to capture additional metadata specific to our lab's stimulation protocols.

1. We have shared all of the code required to replicate the results and provided a figure panel replication key on the home page [code: https://github.com/LorenFrankLab/ms_stim_analysis [\[link\]](#)].
2. We will shortly post a link to all of the raw and intermediate data results, as well as a Docker container that permits full replication using Spyglass [<https://github.com/LorenFrankLab/spyglass> [\[link\]](#)].

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Conceptualization																
Methodology																
Investigation																
Software																
Formal analysis																
Resources																
Data Curation																
Writing - Original Draft																
Writing - Review & Editing																
Visualization																
Supervision																
Project administration																
Funding acquisition																

Funding

Funder	Grant reference number	Author
Simons Foundation	1189761	Abhilasha Joshi
Simons Foundation	1283731	Abhilasha Joshi
Simons Foundation	542981	Loren M Frank
Life Sciences Research Foundation		Abhilasha Joshi
NSF GRFP	1650113	Alison E Comrie
NIH	F31MH124366	Alison E Comrie
UCSF	Jonas Cohler Discovery Fellowship	Alison E Comrie
National Institute of Mental Health	F30MH126483	Jennifer A Guidera
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References

- Atherton Laura A.**, Dupret David, Mellor Jack R. (2015) Memory Trace Replay: The Shaping of Memory Consolidation by Neuromodulation. *Trends in Neurosciences* **38**:560-70
- Bender Franziska**, Gorbati Maria, Cadavieco Marta Carus, Denisova Natalia, Gao Xiaojie, Holman Constance, Korotkova Tatiana, Ponomarenko Alexey (2015) Theta Oscillations Regulate the Speed of Locomotion via a Hippocampus to Lateral Septum Pathway. *Nature Communications* **6**:8521
- Berg Rune W.**, Whitmer Diane, Kleinfeld David (2006) Exploratory Whisking by Rat Is Not Phase Locked to the Hippocampal Theta Rhythm. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience* **26**:6518-22
- Buño W.**, Velluti J. C. (1977) Relationships of Hippocampal Theta Cycles with Bar Pressing during Self-Stimulation. *Physiology & Behavior* **19**:615-21
- Burgess Neil**, Recce Michael, O'Keefe John (1994) A Model of Hippocampal Function. *Neural Networks: The Official Journal of the International Neural Network Society* **7**:1065-81
- Buzsáki G.** (1989) Two-Stage Model of Memory Trace Formation: A Role for 'Noisy' Brain States. *Neuroscience* **31**:551-70
- Buzsáki G.**, Leung L. W., Vanderwolf C. H. (1983) Cellular Bases of Hippocampal EEG in the Behaving Rat. *Brain Research* **287**:139-71
- Buzsáki György** (2002) Theta Oscillations in the Hippocampus. *Neuron* **33**:325-40
- Comrie Alison E.**, Frank Loren M., Kay Kenneth (2022) Imagination as a Fundamental Function of the Hippocampus. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences* **377**:20210336
- Comrie Alison E.**, Monroe Emily J., Kahn Ari E., Denovellis Eric L., Joshi Abhilasha, Guidera Jennifer A., Krausz Timothy A., Berke Joshua D., Daw Nathaniel D., Frank Loren M. (2024) Hippocampal Representations of Alternative Possibilities Are Flexibly Generated to Meet Cognitive Demands. *BioRxiv* <https://doi.org/10.1101/2024.09.23.613567>
- Davidson Thomas J.**, Kloosterman Fabian, Wilson Matthew A. (2009) Hippocampal Replay of Extended Experience. *Neuron* **63**:497-507
- Denovellis Eric L.**, Gillespie Anna K., Coulter Michael E., Sosa Marielena, Chung Jason E., Eden Uri T., Frank Loren M. (2021) Hippocampal Replay of Experience at Real-World Speeds. *eLife* **10** <https://doi.org/10.7554/eLife.64505>
- Diba Kamran**, Buzsáki György (2007) Forward and Reverse Hippocampal Place-Cell Sequences during Ripples. *Nature Neuroscience* **10**:1241-42
- Dragoi George**, Buzsáki György (2006) Temporal Encoding of Place Sequences by Hippocampal Cell Assemblies. *Neuron* **50**:145-57
- Ego-Stengel Valérie**, Wilson Matthew A. (2010) Disruption of Ripple-Associated Hippocampal Activity during Rest Impairs Spatial Learning in the Rat. *Hippocampus* **20**:1-10
- Eichenbaum Howard** (2017) Prefrontal-Hippocampal Interactions in Episodic Memory. *Nature Reviews. Neuroscience* **18**:547-58

- Etter Guillaume**, van der Veldt Suzanne, Choi Jisoo, Williams Sylvain (2023) Optogenetic Frequency Scrambling of Hippocampal Theta Oscillations Dissociates Working Memory Retrieval from Hippocampal Spatiotemporal Codes. *Nature Communications* **14**:410
- Farrell Jordan S.**, Lovett-Barron Matthew, Klein Peter M., Sparks Fraser T., Gschwind Tilo, Ortiz Anna L., Ahanonu Biafra, et al. (2021) Supramammillary Regulation of Locomotion and Hippocampal Activity. *Science* **374**:1492-96
- Fernández-Ruiz Antonio**, Oliva Azahara, de Oliveira Eliezyer Fermio, Rocha-Almeida Florbela, Tingley David, Buzsáki György (2019) Long-Duration Hippocampal Sharp Wave Ripples Improve Memory. *Science* **364**:1082-86
- Foster David J.**, Wilson Matthew A. (2007) Hippocampal Theta Sequences. *Hippocampus* **17**:1093-99
- Freund T. F.**, Antal M. (1988) GABA-Containing Neurons in the Septum Control Inhibitory Interneurons in the Hippocampus. *Nature* **336**:170-73
- Fuhrmann Falko**, Justus Daniel, Sosulina Liudmila, Kaneko Hiroshi, Beutel Tatjana, Friedrichs Detlef, Schoch Susanne, Schwarz Martin Karl, Fuhrmann Martin, Remy Stefan (2015) Locomotion, Theta Oscillations, and the Speed-Related Firing of Hippocampal Neurons Are Controlled by a Medial Septal Glutamatergic Circuit. *Neuron* **86**:1253-64
- Gemzik Zachary M.**, Donahue Margaret M., Griffin Amy L. (2021) Optogenetic Suppression of the Medial Septum Impairs Working Memory Maintenance. *Learning & Memory* **28**:361-70
- Gillespie AK**, Astudillo Maya DA, Denovellis EL, Liu DF, Kastner DB, Coulter ME, Roumis DK, Eden UT, Frank LM (2021) Hippocampal replay reflects specific past experiences rather than a plan for subsequent choice. *Neuron* **109**:3149-3163.e6. <https://doi.org/10.1016/j.neuron.2021.07.029>
- Gillespie Anna K.**, Maya Daniela Astudillo, Denovellis Eric L., Desse Sachi, Frank Loren M. (2024) Neurofeedback Training Can Modulate Task-Relevant Memory Replay Rate in Rats. *eLife* **12**:RP90944 <https://doi.org/10.7554/eLife.90944>
- Gupta Anoopum S.**, van der Meer Matthijs A. A., Touretzky David S., David Redish A. (2012) Segmentation of Spatial Experience by Hippocampal θ Sequences. *Nature Neuroscience* **15**:1032-39
- Hasselmo Michael E.** (2025) Development of the SPEAR Model: Separate Phases of Encoding and Retrieval Are Necessary for Storing Multiple Overlapping Associative Memories. *Hippocampus* **35**:e23676
- Hasselmo Michael E.**, Bodelón Clara, Wyble Bradley P. (2002) A Proposed Function for Hippocampal Theta Rhythm: Separate Phases of Encoding and Retrieval Enhance Reversal of Prior Learning. *Neural Computation* **14**:793-817
- Ito Hiroshi T.**, Zhang Sheng-Jia, Witter Menno P., Moser Edvard I., Moser May-Britt (2015) A Prefrontal-Thalamo-Hippocampal Circuit for Goal-Directed Spatial Navigation. *Nature* **522**:50-55
- Jadhav Shantanu P.**, Kemere Caleb, Walter German P., Frank Loren M. (2012) Awake Hippocampal Sharp-Wave Ripples Support Spatial Memory. *Science* **336**:1454-58
- Jensen O.**, Lisman J. E. (1996) Theta/Gamma Networks with Slow NMDA Channels Learn Sequences and Encode Episodic Memory: Role of NMDA Channels in Recall. *Learning & Memory* **3**:264-78
- Johnson Adam**, David Redish A. (2007) Neural Ensembles in CA3 Transiently Encode Paths Forward of the Animal at a Decision Point. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience* **27**:12176-89
- Jones G. A.**, Norris S. K., Henderson Z. (1999) Conduction Velocities and Membrane Properties of Different Classes of Rat Septohippocampal Neurons Recorded in Vitro. *The Journal of Physiology* **517**:867-77
- Joo Hannah R.**, Frank Loren M. (2018) The Hippocampal Sharp Wave-Ripple in Memory Retrieval for Immediate Use and Consolidation. *Nature Reviews. Neuroscience* **19**:744-57
- Joshi Abhilasha** (2017) Oxford, United Kingdom: University of Oxford.

- Joshi Abhilasha**, Denovellis Eric L., Mankili Abhijith, Meneksedag Yagiz, Davidson Thomas J., Gillespie Anna K., Guidera Jennifer A., Roumis Demetris, Frank Loren M. (2023) Dynamic Synchronization between Hippocampal Representations and Stepping. *Nature* **617**:125-31
- Joshi Abhilasha**, Salib Minas, Viney Tim James, Dupret David, Somogyi Peter (2017) Behavior-Dependent Activity and Synaptic Organization of Septo-Hippocampal GABAergic Neurons Selectively Targeting the Hippocampal CA3 Area. *Neuron* **96**:1342-1357.
- Karlsson Mattias P.**, Frank Loren M. (2009) Awake Replay of Remote Experiences in the Hippocampus. *Nature Neuroscience* **12**:913-18
- Kaur Jaspreet**, Komi Salif A., Dmytriyeva Oksana, Houser Grace A., Bonfils Madelaine C. A., Berg Rune W. (2025) Pedunculopontine-Stimulation Obstructs Hippocampal Theta Rhythm and Halts Movement. *Scientific Reports* **15**:17903
- Kay Kenneth**, Chung Jason E., Sosa Marielena, Schor Jonathan S., Karlsson Mattias P., Larkin Margaret C., Liu Daniel F., Frank Loren M. (2020) Constant Sub-Second Cycling between Representations of Possible Futures in the Hippocampus. *Cell* **180**:552-567.
- Kay Kenneth**, Sosa Marielena, Chung Jason E., Karlsson Mattias P., Larkin Margaret C., Frank Loren M. (2016) A Hippocampal Network for Spatial Coding during Immobility and Sleep. *Nature* **531**:185-90
- Kim J. J.**, Fanselow M. S. (1992) Modality-Specific Retrograde Amnesia of Fear. *Science* **256**:675-77
- Kim Steve M.**, Frank Loren M. (2009) Hippocampal Lesions Impair Rapid Learning of a Continuous Spatial Alternation Task. *PLoS One* **4**:e5494
- Király Bálint**, Domonkos Andor, Jelítai Márta, Lopes-Dos-Santos Vítor, Martínez-Bellver Sergio, Kocsis Barnabás, Schlingloff Dániel, et al. (2023) The Medial Septum Controls Hippocampal Supra-Theta Oscillations. *Nature Communications* **14**:6159
- Kloc Michelle L.**, Velasquez Francisco, Niedecker Rhys W., Barry Jeremy M., Holmes Gregory L. (2020) Disruption of Hippocampal Rhythms via Optogenetic Stimulation during the Critical Period for Memory Development Impairs Spatial Cognition. *Brain Stimulation* **13**:1535-47
- Lee KH**, Denovellis EL, Ly R, Magland J, Soules J, Comrie AE, Gramling DP, Guidera JA, Nevers R, Adenekan P, et al. (2025) Spyglass: a framework for reproducible and shareable neuroscience research. *bioRxiv* <https://doi.org/10.1101/2024.01.25.577295> | PubMed | PubMed Central
- Ledberg Anders**, Robbe David (2011) Hippocampal Theta Oscillations Synchronize with Rhythmical Head Movements during Locomotion. *BMC Neuroscience* **12** <https://doi.org/10.1186/1471-2202-12-S1-P266>
- Lepperød Mikkel Elle**, Christensen Ane Charlotte, Kinden Lensjø Kristian, Buccino Alessio Paolo, Yu Jai, Fyhn Marianne, Hafting Torkel (2021) Optogenetic Pacing of Medial Septum Parvalbumin-Positive Cells Disrupts Temporal but Not Spatial Firing in Grid Cells. *Science Advances* **7**:eabd5684
- Lisman John E.**, Jensen Ole (2013) The θ - γ Neural Code. *Neuron* **77**:1002-16
- Liu Can**, Todorova Ralitsa, Tang Wenbo, Oliva Azahara, Fernandez-Ruiz Antonio (2023) Associative and Predictive Hippocampal Codes Support Memory-Guided Behaviors. *Science* **382**:ead8237
- Martorell Anthony J.**, Paulson Abigail L., Suk Ho-Jun, Abdurrob Fatema, Drummond Gabrielle T., Guan Webster, Young Jennie Z., et al. (2019) Multi-Sensory Gamma Stimulation Ameliorates Alzheimer's-Associated Pathology and Improves Cognition. *Cell* **177**:256-271.
- Mizumori S. J.**, Perez G. M., Alvarado M. C., Barnes C. A., McNaughton B. L. (1990) Reversible Inactivation of the Medial Septum Differentially Affects Two Forms of Learning in Rats. *Brain Research* **528**:12-20
- Mizuseki Kenji**, Sirota Anton, Pastalkova Eva, Buzsáki György (2009) Theta Oscillations Provide Temporal Windows for Local Circuit Computation in the Entorhinal-Hippocampal Loop. *Neuron* **64**:267-80
- Morris R. G.**, Garrud P., Rawlins J. N., O'Keefe J. (1982) Place Navigation Impaired in Rats with Hippocampal Lesions. *Nature* **297**:681-83

- Mouchati Philippe R.**, Kloc Michelle L., Holmes Gregory L., White Sheryl L., Barry Jeremy M. (2020) Optogenetic ‘Low-Theta’ Pacing of the Septohippocampal Circuit Is Sufficient for Spatial Goal Finding and Is Influenced by Behavioral State and Cognitive Demand. *Hippocampus* **30**:1167-93
- O’Keefe J.** (1976) Place Units in the Hippocampus of the Freely Moving Rat. *Experimental Neurology* **51**:78-109
- O’Keefe J.**, Recce M. L. (1993) Phase Relationship between Hippocampal Place Units and the EEG Theta Rhythm. *Hippocampus* **3**:317-30
- Panoz-Brown Danielle**, Iyer Vishakh, Carey Lawrence M., Sluka Christina M., Rajic Gabriela, Kestenman Jesse, Gentry Meredith, et al. (2018) Replay of Episodic Memories in the Rat. *Current Biology: CB* **28**:1628-1634.
- Pastalkova Eva**, Itskov Vladimir, Amarasingham Asohan, Buzsáki György (2008) Internally Generated Cell Assembly Sequences in the Rat Hippocampus. *Science* **321**:1322-27
- Petersen Peter Christian**, Buzsáki György (2020) Cooling of Medial Septum Reveals Theta Phase Lag Coordination of Hippocampal Cell Assemblies. *Neuron* **107**:731-744.
- Petsche H.**, Stumpf C., Gogolak G. (1962) [The Significance of the Rabbit’s Septum as a Relay Station between the Midbrain and the Hippocampus. I. The Control of Hippocampus Arousal Activity by the Septum Cells]. *Electroencephalography and Clinical Neurophysiology* **14**:202-11
- Pfeiffer Brad E.**, Foster David J. (2013) Hippocampal Place-Cell Sequences Depict Future Paths to Remembered Goals. *Nature* **497**:74-79
- Quirk Clare R.**, Zutshi Ipshita, Srikanth Sunandha, Fu Maylin L., Marciano Naomie Devico, Wright Morgan K., Parsey Darian F., et al. (2021) Precisely Timed Theta Oscillations Are Selectively Required during the Encoding Phase of Memory. *Nature Neuroscience* **24**:1614-27
- Robinson Jennifer C.**, Wilmot Jacob H., Hasselmo Michael E. (2023) Septo-Hippocampal Dynamics and the Encoding of Space and Time. *Trends in Neurosciences* **46**:712-25
- Robinson Jennifer C.**, Ying Johnson, Hasselmo Michael E., Brandon Mark P. (2023) Optogenetic Silencing of Medial Septal GABAergic Neurons Disrupts Grid Cell Spatial and Temporal Coding in the Medial Entorhinal Cortex. *BioRxiv* <https://doi.org/10.1101/2023.11.08.566228>
- Robinson Jennifer**, Manseau Frédéric, Ducharme Guillaume, Amilhon Bénédicte, Vigneault Erika, El Mestikawy Salah, Williams Sylvain (2016) Optogenetic Activation of Septal Glutamatergic Neurons Drive Hippocampal Theta Rhythms. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience* **36**:3016-23
- Rübel Oliver**, Tritt Andrew, Ly Ryan, Dichter Benjamin K, Ghosh Satrajit, Niu Lawrence, Baker Pamela, Soltesz Ivan, Ng Lydia, Svoboda Karel, et al. (2023) The Neurodata Without Borders ecosystem for neurophysiological data science. *eLife* **11**:e78362
- Salib Minas**, Joshi Abhilasha, Katona Linda, Howarth Michael, Micklem Benjamin R., Somogyi Peter, Viney Tim J. (2019) GABAergic Medial Septal Neurons with Low-Rhythmic Firing Innervating the Dentate Gyrus and Hippocampal Area CA3. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience* **39**:4527-49
- Scoville W. B.**, Milner B. (1957) Loss of Recent Memory after Bilateral Hippocampal Lesions. *Journal of Neurology, Neurosurgery, and Psychiatry* **20**:11-21
- Siegle Joshua H.**, Wilson Matthew A. (2014) Enhancement of Encoding and Retrieval Functions through Theta Phase-Specific Manipulation of Hippocampus. *eLife* **3**:e03061 <https://doi.org/10.7554/eLife.03061>
- Skaggs W. E.**, McNaughton B. L., Wilson M. A., Barnes C. A. (1996) Theta Phase Precession in Hippocampal Neuronal Populations and the Compression of Temporal Sequences. *Hippocampus* **6**:149-72
- Smith Anne C.**, Frank Loren M., Wirth Sylvia, Yanike Marianna, Hu Dan, Kubota Yasuo, Graybiel Ann M., Suzuki Wendy A., Brown Emery N. (2004) Dynamic Analysis of Learning in Behavioral Experiments. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience* **24**:447-61

- Vancura Bert**, Geiller Tristan, Grosmark Andres, Zhao Vivian, Losonczy Attila (2023) Inhibitory Control of Sharp-Wave Ripple Duration during Learning in Hippocampal Recurrent Networks. *Nature Neuroscience* **26**:788-97
- Varga Csaba**, Golshani Peyman, Soltesz Ivan (2012) Frequency-Invariant Temporal Ordering of Interneuronal Discharges during Hippocampal Oscillations in Awake Mice. *Proceedings of the National Academy of Sciences of the United States of America* **109**:E2726-34
- Viney Tim J.**, Lasztocki Balint, Katona Linda, Crump Michael G., Tukker John J., Klausberger Thomas, Somogyi Peter (2013) Network State-Dependent Inhibition of Identified Hippocampal CA3 Axo-Axonic Cells in Vivo. *Nature Neuroscience* **16**:1802-11
- Viney Tim James**, Salib Minas, Joshi Abhilasha, Unal Gunes, Berry Naomi, Somogyi Peter (2018) Shared Rhythmic Subcortical GABAergic Input to the Entorhinal Cortex and Presubiculum. *eLife* **7** <https://doi.org/10.7554/eLife.34395>
- Wang Yingxue**, Romani Sandro, Lustig Brian, Leonardo Anthony, Pastalkova Eva (2015) Theta Sequences Are Essential for Internally Generated Hippocampal Firing Fields. *Nature Neuroscience* **18**:282-88
- Wang Yingxue**, Roth Zachary, Pastalkova Eva (2016) Synchronized Excitability in a Network Enables Generation of Internal Neuronal Sequences. *eLife* **5** <https://doi.org/10.7554/elife.20697>
- Yamamoto Jun**, Tonegawa Susumu (2017) Direct Medial Entorhinal Cortex Input to Hippocampal CA1 Is Crucial for Extended Quiet Awake Replay. *Neuron* **96**:217-227.
- Yong Hyun Choong**, Chang Haoran, Brandon Mark P. (2022) Optogenetic Reduction of Theta Oscillations Reveals That a Single Reliable Time Cell Sequence Is Not Required for Working Memory. *BioRxiv* <https://doi.org/10.1101/2022.06.25.497592>
- Yu Jai Y.**, Pettibone Jeffrey R., Guo Caiying, Zhang Shuqin, Saunders Thomas L., Hughes Elizabeth D., Filipiak Wanda E., et al. (2018) Knock-in Rats Expressing Cre and Flp Recombinases at the Parvalbumin Locus. *BioRxiv* <https://doi.org/10.1101/386474>
- Zutshi Ipshita**, Brandon Mark P., Fu Maylin L., Donegan Macayla L., Leutgeb Jill K., Leutgeb Stefan (2018) Hippocampal Neural Circuits Respond to Optogenetic Pacing of Theta Frequencies by Generating Accelerated Oscillation Frequencies. *Current Biology: CB* **28**:1179-1188.

Peer reviews

Reviewer #1 (Public review):

Summary:

This manuscript by Joshi and colleagues demonstrates that the precise theta-phase timing of spikes is causal for CA1 hippocampal theta sequences during locomotion on a linear track and is necessary for learning the cognitively demanding outbound component of a hippocampus-dependent alternation task (W-maze), independently of replay during immobility. To reach these conclusions, the authors developed a theta-phase-specific, closed-loop manipulation that used optogenetic activation of medial septal parvalbumin (PV) interneurons at the ascending phase of theta during locomotion. This protocol preserved immobility periods, allowing a clean and elegant dissociation from SWR-associated replay.

The manuscript is well written and was a pleasure to read. The work described is of high quality and introduces several notable advances to the field:

- (a) It extends prior studies that manipulated theta oscillations by examining precise temporal structure (specifically theta sequences) rather than only LFP features.
- (b) The closed-loop manipulation enabled dissociation between deficits in theta sequences during a behavioural task and SWR-associated replay activity.

(c) As controls, the authors included rats with suboptimal viral transduction or optic-fibre placement, and, within subjects, both stimulation-on (stim-on) and stimulation-off (stim-off) trials. Notably, sequence disruption persisted into stim-off periods within the same session.

Overall, this is a strong manuscript that will provide valuable insights to the field. I have only minor comments:

(1) As the authors note, it is striking that both behavioural performance and spike patterns are altered during stim-off trials. They propose that "disruption of theta sequences during the initial experience in an environment is sufficient to have lasting effects," implying that rapid, experience-dependent plasticity is driven by sequential firing. Does this imply that if rats were previously trained on the task, subsequent stim-on and stim-off trials would yield different outcomes, with stim-off trials showing improved performance and intact theta sequences? For example, if the sequence of one-third stim-on, one-third stim-off, one-third stim-on were inverted to off-on-off, would theta sequences be expected to emerge, disappear, and potentially re-emerge? While I am not asking for additional experiments, I think the discussion could be extended in this aspect.

Alternatively, could the number of stim-off trials (one third of the total) be insufficient to support learning/induce plasticity? In the controls, ~50-100 trials appear necessary to achieve high performance.

(2) In line with the point above, the authors characterise the behavioural changes induced by MS optogenetic stimulation specifically as a "learning deficit," as rats failed to improve across 300 trials in an initially novel environment (W-maze). While they present this as complementary to prior demonstrations of impaired performance on previously learned tasks (Zutshi et al., 2018; Quirk et al., 2021; Etter et al., 2023; Petersen et al., 2020), an alternative interpretation is a working-memory deficit. This would produce the same behavioural pattern, with reference memory (the less cognitively demanding trials) remaining intact despite stimulation and concomitant changes in theta sequences. This interpretation would also be consistent with work in certain disease models, where reduced synaptic plasticity and working-memory deficits co-occur with preserved place coding despite impaired theta sequences (e.g., Viana da Silva et al., 2024; Donahue et al., 2025).

(3) It was not immediately clear whether SWR-associated activity was derived from the interleaved ~15-min rest sessions in a rest box, or from periods of immobility or reward consumption in the maze (aSWR, as in Jadhav et al 2012). Regardless, it would be informative to compare aSWR events within the maze to rest-box SWRs that may occur during more prolonged slow-wave episodes (even if not full sleep). This contrasts with Liu et al. (2024), who analysed replay during ~1.5-h sleep sessions.

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Reviewer #2 (Public review):

Summary:

The authors of this study developed a closed-loop optogenetic stimulation system with high temporal precision in rats to examine the effect of medial septum (MS) stimulation on the disruption of hippocampal activity at both behavioral and compressed time scales. They found that this manipulation preserved hippocampus single-cell-level spatial coding but affected theta sequences and performance during a spatial alternation task. The performance deficits were observed during the more cognitively demanding component of the task and even persisted after the stimulation was turned off. However, the effects of this disruption were confined to locomotor periods and did not impact waking rest replay, even during the

early phase of stimulation-on. Their conclusion is consistent with previous findings from the Pastalkova lab, where MS disruption (using different methods) affected theta sequences and task performance but spared replay (Wang et al., 2015; Wang et al., 2016). However, it differs from a recent study in which optogenetic disruption of EC inputs during running affected both theta sequences and replay (Liu et al., 2023).

Strengths:

The experiments were well designed and controlled, and the results were generally well presented.

Weaknesses:

Major concerns are primarily technical but also conceptual. To further increase the impact of this study by contrasting findings from different disruptions, it is necessary to better align the analysis and detection methods.

Major concerns:

(1) To show that MS disruption does not affect spatial tuning, the authors computed the KL divergence of tuning curves between stimulation-on and stimulation-off conditions. I have two main questions about this analysis:

(1.1) The authors seem to impose stringent inclusion criteria requiring a large number of spikes and a strong concentration of tuning curves. These criteria may have selected strongly spatially tuned cells, which are typically more stable and potentially less vulnerable to perturbations. Based on the Figure 2 caption, it seems that fewer than 10% of cells were included in the KL divergence analysis, which is lower than the usual proportion of place cells reported in the literature. What is the rationale for using such strict inclusion criteria? What happens to the cells that are not as strongly tuned but are still identified as significant place cells?

(1.2) The KL divergence was computed between stimulation-on and stimulation-off conditions within the same animal group. However, the authors also showed that MS stimulation had lasting effects on theta sequences and performance even during stimulation-off periods. Would that lasting effect also influence spatial tuning? Based on these questions, the authors should perform additional analyses that directly measure spatial tuning quality and compare results across control and experimental groups - for example, spatial information of spikes (Skaggs et al., 1996), tuning stability, field length, and decoding error during running.

(2) The authors compared their results with those from Liu et al. (2023) and proposed that the different outcomes could be explained by different sites of disruption. However, the detection and quantification methods for theta sequences and replay differ substantially between the two studies, emphasizing different aspects of the phenomenon. I am not suggesting that either method is superior, but providing additional analyses using aligned detection methods would better support the authors' interpretations and benefit the field by enabling clearer comparisons across studies. In the current analysis, the power spectrum of the decoded ahead/behind distance only indicates that there is a rhythmic pattern, without specifying the decoding features at different theta phases. Moreover, the continuous non-local representations during ripples could include stationary representations of a location or zigzag representations that do not exhibit a linear sequential trace. Given that, the authors should show averaged decoding results corrected by the animal's actual position within theta cycles and compute a quadrant ratio. For replay analysis, they could use a linear fit (as in Liu et al., 2023) and report the proportion of significant replay events.

(3) The finding that theta sequences and performance were impaired even during stimulation-off periods is particularly interesting and warrants deeper exploration. In the

Discussion, the authors claim that this may arise from "the rapid plasticity engaged during early learning." However, this explanation does not fully account for the observation. Previous studies have shown that theta sequences can develop very rapidly (Feng et al., Foster lab, 2015; Zhou et al., Dragoi lab, 2025). If the authors hypothesize that rapid plasticity during early stimulation-on disrupts the theta sequence, then the plasticity window must also be short and terminate during the subsequent stimulation-off period. Otherwise, why can't animals redevelop theta sequences during stimulation-off? The authors should conduct additional analyses during the stimulation-off periods of the W-maze task. For example:

- (3.1) What is the spike-theta phase relationship? Do the phases return to normal or remain altered as during stimulation-on?
- (3.2) Is there a significant place-field remapping from stimulation-on to stimulation-off? (Supplementary Figure 3F includes only a small subset of cells; what if population vector correlations are computed across all cells, or Bayesian decoding of stimulation-on spikes is performed using stimulation-off tuning curves?)
- (3.3) The authors should also discuss why the stimulation-off epochs were not sufficient to support learning, and if the stimulation-off place cell sequences could have supported replay.
- (4) Citations and/or discussion of key studies relevant to the current work are missing: Wang et al. in Pastalkova lab 2015-2016 studies for disruption of theta sequence (but not place cell sequence) disrupting learning but not replay, Drieu et al. in Zugaro lab 2018 study on disruption of theta sequence affecting sleep replay, Farooq and Dragoi 2019 for association between a lack of theta sequence and presence of waking rest replay during postnatal development, etc. The authors should discuss what the conceptually new findings in the current study are, given the findings of the previous literature above.
- (5) The assessment of theta sequence is not state-of-the-art:
 - (5.1) Detecting the peak of cross-correlograms between neurons (CCG) relates to behavioral timescale CCG, not the theta sequence one; for the theta sequence, the closest to zero local peak should be used instead.
 - (5.2) How were other methods of detecting theta sequences performing on the stimulation-on/stimulation-off data: Bayesian decoding, firing sequences?
 - (5.3) How was phase precession during stimulation-on/stimulation-off?
- (6) It would be important to calculate additional variables in the replay part of the study to compare the quality of replay across the 2 groups:
 - (6.1) Proportion of significant replay events out of the detected multiunit events.
 - (6.2) The average extent of trajectory depicted by the significant replay events in the targeted compared to the control, stimulation-on/stimulation-off.

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Reviewer #3 (Public review):

Joshi et al. present an elegant and technically rigorous study examining how the temporal structure of hippocampal spiking during locomotion contributes to spatial learning. Using a closed-loop, theta phase-specific optogenetic manipulation of medial septal parvalbumin-expressing neurons in rats, the authors demonstrate that disrupting theta-timescale coordination impairs performance on the cognitively demanding component outbound trajectory of a spatial alternation task, while sparing hippocampal replay, place coding, and the simpler inbound learning. The work aims to dissociate the role of theta-associated

temporal organization during navigation from sharp-wave ripple-associated replay during subsequent rest periods, providing a mechanistic link between theta sequences and learning. The findings have important implications for models of septo-hippocampal coordination and the functional segregation between online (theta) and offline (SWR) network states. That said, there are a few conceptual and methodological issues that need to be addressed.

One concern is the overall novelty of this work; the dissociation between online temporal sequence and offline replay events following memory deficits has previously been shown by Wang et al., 2016 *eLife*. While the authors discuss Lui et al., 2023, which demonstrates MEC activation of inhibitory neurons at gamma frequencies during locomotion disrupts theta sequences, subsequent replay and learning (line 65-66), they do not reference Wang et al., 2016 who performed a very similar study with MS pharmacological inactivation, and report large decreases in theta power, attenuated theta frequencies together with behavioural deficits but SWR replay persisted. Given strong similarities in the manipulation and findings, this study should be discussed.

Along the same lines, it should be noted that Brandon et al. (2014, *Neuron*) demonstrated that hippocampal place codes can still form in novel environments despite MS inactivation and loss of theta, indicating that spatial representations can emerge without intact septal drive. Referencing this study would strengthen the discussion of how temporal coordination, rather than spatial coding per se, underlies the learning deficits observed here.

The conclusion that disrupting "theta microstructure" impairs learning relies on the assumption that the observed behavioral deficits arise from altered temporal coding from within hippocampal CA1 only. However, optogenetic modulation of medial septal PV neurons influences multiple downstream regions (entorhinal cortex, retrosplenial cortex) via widespread GABAergic projections. While the authors do touch on this, their discussion should expand to include the network-level consequences of entorhinal grid-cell disruption and how this could affect temporal coding both online and offline.

The finding that replay content, rate, and duration are unchanged is critical to the paper's claim of dissociation. However, the analysis is restricted to immobility on the track. Given evidence for distinct awake vs. sleep replay, confirming that off-track rest and post-session sleep replays are similarly unaffected would confirm the conclusions of the paper. If these data are unavailable, the limitation should be acknowledged explicitly. Moreover, statistical power for detecting subtle differences in replay organization or spatial bias should be added to the supplement (n of events per animal, variability across sessions).

The exact protocol for optogenetic stimulation is a bit confusing. For the task, the first and final third (66%) of trials were disrupted and were only stimulated when away from the reward well and only when the animal was moving. What proportion of time within "stimulated" trials remained unstimulated? Why were only 66% of trials stimulated?

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Author response:

We thank all reviewers for their overall assessment, thoughtful comments, and suggestions. We are working to address each reviewer's comment in detail. In this provisional response, we provide clarifications regarding our experimental approach and the novelty of our work, and include additional analyses that we have performed since the submission of the manuscript. We are also happy to report that we have now shared the raw data, intermediate analysis files, and the complete repository to facilitate replication of the analysis and figures.

Code repo: github.com/LorenFrankLab/ms_stim_analysis

Data repo: dandiarchive.org/dandiset/001634

Docker containers (see GitHub repo for use instructions):

Database: https://hub.docker.com/r/samuelbray32/spyglass-db-ms_stim_analysis

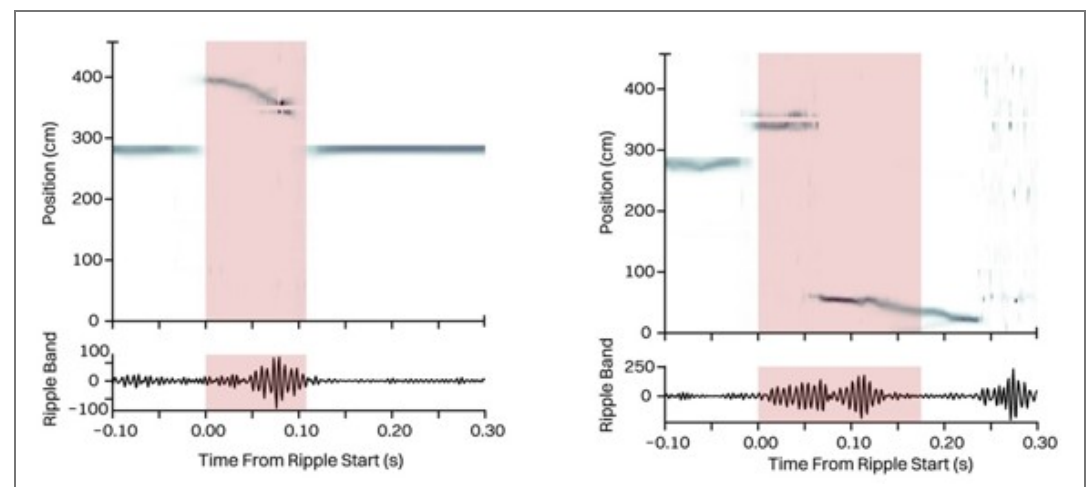
Python notebooks: https://hub.docker.com/r/samuelbray32/spyglass-hub-ms_stim_analysis

(1) *Novelty and contrast with earlier manipulations:*

We thank the reviewers for suggesting that we explicitly contrast our results with prior pharmacological (Wang et al., 2016; Wang et al., 2015; Koenig et al., 2011; Brandon et al., 2014), systemic (Robbe & Buzsáki 2009; Petersen and Buzsáki 2020), and behavioral (Drieu et al., 2018) manipulations that also assessed some of the physiological features we evaluated. We will add a discussion of these studies, which will help us emphasize both the insights and discrepancies observed using these prior approaches. We will also more clearly explain the the novelty and importance of our specific approach for temporally and physiologically precise manipulation. Specifically, our approach (closed-loop theta-phase stimulation during locomotion) provides a level of physiological specificity that made it possible to dissociate theta-state dynamics from other hippocampal processes. This in turn allowed us to address a question that has remained unresolved across prior studies: Are hippocampal spatial sequences during locomotion (i.e., theta sequences) necessary to learn a novel hippocampal-dependent task?

(2) *Additional analysis on SWRs during rest:*

since submitting the manuscript, we have conducted additional analysis on the rate and length of SWRs in the rest box and found that their rate and length are also indistinguishable between targeted and control animals (effect of manipulation between control and targeted animals; rSWR rate: $p=0.45$; rSWR length: $p=0.94$, mixed effect model). We also find evidence for sequential neural representations in the rest box, when the encoding was performed in the behavioral arena. Example trajectories are shown below. These results are consistent with our observations on SWRs rate, length, and content in the behavioral arena. Additionally, we are in the process of evaluating and quantifying the results of decoding the rSWRs and will include those in the next version of the manuscript.



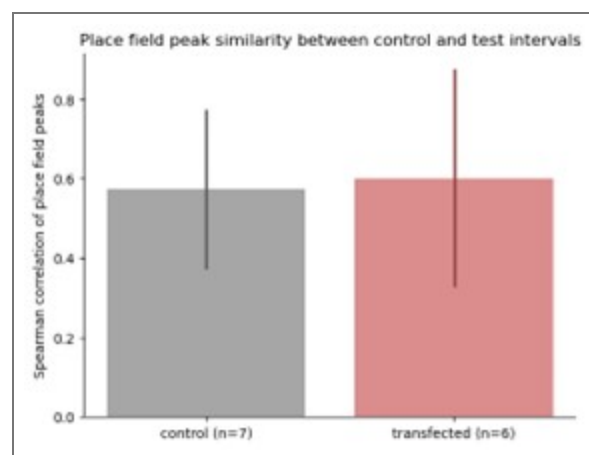
Author response image 1. Sequential replay events observed in the rest box

(3) *Theta sequence measurement in the absence of theta:*

In the next version of the manuscript, we will explicitly explain why our manipulation makes it more appropriate to measure sequential hippocampal representations during locomotion (i.e., theta sequences) without using theta oscillation or an epoch-averaged relatively large sliding window as a reference. The key insight here is that our manipulation suppresses theta and thus makes it difficult or impossible to accurately identify theta phase. We understand that theta-phase based approaches were used in prior work; however, these prior analyses may have confounded the absence of hippocampal theta sequences during locomotion by the inability to detect theta oscillatory phase reliably. We will show that our method of using clusterless Bayesian decoding in which we estimate the decoded position at every 2ms timestep is indeed able to capture endogenous hippocampal sequences even without imposing any requirements of aligning to theta oscillations, thus providing an unbiased estimate of the rhythmicity of hippocampal spatial representations.

(4) Additional analysis on place cell stability and tuning:

We thank the reviewer for this question. For the KL divergence analysis, we have imposed a spike-count criterion (100 spikes for each interval type —stimulation-off, stimulation-on, and the stimulus sub-interval) and a coverage criterion (50% HPD of the units' spatial firing distribution was contained within 40cm on the linear track and 100cm on the w-track). These criteria were chosen to ensure that spatial tuning curves were sufficiently well sampled and localized to allow reliable estimation of KL divergence, which is particularly sensitive to noise arising from low spike counts or diffuse firing. Based on the reviewer's suggestion, we have relaxed the unit inclusion criteria for KL divergence by relaxing the criteria for number of spikes and spatial coverage criterion to include more weakly tuned place cells and replicated our results ($p=.146$). Further, we have also evaluated the stability of place field order between stimulation-on and stimulation-off conditions using more standard methods (as in Wang et. al., 2015; spearman correlation of place field order, control vs targeted, $p = .920$, t-test). These results are consistent with our observations about place field stability during stimulation-off and stimulation-on conditions (Fig. 2F).



Author response image 2. Spearman correlation of place field order during stimulation-on and stimulation-off conditions.

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