

Reviewed Preprint

v1 • January 22, 2026

Not revised

Reviewed Preprint

v2 • April 2, 2026

Revised by authors

✉ For correspondence:

a-contractor@northwestern.edu

Competing interests: No

competing interests declared

Funding: See [page 23](#)

Reviewing editor: Laura L Colgin,
University of Texas at Austin, United
States

© 2026, Castillon et al. This article is
distributed under the terms of the
[Creative Commons Attribution
License](#), which permits unrestricted
use and redistribution provided that
the original author and source are
credited.

Subregional activity in the dentate gyrus is amplified during elevated cognitive demands

Charlotte CM Castillon¹, Shintaro Otsuka¹, John N Armstrong¹, Anis Contractor^{1,2,3} ✉¹Department of Neuroscience, Weinberg College of Arts and Sciences Northwestern University, Evanston, United States• ²Department of Psychiatry and Behavioral Sciences Feinberg School of Medicine, Weinberg College of Arts andSciences Northwestern University, Evanston, United States • ³Department of Neurobiology, Weinberg College of Arts and Sciences Northwestern University, Evanston, United States

eLife Assessment

This manuscript presents a **valuable** study of the activity and functional relevance of different circuits in the dentate gyrus of mice performing a pattern separation task. **Solid** evidence is presented to support the paper's central conclusions. The study is likely to be of interest to those studying the subregional organization and cell type-specific functions of the dentate gyrus.

<https://doi.org/10.7554/eLife.109611.2.sa4>

Abstract

Neural activity in the dentate gyrus (DG) is required for the detection and discrimination of novelty, context and patterns, amongst other cognitive processes. Previous studies have shown that granule cell activation differs between the supra and infrapyramidal blades of the DG during a range of hippocampal dependent tasks yet how excitatory dynamics within the DG drive and modulate this blade-specific bias under varying cognitive demands remains unclear. Here we used an automated touch screen pattern separation task combined to temporally controlled tagging of active neurons to determine how increasing cognitive demand shapes the spatial patterns of activity in the DG. We observed that as task difficulty increased, DG activation became progressively biased toward the suprapyramidal blade, accompanied by structured distributions of active mature granule cells (mGCs) along the apex-to-blade and hilar-to-molecular layer axes. Selective inhibition of mature granule cells (mGCs) did not affect the patterned distribution of active cells but profoundly impaired performance, as mice were no longer able to discriminate between closely spaced locations. In contrast, chemogenetic inhibition of adult-born granule cells (abDGCs) beyond a critical window of their maturation significantly impaired performance of mice during high-demand conditions, elevated overall mGC activity and disrupted the blade-specific distribution of active mGCs even in animals that successfully completed the task. These findings demonstrate how a high cognitive demand pattern separation task preferentially activates mGCs in subregions of the DG and are consistent with a modulatory role for abDGCs on the dentate circuit which in part governs the spatially organized patterns of activity of mGCs.

Introduction

The dentate gyrus (DG) is the input structure to the hippocampus and plays important roles in episodic memory¹, spatial coding², novelty detection³ as well as in separating patterns⁴. A long-standing proposal stemming from early computational studies, suggest that the DG contribute to pattern separation based on its anatomical connectivity⁵⁻⁷. The DG is the gateway to the hippocampus receiving direct excitatory input from the entorhinal cortex (EC). The large number

of granule neurons in the DG receiving input from a relatively smaller population of EC has been proposed to promote expansion recoding which, along with sparse firing of granule cells, allows the separation of inputs at the level of neural activity⁸.

The granule cell layer of the DG is organized into two blades, termed the suprapyramidal (SB) and infrapyramidal (IB) blades. When viewed in transverse cross-section⁹, mature granule cells (mGCs) exhibit asymmetric activation across these blades during hippocampal-dependent tasks^{10–13}, with greater activity typically observed in the SB compared to the IB^{10–12}. However, the circuit-level mechanisms underlying this functional asymmetry remain unresolved.

In addition to mature populations, the subgranular zone (SGZ) of the DG serves as a neurogenic niche where new granule neurons continue to be generated throughout adulthood^{14–17}. This process of adult neurogenesis gives rise to adult-born dentate granule cells (abDGCs), which can remodel local circuitry and potentially contribute to memory flexibility¹⁸. Interestingly, the asymmetry in mGC activation across the blades parallels an asymmetry in neurogenesis, with higher numbers of abDGCs reported in the SB^{19–22}.

In this study, we investigated whether performance in a cognitively demanding task alters the patterns of neural activity in the dorsal dentate gyrus (DG), and how manipulation of excitatory populations within the DG can modulate these activity patterns. Using TRAP2 mice²³ to label neurons activated during a touchscreen-based spatial discrimination task, we observed a consistent bias in activity between the two blades of the DG. When the cognitive demand of the task was elevated by reducing the degree of spatial separation, there was an increase of TRAP cells activity and a more prominent and amplified spatial bias with active neurons distributed preferentially to the SB and relatively reduced in the IB. To dissect the contribution of distinct excitatory populations, we selectively inhibited either mature or adult-born granule cells during task performance. We found that manipulating these populations had differential effects on the blade-specific bias of activation, indicating distinct roles for mGCs and abDGCs in shaping DG representations during spatial cognition. Inhibition of mature granule cells did not alter the blade bias of activation but markedly impaired performance, as mice were no longer able to discriminate closely spaced locations under high cognitive demand. In contrast, chemogenetic inhibition of abDGCs beyond a critical window of their maturation, decreased task performance, increased overall mGC activity, and reduced the blade-specific bias of activation, suggesting that abDGC activity is required to maintain proper DG network organization under high cognitive load. Together, these findings reinforce the significance of neural activity in the dorsal DG in spatial pattern separation and highlight the circuit contributions of both abDGCs and mGCs to behaviors requiring a high cognitive demand.

Results

Amplified blade-biased activity of GCs during a high cognitive demand pattern separation task

Prior work has demonstrated that expression of immediate early genes in the DG including Arc¹⁰, c-Fos¹² and Zif268²⁴ correlate with behavioral state and are active in both mature and young abDGCs. Genetic strategies using TRAP1 mice²⁵ and TetTag mice^{26,27} have also demonstrated that active GC populations can be labeled with these strategies. To determine whether behaviorally induced labeling of both mGCs and abDGCs is observed in TRAP2 mice (Fos^{2a-iCreERT2};Ai9) we combined birth-dating of abDGCs with a single exposure to voluntary wheel running which is known to activate DG neurons²⁵ (Figure S1A [↗](#)). Mice given access to running wheels for 4 hours had elevated numbers of TRAP+ neurons throughout the DG, with both mature (mGCs; defined as granule cells not birth-dated with BrdU or EdU) and immature neurons labeled (Figure S1B–F [↗](#)). TRAP+ labeling of abDGCs was observed in birth-dated neurons 14 days post injection (dpi) from BrdU (Figure S1E & F [↗](#)). No double labeled TRAP+ BrdU+ neurons were observed in younger neurons prior to this timepoint (Figure S1F [↗](#)).

Importantly, an analysis of the blade specific localization of TRAP⁺ neurons demonstrated that in home cage controls there was a small bias in the distribution of active neurons to the SB of the DG and this distribution of labeled GCs was not different in mice exposed to a single episode of running (Figure S1D [↗](#)). Moreover, consistent with previous reports¹⁹⁻²¹, we observed an asymmetry in neurogenesis, with a greater number of abDGCs in the SB compared to the IB (Figure S1G [↗](#)).

In order to determine the activation patterns of GCs (mGCs and abDGCs) when mice successfully performed a high cognitive demand spatial discrimination pattern separation task, TRAP2 mice were trained in a modified touch screen task that tests the ability to separate a unique pattern of illuminated squares posing a relatively low or high cognitive demand (Figure 1A [↗](#) and Figure S2A & B [↗](#)). The trial unique nonmatching-to-location (TUNL) task has been demonstrated to require the hippocampus, particularly when the cognitive demand is high when the separation of the sample to test squares is reduced, or the delay between sample and choice is increased^{28,29}. Mice were injected with BrdU to birthdate neurons at day 0 (D0). At D7 mice were water restricted before EdU injections and shaping at D14 to habituate them to the operant task and learn to touch the screen for a saccharine water reward. Mice were then trained until they reached criterion (70% correct during one of two daily sessions on 2 consecutive days) in the TUNL task with a 1s delay between the sample and choice and a large (L) separation between sample panel and the test panel (Figure 1A [↗](#) and Figure S2A [↗](#)). Mice reached criterion on average at 10.7 ± 0.8 days in the large separation sessions (Figure 1B & D [↗](#) left panel). After reaching criterion the mice were tested with alternating trials of large and small (LS) separation between the sample and test panels. In this case the mice were always above criterion for the L trials but took another 9.3 ± 1.0 days to reach criterion on the S trials (Figure 1C [↗](#) & D middle and right panel).

Different cohorts of mice were analyzed at each level of cognitive demand of the task to assess the activation of mGCs and abDGCs. TRAP labeling was used to identify active neurons in the DG, together with thymidine analogs that birth-dated abDGCs at approximately 3 weeks (25 ± 1.4 days, EdU) and 5 weeks (39 ± 1.2 days, BrdU) of age (Figure 1E [↗](#)). We found a strong relationship between the level of cognitive demand of the task and the degree of mGC activation with a greater number of TRAP⁺ neurons in mice tested under higher cognitive load.

Specifically, TRAP⁺ neuron density increased from shaping to the L separation stage, and was highest in mice performing the most cognitively demanding condition (LS) (one-way ANOVA, $F(2,25) = 7.97$, $p = 0.0021$; Tukey post-hoc test. Groups Shap vs LS and Large vs LS were significantly different) (Figure 1F [↗](#)), regardless of whether the mice reach the criterion (C) or not (NC) (Figure S2H [↗](#)). A similar trend was observed when assessing c-Fos labeling 90 minutes after the final training session, further supporting the link between task demand and mGC activation (Figure S2F [↗](#)). Furthermore, when we examined the correlation between successful performance to criterion and the number of TRAP⁺ neurons, we found no significant relationship (Figure S2C [↗](#)), demonstrating that increased DG activity is driven by engagement in a high-demand task rather than task success.

Activation of adult-born granule cells (abDGCs) birth-dated with BrdU and EdU was assessed by colocalization with TRAP⁺ cells, rather than density, because the low number of newborn neurons precluded reliable density measurements. Accordingly, results are expressed as the proportion of birth-dated cells that were TRAP⁺. Using this approach, we did not observe a relationship between the level of cognitive demand and the recruitment of newborn neurons. Only a small fraction (~1%) of 25- to 39-day-old abDGCs were active (TRAP⁺) during the high cognitive demand task (LS) (Figure 1G [↗](#)), regardless of whether the mice reached criterion (C) or not (NC) (Figure S2J [↗](#)). Given the sparse labeling in this population, this percentage reflects very low absolute cell numbers per animal. In contrast, mice that performed only the low-demand version of the task (L configuration) exhibited almost no TRAP-labeled birth-dated cells (Figure 1G [↗](#)). Moreover, in the shaping group, no newborn neurons were co-labeled with TRAP, indicating that minimal cognitive engagement does not recruit this population (Figure 1G [↗](#)). Importantly, we did not detect qualitative differences in the overall abundance or blade distribution of BrdU⁺ or EdU⁺ neurons across behavioral groups (not shown).

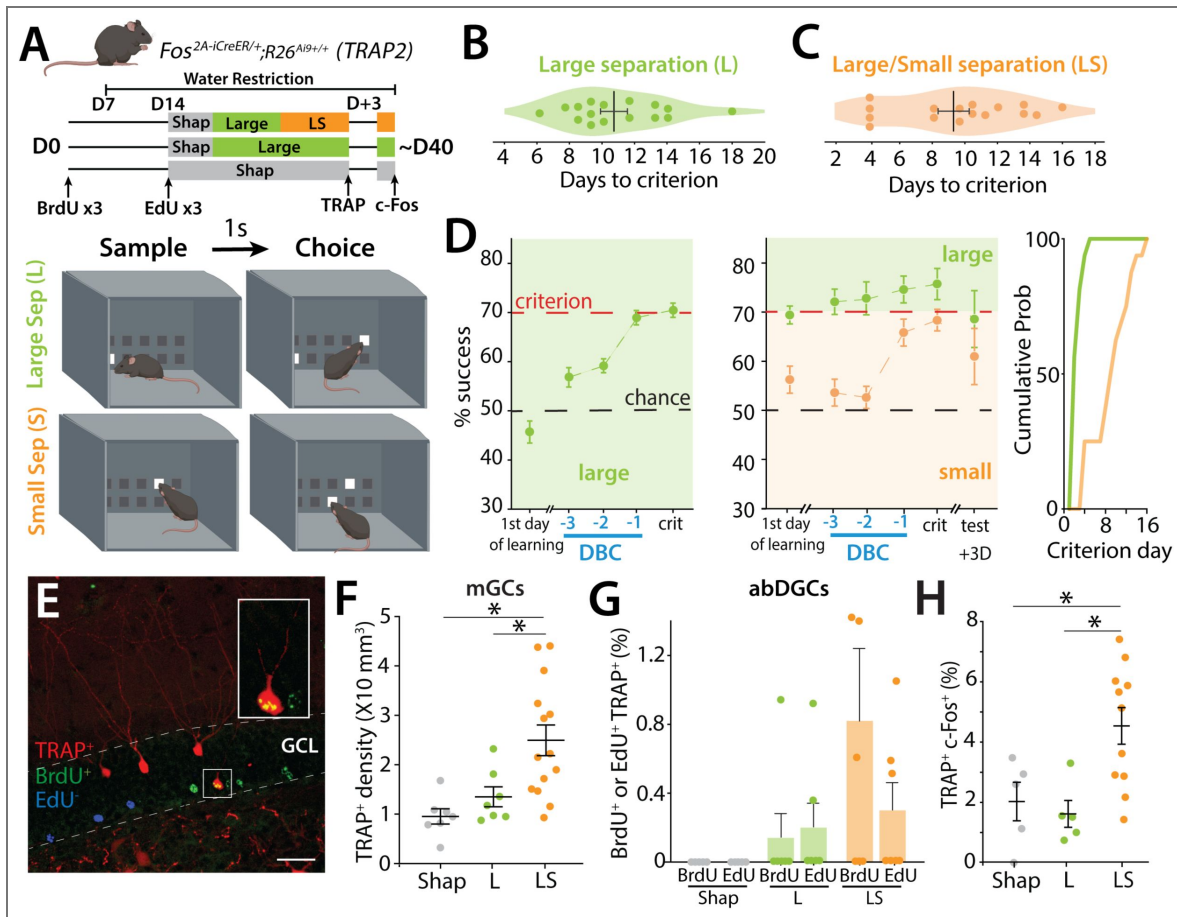


Figure 1. TRAP labeling of mGCs and birth-dated abDGCs after the TUNL pattern separation task.

(A) *Top*: Schematic illustrating the pattern separation paradigm. Male ($n = 9$) and female ($n = 7$) mice (6-8-week-old) received 3 injections of BrdU at D0 and 3 injections of EdU two weeks later (D14) before starting the behavioral paradigm. Mice received one injection of 4-OHT on the day they reached criterion (TRAP) and were perfused 3 days later 90 minutes after one exposure to the behavioral paradigm to allow endogenous c-Fos expression (c-Fos). Control mice (males $n = 6$, females $n = 4$) underwent the same treatment but only participated in the Large separation task (L). Mice were water-restricted during the behavioral experiment. *Bottom*: Cartoon representation of the chamber in TUNL task with examples of Large separation (L) and Small (S) separation configurations. High cognitive demand paradigm consisted of interleaved L and S trials (LS). (B) Number of days for each mouse to reach the 70% success criterion in S trials, during the LS separation task. (C) Number of days for mice to reach 70% success criterion in S trials, during the LS separation task. (D) *Left*: Average percentage of correct choices (% success) in the L separation on the first day of training, as well as during the three days prior to reaching criterion (days before criterion, DBC) and the day of reaching the criterion (crit) (one-sample t -test to chance at criterion Large $t(16) = 12.04$, $p < 0.0001$). *Middle*: Average percentage of correct choices in the L and S separation on the first day of training, during the three days before criterion (DBC), on the day of reaching criterion and 3 days later (one-sample t -test to chance at criterion Large $t(16) = 8.38$, $p < 0.0001$; Small $t(16) = 7.37$, $p < 0.0001$). *Right*: Cumulative probability of days to criterion of mice in S for both controls L and LS groups. (E) Example section of BrdU+ labeling (green) EdU+ labeling (blue) and TRAP+ labeling (red) in TRAP2 mice. Calibration bar: 30 μm . (F) Density of TRAP-labeled GCs in mice undergoing LS compared to control mice engaged in either shaping (Shap) or the Large configuration task only (L) (one-way ANOVA, $F(2,25) = 7.97$, $p = 0.0021$; Tukey post-hoc test. Groups Shap vs LS and Large vs LS were significantly different). (G) Percentage of BrdU labeled cells (39 days ± 1.23) or Edu labeled cells (25 days ± 1.37 days) also TRAP+ after L or LS training (BrdU+/TRAP+ L: $0.145 \pm 0.145\%$, 6 mice; LS BrdU+/TRAP+ $0.822 \pm 0.474\%$, 6 mice; ns; Edu+/TRAP+ L: $0.203 \pm 0.144\%$, 6 mice; LS BrdU+/TRAP+ $0.304 \pm 0.155\%$, 7 mice; ns; Mann-Whitney). (H) Percentage of TRAP+ GCs expressing c-Fos+ in Shap, L or LS task after reaching criterion (expressed as a percentage of TRAP+ cells) (one-way ANOVA, $F(2,18) = 6.66$, $p = 0.0069$; Tukey post-hoc test. Groups Shap vs LS and Large vs LS were significantly different).

In a separate cohort, we TRAPed neurons during the final day of LS trials and re-exposed mice to the same task 3 days later. After two LS sessions, mice were perfused 90 minutes later for immunohistochemical detection of endogenous c-Fos (Figure 1A). Task performance was reduced compared to the final day of prior training in both the L and S trials (Figure 1D, middle panel). Analysis of mGC reactivation (TRAP⁺/c-Fos⁺ cells) revealed greater overlap in mice performing LS trials than in those performing only L separations or shaping sessions (one-way ANOVA, $F(2,18) = 6.66$, $p = 0.0069$; Tukey post-hoc test. Groups Shap vs LS and Large vs LS were significantly different) (Figure 1H). Notably, across all animals and conditions, no BrdU⁺ or EdU⁺ cells were found to be labelled with both TRAP and c-Fos, indicating that newborn neurons activated during the final LS session were not reactivated three days later. Collectively, these results indicate that while increasing cognitive demand enhances activation and reactivation of mature granule cells, the recruitment of adult-born neurons remains low and does not scale with task difficulty.

Consistent with previous reports^{10–13}, we observed a bias in granule cell activity within the DG, with TRAP⁺ mGCs preferentially localized to the suprapyramidal blade (SB). Interestingly, this bias was not fixed but varied according to the level of cognitive demand of the task (Figure 2A). Mice performing the high-demand LS condition showed a pronounced enrichment of TRAP⁺ mGCs in the SB compared to the infrapyramidal blade (IB), driven by both an increase in labeled cells in the SB and a decrease in the IB (two-way ANOVA (unbalanced) (group: $F(1,38) = 146.34$, $p = 1.33 \times 10^{-14}$) (blade: $F(2,38) = 1.14 \times 10^{-7}$, $p = 1$) (group \times blade: $F(2,38) = 13.99$, $p = 2.81 \times 10^{-5}$, Tukey post-hoc test, LS and Large ($p=0.03$), LS and Shap ($p=0.05$) for SB and IB) (Figures 2A and S2E). In contrast, mice performing lower-demand versions of the task still exhibited a significant SB–IB difference, but the magnitude of this bias was reduced, resulting in a more balanced distribution of activity across blades. However, as with overall TRAP⁺ cell density, there was no significant correlation between individual task performance and the distribution of TRAP⁺ neurons in the SB (Figure S2D).

Further spatial mapping of TRAP⁺ cells relative to the SGZ and dentate apex revealed that active neurons were concentrated in the outer radial portion of the granule cell layer—an area enriched in spatially tuned semilunar granule cells^{11,30}, and this localization was particularly evident after the high-demand LS task (Figures 2B and 2C). In addition, active neurons were preferentially located toward the distal tip of the suprapyramidal blade (two-way ANOVA (unbalanced) (group: $F(1,26) = 26.05$, $p = 2.56 \times 10^{-5}$) (distance (0%-100%): $F(2,26) = 0.044$, $p = 0.957$) (group \times distance (0%-100%): $F(2,26) = 23.77$, $p = 1.35 \times 10^{-6}$). Tukey post-hoc test: significant differences were observed between Shap; 0%-50% and 50%-100% ($p<0.0001$), LS; 0%-50% and 50%-100% ($p=0.03$) (Figure 2C). Together, these results demonstrate that the suprapyramidal bias in granule cell activity is dynamically modulated by cognitive demand, with the separation becoming more pronounced as task difficulty increases and producing distinct distributions of active neurons along both the apex–distal and hilar–outer axes of the DG (Figures 2D and 2E).

Activation of GCs during remote recall in the high cognitive demand task

We next investigated whether a similar spatially biased activity of GCs was observed in a remote recall (R) test three weeks after reaching the criterion during the initial training (I), and whether there was overlap in the population of active DG neurons (Figure 3A). In the remote session, mice had a high success rate performing close to criterion in the L trials on the first day of retesting suggesting that 3 weeks after initial learning they retained the ability in task (Figure 3B). However, when confronted with the more demanding trials with inclusion of the small separation configuration, mice required additional days to achieve the criterion level (Figure 3C), although most mice required less time to reach criterion in LS training during the remote trials than on the initial testing (Figure 3D). We again observed that the density of TRAP⁺ cells was increased in mice that underwent LS compared to L only trials during the initial training (not shown) and importantly the distribution of active neurons during both the initial training (TRAP⁺) and remote training (c-Fos⁺) were biased to the SB after LS training (Figure 3F & G) (Two-way

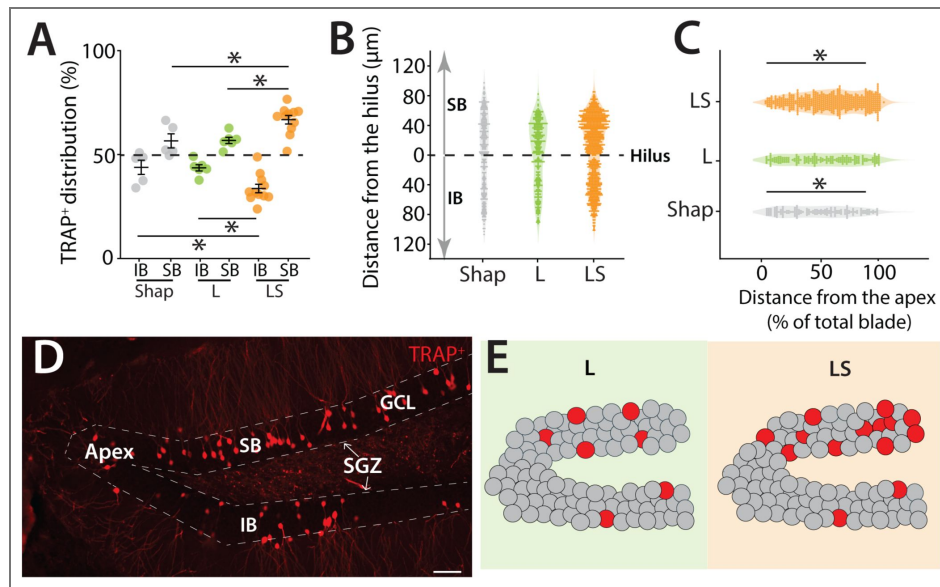


Figure 2. Blade-biased activity of mGCs during a high cognitive demand pattern separation task.

(A) Spatial distribution of TRAP-labeled cells in the infrapyramidal (IB) and suprapyramidal (SB) blades of the dorsal DG in TRAP2 mice performing the LS separation, compared to control mice that performed only the Shap or L configuration (two-way ANOVA (unbalanced) (group: $F(1,38) = 146.34, p = 1.33 \times 10^{-14}$) (blade: $F(2,38) = 1.14 \times 10^{-7}, p = 1$) (group \times blade: $F(2,38) = 13.99, p = 2.81 \times 10^{-5}$). Tukey post-hoc test: significant differences were observed between IB; LS and Large ($p=0.03$), LS and Shap ($p=0.05$), SB; LS and Large ($p = 0.03$), LS and Shap ($p=0.05$)). (B) Spatial distribution of TRAP+ cells in the hilar to outer radial GCL axes in the SB (0 to 120 μ m) and the IB (0 to 120 μ m) of the dorsal DG with 0 μ m indicating location at the hilar border (Shap: 5 mice, 255 cells; L: 5 mice, 294 cells; LS: 5 mice, 639 cells). (C) Distribution of TRAP-labeled cells along the apex to blade extremity axes of the SB of the dorsal DG, with 0% indicating location at the dentate apex (Shap: 5 mice, 152 cells; L: 5 mice, 176 cells; LS: 5 mice, 517 cells) (two-way ANOVA (unbalanced) (group: $F(1,26) = 26.05, p = 2.56 \times 10^{-5}$) (distance (0%-100%): $F(2,26) = 0.044, p = 0.957$) (group \times distance (0%-100%): $F(2,26) = 23.77, p = 1.35 \times 10^{-6}$). Tukey post-hoc test: significant differences were observed between Shap; 0%-50% and 50%-100% ($p<0.0001$), LS; 0%-50% and 50%-100% ($p=0.03$)). (D) Example section of TRAP+ cells in the dorsal DG illustrating the distinct boundaries between the two blades, the SGZ, and the apex. Calibration bar: 50 μ m. (E) Cartoon illustrating the two blades of the DG: IB and SB. Dentate granule cells are depicted in grey while red circles represent TRAP+ cells in the DG. In the L group, a greater number of labeled cells are localized in the SB compared to the IB. This bias of activity is also observed in the LS group, where the bias is more pronounced, and the overall activity of the DG is increased. In the SB, the majority of the activated cells are located closer to the tip of the blade.

ANOVA: group, $F(1,12) = 32.98$, $p < 0.001$; blade, $F(1,12) = 9.2 \times 10^{-30}$, $p = 1$; group \times blade interaction, $F(1,12) = 16.92$, $p = 0.0014$. Tukey post-hoc test: significant differences were observed between IB; LS and Large ($p=0.05$), SB; LS and Large ($p= 0.05$)(3G: Two-way ANOVA: group, $F(1,12) = 56.03$, $p < 0.00001$; blade, $F(1,12) = 0$, $p = 1$; group \times blade interaction, $F(1,12) = 7.02$, $p = 0.021$. Tukey post-hoc test: no significant differences were observed between LS and Large for IB or SB).

An analysis of birth-dated active neurons labeled during the initial training (TRAP⁺) or during the remote trials (c-Fos⁺) demonstrated a small number of BrdU (~4 weeks) neurons were labeled during the initial training (Figure 3H) whereas in the remote training we observed both BrdU labeled (7.7 weeks) and EdU labeled (5.7 weeks) abDGCs that were c-Fos⁺ only in mice after LS training (Figure 3I). No birth-dated abDGCs were found that were double labeled TRAP⁺ and c-Fos⁺, however we observed dual labeled mGCs with more TRAP⁺/c-Fos⁺ labeled neurons in mice that had performed LS training than those that only performed L training (Unpaired t-test, $t(6) = 2.90$, $p = 0.027$) (Figure 3J).

Taken together, these results demonstrate that during initial training and remote training in the pattern separation task, spatially biased activity of neurons is observed in the SB but there is little overlap in activity of neurons in the DG of mice during these epochs of training.

mGC activity is required for DG function and efficient pattern separation in TUNL

Given that the suprapyramidal bias in DG activity scaled with cognitive demand, we hypothesized that altering the excitatory drive within the DG might modulate this spatial pattern of activation. We began by targeting mature granule cells (mGCs), the principal excitatory population in the DG, which are under strong inhibitory control and sparsely active in vivo^{31,32}.

To selectively suppress their activity, Dock10-Cre mice³³ were crossed with R26-LSL-Gi-DREADD mice (designer receptor exclusively activated by designer drugs) to express the inhibitory DREADD hM4Di, which can be activated by the ligand deschloroclozapine (DCZ)(50 $\mu\text{g.kg}^{-1}$)³⁴ during the LS sessions of the task (Figure 4C). Characterization of Dock10-Cre expression in newborn neurons (BrdU⁺) revealed few labeled cells at 2 weeks postmitosis, with progressively more double-labeled abDGCs in older, non-DCX-positive neurons (Figures 4A and 5B), reaching ~80% of abDGCs by 7 weeks (Figures 4A and 4B). This pattern confirms that Dock10-Cre is expressed primarily in mGCs and older abDGCs. As predicted, DREADD-mediated inhibition of mGCs impaired task performance, with the majority of mice failing to reach criterion (62.5%) within the 14-day LS cutoff, compared to controls (34.8%) ($p = 0.0452$) (Figures 4D and 4E). Performance in the interleaved low-demand L trials was not affected (not shown). Consistent with reduced DG activity, we observed lower c-Fos expression in both the DG (Unpaired t-test, $t(9) = 2.30$, $p = 0.047$) and CA3 (Unpaired t-test, $t(9) = 4.93$, $p = 0.0008$) of mice that did not reach criterion after 14 days (Figures 4F, 4G, and 4I). Interestingly, despite the overall reduction in activity, the distribution of active c-Fos⁺ neurons remained biased toward the SB, similar to control mice that reached criterion (Figures 4H) (two-way ANOVA (unbalanced) (group: $F(1,16) = 315.83$, $p = 5.87 \times 10^{-12}$; blade: $F(1,16) = 0$, $p = 1$; group \times blade: $F(1,16) = 0.040$, $p = 0.845$). This suggests that the overall excitability is a key feature of DG function during performance of a high cognitive demand pattern separation task (Figures 4H, 4J, and 4K).

DREADD inhibition of abDGCs disrupts blade-biased activity of mGCs and performance of mice in a high cognitive demand task

Having established that mGCs regulate overall DG activity but do not determine the SB-IB bias, we next asked whether manipulating abDGCs could influence this bias and the pattern of DG activation. Because mGC inhibition reduced global excitability without altering the spatial organization of active cells, this suggested that the blade-biased pattern may be set upstream by a more selective cell population. Given that abDGCs constitute a smaller, more plastic excitatory population, we hypothesized that selectively altering their activity might differentially affect both overall excitability and the spatial distribution of active neurons during high cognitive demand

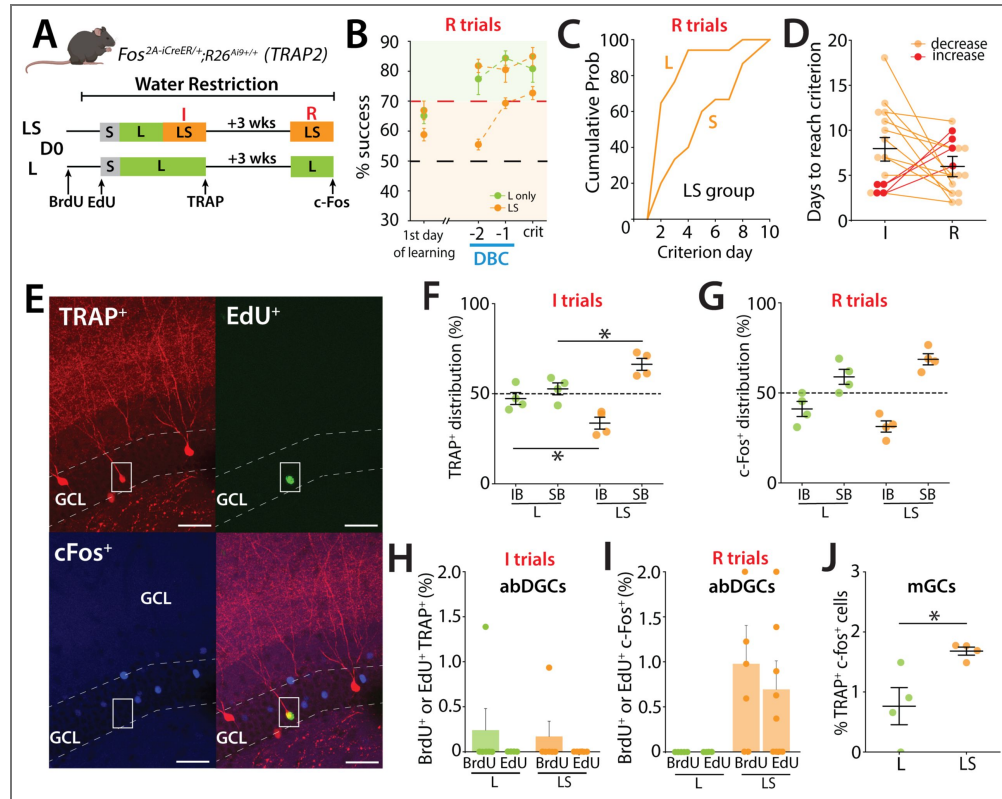


Figure 3. Activation of GCs during remote recall in the high cognitive demand task.

(A) Schematic representation of the pattern separation paradigm used in the study. Mice (male $n=11$, female $n=8$) performed two training sessions: initial (I) and remote (R). The second training (R) took place 3 weeks after the mice reached the S criterion in LS. Mice were perfused when they reached the criterion in S during R. A second group that performed only in the L configuration during both training sessions was included as control (males $n=3$, females $n=3$). (B) Learning curves representing the percentage of success during the remote test (R). The control group trained only in the large-separation condition (L group) is shown in green and includes performance in the large-separation trials only. The LS group (orange) was trained in both large- and small-separation conditions and therefore includes two traces: the upper orange trace represents performance during large-separation trials, and the lower orange trace represents performance during small-separation trials. (one-sample t -test to chance at criterion; LS: Large $t(14) = 11.8$, $p < 0.0001$, Small $t(14) = 10.3$, $p < 0.0001$; L: Large $t(3) = 6.78$, $p = 0.0066$). (C) Percentage of mice reaching the L and S criterion in LS group, during the remote test (R) represented as cumulative probability. During the remote test, mice reach the L criterion in just a few days but require more time to reach the S criterion. (D) Performance of individual mice indicating a non-significant reduction in the number of days needed to reach the criterion between the initial and remote training sessions. Only 4 mice (red lines) required more days to reach the criterion during the remote test. (I: 8.067 ± 1.173 days, R: 5.93 ± 0.753 days, 15 mice; $p = 0.204$; Mann-Whitney). (E) EdU labeled cell that was also TRAP+ labeled after the initial training but was that was not reactivated (c-Fos) during the remote test. EdU (green), TRAP (red), c-Fos (blue) in TRAP2 mice. Calibration bar, 10 μm . (F) Distribution of TRAP-labeled cells in the IB and SB blades of the dorsal DG in TRAP2 mice. Labeling was performed in LS separation during initial training (I) and compared to control mice that performed only the L configuration. Active neurons show a preferential distribution to the SB over the IB in the DG. This bias is more prominent in the LS group of mice (Two-way ANOVA: group, $F(1,12) = 32.98$, $p < 0.001$; blade, $F(1,12) = 9.2 \times 10^{-30}$, $p = 1$; group \times blade interaction, $F(1,12) = 16.92$, $p = 0.0014$. Tukey post-hoc test: significant differences were observed between IB; LS and Large ($p=0.05$), SB; LS and Large ($p=0.05$). (G) Distribution of c-Fos labeled cells between the IB and SB blades of the dorsal DG in TRAP2 mice performing the LS separation during remote training (R), compared to control mice that performed only the L configuration (Two-way ANOVA: group, $F(1,12) = 56.03$, $p < 0.00001$; blade, $F(1,12) = 0$, $p = 1$; group \times blade interaction, $F(1,12) = 7.02$, $p = 0.021$. Tukey post-hoc test: no significant differences were observed between LS and Large for IB or SB). (H) Percentage of newborn neurons (BrdU (7.7 weeks) or EdU (5.7 weeks)) activated during the initial training (I), as indicated by co-labeling of TRAP+ (BrdU+ c-Fos+; L: 0.242 ± 0.242 %, 6 mice, LS: 0.170 ± 0.170 %, 6 mice; ns; Mann-Whitney). (I) Percentage of newborn neurons (BrdU or EdU) activated during the remote test (R), as indicated by c-Fos expression (BrdU+ c-Fos+; 0.977 ± 0.426 %, 6 mice, $p=0.05$; EdU+ c-Fos+; 0.694 ± 0.317 %, 9 mice; $p=0.0785$; Mann-Whitney). (J) Percentage of mGCs activated during the (I) training (TRAP+) that were reactivated during the R test (c-Fos+) (expressed as a percentage of TRAP+ cells) (Unpaired t -test, $t(6) = 2.90$, $p = 0.027$).

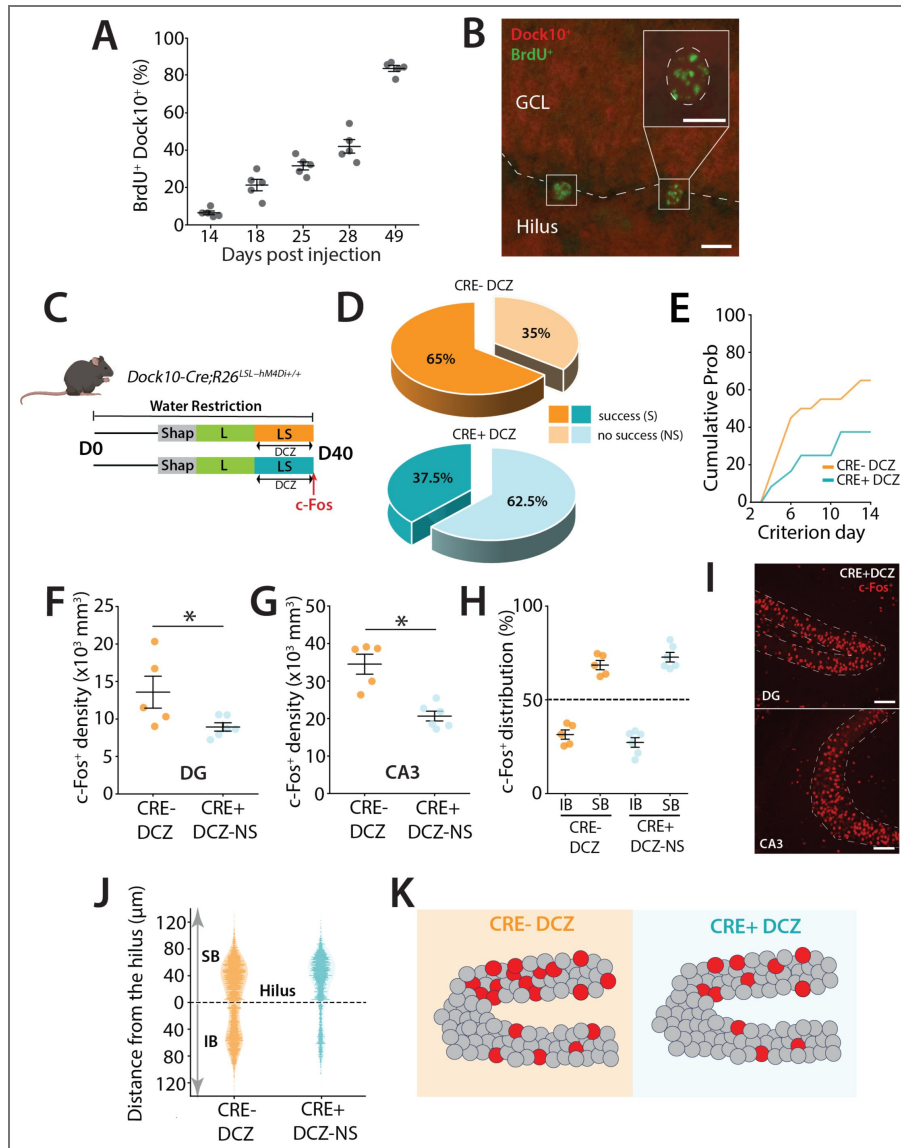


Figure 4. DREADD inhibition of mGCs during a high cognitive demand task.

(A) Percentage of BrdU-labeled cells that are also Dock10+ at different time points following BrdU injections (in days). (B) Representative section showing BrdU+ labeling (green) and Dock10+ labeling (red) in Dock10 cre;Ai9 mice. Calibration bars: 10 μ m. (C) Schematic representation of the pattern separation paradigm. All mice (males n = 17, females n = 27) received DCZ i.p. injections 30 minutes before the beginning of their LS trials. (D) Percentage of mice reaching criterion in (S) and percentage that were not successful (NS) after 14 days of training in S configuration of LS. A majority of mice never succeeded in reaching the criterion in the Cre+ DCZ group ($z = 2.00$, $p = 0.0453$, z -test - Group NS). (E) Percentage of mice reaching the criterion in S trials across days represented as the cumulative probability including all successful and not successful mice in the group. (F) Density of c-Fos+ GCs in the dorsal DG across the two groups. A significantly reduced density was observed in the Cre+ DCZ group compared to controls (Unpaired t-test, $t(9) = 2.30$, $p = 0.047$). (G) Density of c-Fos+ GCs in the dorsal CA3 across the two groups. A significant reduction was observed in the Cre+ DCZ group compared to the control groups (Unpaired t-test, $t(9) = 4.93$, $p = 0.0008$). (H) Blade specific distribution of c-Fos+ GCs in the IB and SB of the dorsal DG in mice performing LS separation (Two-way ANOVA (unbalanced, Type II) revealed a significant main effect of group ($F(1,16) = 315.83$, $p = 5.87 \times 10^{-12}$), but no significant main effect of blade ($F(1,16) = 0$, $p = 1$) and no significant group \times blade interaction ($F(1,16) = 0.040$, $p = 0.845$). (I) Example of dorsal DG section from Dock10-cre;hM4Di mice after LS training. c-Fos+ neurons are labeled in red. Calibration bars: 100 μ m. (J) Spatial distribution of c-Fos+ along the hilar to outer radial GCL axis of the DG (0 to 120 μ m), with 0 μ m indicating location at the SGZ. (K) Cartoon illustrating the distribution of activity labeled GCs in the SB and IB blades of the DG. GCs are depicted in grey while red circles represent activated cells in the DG. The overall distribution of neurons in mice in which mGCs were inhibited (Cre+ DCZ) and which did not reach criterion was not different to controls even though the total density of activity labeled neurons was lower.

tasks. abDGCs are known to affect the excitability of mGCs^{35,36} and there is a demonstrated blade bias in neurogenesis in the DG. However, it remains unknown whether abDGCs affect the blade-biased activity patterns of mGCs that are evident during a pattern separation task. To acutely modulate the activity of abDGCs during task performance, we created mice in which the inhibitory DREADD (designer receptor exclusively activated by designer drugs) could be conditionally expressed in abDGCs by crossing R26-LSL-Gi-DREADD mice (hM4Di)³⁷ to *Ascl1*^{CreERT2} mice. This selectively expresses the hM4Di inhibitory DREADD in a birth-dated cohort of abDGCs by the delivery of TAM (tamoxifen, see Methods) in the diet of mice for 7 weeks allowing the inhibition of their activity with a selective ligand deschloroclozapine (DCZ)(50 $\mu\text{g}\cdot\text{kg}^{-1}$)³⁴ during the LS sessions of the task (Figure 5A & B) ^{38,39}. Inhibition of abDGCs (≤ 7 weeks old) using the DREADD ligand DCZ increased the time animals took to reach criterion compared to control groups of animals that received DMSO (one-way ANOVA, $F(2,31) = 3.77$, $p = 0.034$; Tukey post-hoc test indicate that Cre+ DCZ group is significantly different from Cre+ DMSO, while other pairs are not significantly different) (Figure 5C & D). This impairment was specific to the cognitively more demanding LS task, as inhibiting ≤ 7 -week-old abDGCs did not affect the performance of mice once they had learned the L configuration (not shown). Importantly control groups that had also received TAM in their diet showed no effect in the pattern separation task or in the biased blade activity of mGCs (Figure 5C-J). Taken together this experiment suggests that abDGCs in this cohort (≤ 7 weeks) are beneficial to performance in a pattern separation task.

Prior work has demonstrated that abDGC loss causes an increase in activity of mGCs^{35,40} which is likely through a loss of connections to hilar interneurons which provide feedback inhibition to the DG. Consistent with this, analysis of c-Fos in the DG demonstrated increased labeling across the DG when abDGCs were inhibited with DCZ activation of hM4Di compared to control groups ($p=0.0271$) (Figure 5E & F). Conversely, c-Fos+ labeling in the downstream CA3 region of the hippocampus demonstrated no significant difference in density between the control groups and abDGC inhibition (Figure 5G) (one-way ANOVA, $F(2,13) = 2.27$, $p = 0.143$; Tukey post-hoc test indicates that there is no significant difference between any pairs of groups).

Interestingly, we found that the biased distribution of c-Fos⁺ neurons in the two control groups showed the same profile as we had observed before in the IB and SB after mice perform the high cognitive demand task (Figure 5H). However, in the cohort of mice in which abDGCs were inhibited, the prominent biased distribution pattern of activity was not observed (two-way ANOVA (unbalanced) (group: $F(1,24) = 659.55$, $p < 2.2 \times 10^{-16}$; blade: $F(2,24) = 3.48 \times 10^{-13}$, $p = 1$; group \times blade: $F(1,16) = 0.040$, $p = 0.845$). Tukey post-hoc test. Significant differences were observed for IB comparisons: Cre+ DCZ vs Cre-DCZ ($p=0.000548$) and Cre+ DCZ vs Cre+ DMSO ($p=0.00765$). Significant differences were observed for SB comparisons: Cre+ DCZ vs Cre-DCZ ($p=0.000548$) and Cre+ DCZ vs Cre+ DMSO ($p=0.00765$) (Figure 5H).

Furthermore, the distribution of c-Fos⁺ cells within the blades showed that when abDGCs were inhibited during LS training there was a marked prominence of labeled neurons located near the hilus in both the IB and SB in the region closest to the subgranular zone (SGZ), the neurogenic niche of the DG where abDGCs are produced, raising the possibility that these neurons are under greater feedforward inhibitory control via abDGCs (Figure 5I). In summary, DREADD inhibition of ≤ 7 -week-old abDGCs during the LS high cognitive demand task resulted in a reduction in both in biased SB-IB spatial activity pattern as well as the activity labeling of neurons along both axes of the transverse DG (Figure 5J), and this altered activity correlated with a reduction in performance of animals which required increased days to complete the high cognitive demand task.

In the previous experiments we had provided TAM to mice in their diet to cause expression of hM4Di in neurons that range from 0 days to 7 weeks. This covers abDGCs that are within their critical period when they demonstrate strong plasticity and excitability and are well connected to the hippocampal network^{41,42}. Because inhibition of this 0–7-week population altered both performance and the blade-biased spatial distribution of mGC activity, we next asked whether these effects depend specifically on the maturity of the abDGCs. To test whether younger, ≤ 4 -week-old neurons contribute similarly to DG function during high cognitive demand, we provided TAM

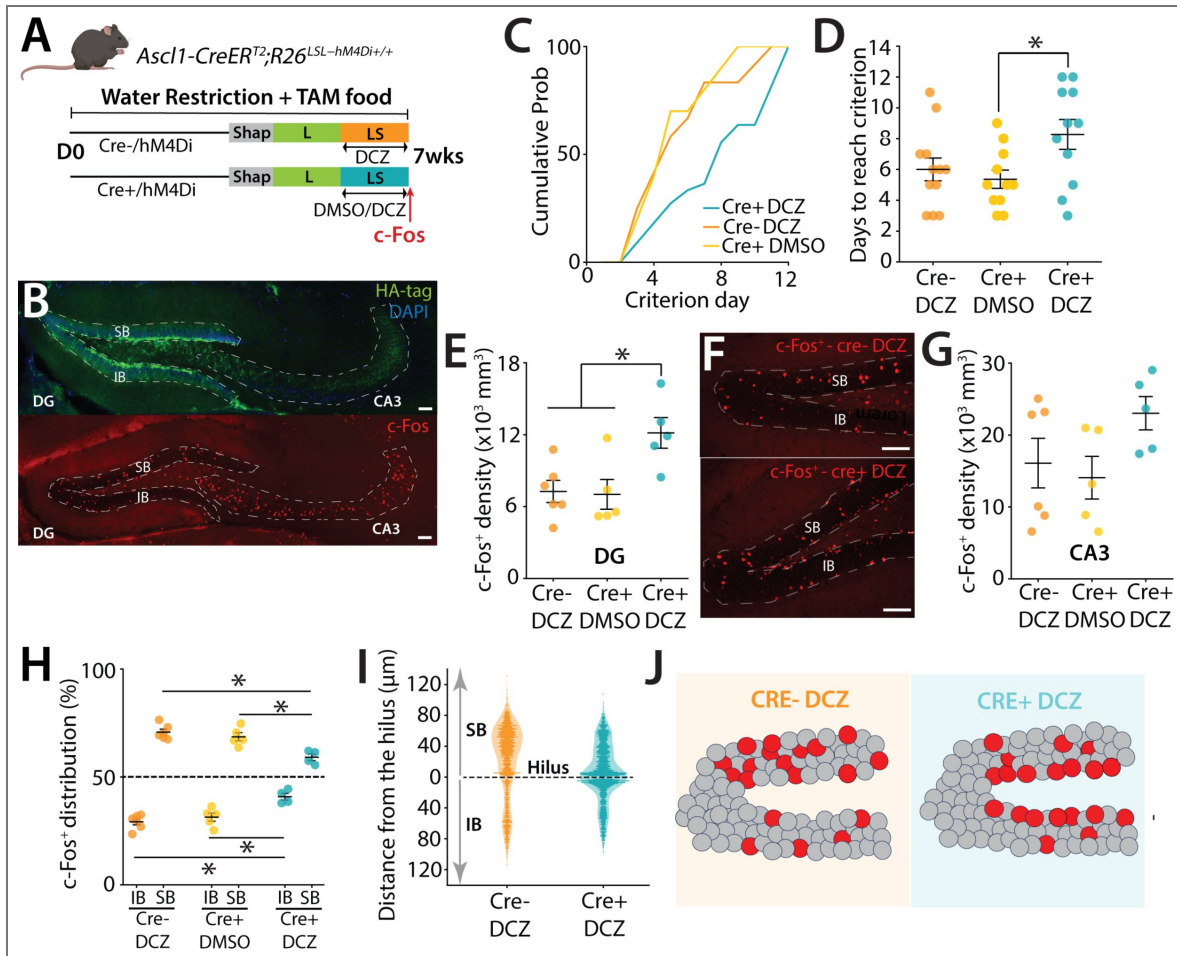


Figure 5. DREADD inhibition of ≤ 7 -week abDGCs during a high cognitive demand task

(A) Schematic representation of the pattern separation paradigm. Prior to onset of the training, mice (males $n = 17$, females $n = 16$) were provided tamoxifen-containing food ad libitum in the home cage for 7 weeks to activate TAM inducible Cre recombinase (*Ascl1-CreER^{T2}*) in abDGCs. All mice received DCZ (deschloroclozapine, $50 \mu\text{g}\cdot\text{kg}^{-1}$) or DMSO injections 30 minutes before the beginning of LS trials. (B) Example sections from *Ascl1-CreER^{T2}*; hM4Di mice with immunohistochemical localization of the HA tagged DREADD (top) and after LS training with c-Fos⁺ neurons labeled in red. Calibration bars: 140 μm . (C) Percentage of mice reaching the criterion in the S trials across days represented as the cumulative probability. The Cre+ DCZ treated group needed additional days to reach criterion compared to controls. (D) Number of days for each mouse to reach the 70% success criterion in S trials, (one-way ANOVA, $F(2,31) = 3.77$, $p = 0.034$; Tukey post-hoc test indicate that Cre+ DCZ group is significantly different from Cre+ DMSO, while other pairs are not significantly different). (E) c-Fos⁺ cell density in all groups of mice. There was an increase in c-Fos⁺ mGCs in mice in which ≤ 7 week abDGCs were inhibited by DCZ ($H(2) = 7.22$, $p = 0.0271$; Kruskal Wallis)(Cre+ DCZ comparison to Cre+ DMSO, $p = 0.014$; Cre+ DCZ comparison to Cre-DCZ, $p = 0.027$ with Dunn's multiple comparison post hoc analysis). (F) Example section after c-Fos immunohistochemistry to assess distribution of c-Fos⁺ mGCs between the IB and SB of the dorsal DG in groups of mice who received DCZ (cre-vs cre+). c-Fos cells are localized closer to the SGZ in the Cre+ group. Calibration bar, 100 μm . (G) c-Fos⁺ density in the CA3 region of the hippocampus in each of the groups of mice. No significant difference was observed in any of the groups (one-way ANOVA, $F(2,13) = 2.27$, $p = 0.143$; Tukey post-hoc test indicates that there is no significant difference between any pairs of groups). (H) Blade distribution of c-Fos⁺ mGCs in the IB and SB of the dorsal DG in mice performing the LS separation receiving either DCZ or DMSO 30 min prior to task performance. Active neurons are distributed preferentially to the SB than the IB in the DG for the control groups. However this distribution bias is significantly reduced in Cre+ DCZ group (two-way ANOVA (unbalanced) (group: $F(1,24) = 659.55$, $p < 2.2 \times 10^{-16}$; blade: $F(2,24) = 3.48 \times 10^{-13}$, $p = 1$; group \times blade: $F(1,16) = 0.040$, $p = 0.845$). Tukey post-hoc test. Significant differences were observed for IB comparisons: Cre+ DCZ vs Cre-DCZ ($p = 0.000548$) and Cre+ DCZ vs Cre+ DMSO ($p = 0.00765$). Significant differences were observed for SB comparisons: Cre+ DCZ vs Cre-DCZ ($p = 0.000548$) and Cre+ DCZ vs Cre+ DMSO ($p = 0.00765$). (I) Spatial distribution of c-Fos⁺ GCs along hilar to molecular layer axes of the SB (0 to 120 μm) and the IB (0 to 120 μm) of the dorsal DG, with 0 μm indicating the hilar position. c-Fos⁺ cells are localized closer to the SGZ in the Cre+ DCZ group. (J) Cartoon illustrating the DG with its two blades: IB and SB. Dentate granule cells are depicted in grey while red circles represent activity labeled cells in the DG. For Cre+ DCZ mice, the distribution of labeled neurons is closer to SGZ, away from outer radial GCL and more evenly distributed in IB and SB.

for only 4 weeks to restrict hM4Di expression to these younger abDGCs (Figure 6A). In the LS high cognitive demand task, there was no difference in the time to criterion of animals that had received DCZ administration for hM4Di inhibition of abDGCs compared to the control groups (Figure 6B). Similarly, analysis of c-Fos+ density in the DG found no difference in overall density (Figure 6C) (one-way ANOVA, $F(2,13) = 0.157$, $p = 0.857$) and the blade distribution of active mGCs after hM4Di (Figure 6D) or in the hilar to molecular layer axes distribution (Figure 6E).

Taken together these results demonstrate that ≤ 7 -week-old cohort of abDGCs that include neurons in their critical period of development influence the excitability and spatially biased pattern of activity of mGCs in the DG and also affect the performance of mice in a high cognitive demand pattern separation task. In contrast young neurons at ≤ 4 weeks do not influence either the performance of mice in the task or the spatial distribution of activity in the DG.

Discussion

Blade-specific activation patterns in the dorsal dentate gyrus during high-demand discrimination

In this study we used activity labeling techniques combined with a complex, automated visual spatial pattern separation task to capture patterns of activity of GCs in the dorsal DG. The DG is a critical part of the hippocampal formation that receives extrinsic input from cortex and other structures but is also highly regulated by local connections with hilar and CA3 neurons⁸. While many roles have been ascribed to the dorsal DG, a prominent theory has been that its activity is vital to pattern separation⁴³. We found that activity labeling of neurons demonstrated unique spatial distributions of active GCs which were amplified when the separation task was made more stringent for the mice. We characterized this increased difficulty in the task as an increase in cognitive demand as it reflected the increased difficulty associated with discriminating highly similar spatial locations. However, the change in the task parameters likely engage a complex multitude of behavioral processes, including attention, motivation, reward expectation, and engagement. Therefore, our findings should be interpreted as reflecting activity patterns in the dentate gyrus in response to increased task difficulty rather than a single isolated cognitive operation.

In addition, TRAP and c-Fos labeling integrate neuronal activity across a temporal window, these approaches primarily capture overall task engagement rather than activity linked to specific trial types or behavioral epochs. Accordingly, our findings should be interpreted as reflecting sustained neural recruitment during task performance rather than moment-to-moment encoding of individual trial events. In particular, we observed a biased distribution of active neurons in the SB, which was elevated in mice that completed the LS task. There have been prior descriptions of a blade-biased distribution of active neurons when mice engage in hippocampal dependent behaviors^{10–13} however it is still not known how these spatially defined activity patterns are modulated or established. We found that not only is there a bias towards more activity in SB and less in the IB during a more demanding task, there is also an increase in activity density in the outer radial areas of the granule cell layer (GCL) where semilunar granule cells are enriched^{11,30}, and in regions closer to the dentate apex. These elevated biased spatial distributions of active neurons were not observed when mice were assessed after running or after the low cognitive demand version of the task, two behaviors that increase the activity of mGCs significantly but do not enhance their blade-biased distribution (Figure S2E). However, while blade-specific differences in activity are evident, no distinct, independent microcircuit has been identified that exclusively governs the function of either the SB or IB. This raises an important question as to whether these differences arise solely from variations in afferent connectivity and intrinsic GC properties or if they reflect an underlying, segregated network organization within the DG. Future studies exploring whether the SB and IB operate as distinct computational units or as interconnected components of a larger network will be essential to clarify the functional basis of this observed bias.

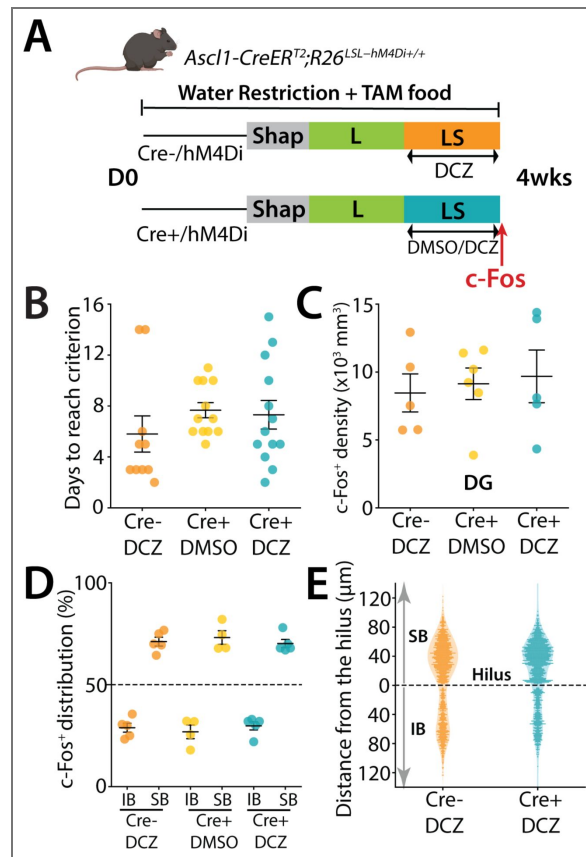


Figure 6. DREADD inhibition of ≤ 4 -week-old abDGCs does not affect performance of mice

(A) Schematic representation of the pattern separation paradigm used in the study. Prior to the behavioral experiment, mice (males $n = 12$, females $n = 23$) were given tamoxifen-enriched food for 4 weeks to activate the Cre recombinase and label newborn neurons. All mice received DCZ or DMSO injections 30 minutes before the beginning of LS trials. (B) Number of days for each mouse to reach the 70% success criterion in S trials ($H(2) = 4.21$, ns; Kruskal Wallis). (C) Density of c-Fos⁺ GCs in the dorsal DG across the different groups (one-way ANOVA, $F(2,13) = 0.157$, $p = 0.857$). (D) Blade specific distribution of c-Fos⁺ cells in the IB and SB of the dorsal DG in mice performing LS separation ($H(2) = 0.46$, ns; Kruskal Wallis). (E) Spatial distribution of c-Fos⁺ along the hilar to outer radial GCL axis of the DG (0 to 120 μ m), with 0 μ m indicating location at the border of the SGZ and hilus.

Activity level of mGCs is required for a high demand task

Mature granule cells (mGCs) constitute the primary excitatory population of the DG and are essential for its sparse coding properties. Despite their large number, mGCs are strongly regulated by local inhibition and typically remain sparsely active in vivo^{31,32}. In our study, we used an acute and reversible DREADD inhibition of mGCs using a Cre driver line expressed in mature but not in young adult-born neurons (Figure 4A, S3). Suppressing mGC activity during the LS task had a major effect on performance: most mice in the inhibited cohort failed to reach criterion. Analysis of behavior revealed that during S trials, performance remained at chance, whereas in interleaved L trials, mice continued to perform above criterion. Consistent with reduced mGC excitability, the density of c-Fos+ neurons was significantly decreased in the DG. Interestingly, despite this reduction in overall activation, the blade-biased distribution of active neurons was still evident in mice that performed the task but failed to reach criterion. Thus, unlike the effects observed when inhibiting adult-born neurons, there was no clear correlation between behavioral impairment and changes in the spatial distribution of active cells. Direct inhibition of mGCs therefore produced a fundamentally different outcome on DG circuit dynamics and task performance. While abDGC inhibition slowed performance and altered the spatial distribution of active mGCs even in mice that reached criterion, direct suppression of mGCs reduced global excitability without changing the spatial organization of active cells in animals that did or did not reach criterion (not shown). Moreover, the subset of mice that did reach criterion performed with latencies indistinguishable from control cohorts.

Prior studies using genetic manipulations to increase neurogenesis, have identified blade-specific changes in the number of c-Fos+ cells, with an increase in the IB following contextual discrimination⁴⁴, and a corresponding increase or decrease in the IB or SB respectively, after navigational learning⁴⁵. These prior findings support the hypothesis that the SB is context-specific and involved in pattern separation, while the IB is more associated with spatial representation and memory precision⁴⁶. Our study extends these observations by demonstrating that while overall mGC excitability is required for successful performance in high cognitive demand conditions, the SB–IB bias itself can persist independently of excitatory drive, suggesting that it is not directly determined by mGC activity.

Adult-born granule cells modulate DG network organization during high cognitive demand

The DG, uniquely among temporal lobe structures, contains a neurogenic niche where new neurons are continuously generated and integrated into the existing network. Post-mitotic adult-born dentate granule cells (abDGCs) mature over several weeks⁴⁷, reaching maturity by around 8 weeks post mitosis where their functional properties and connectivity resemble those of other mGCs^{48,49}. During a critical period between 4–8 weeks, however, they exhibit heightened plasticity and distinct circuit interactions⁵⁰, making them uniquely positioned to influence local computation.

It has been proposed that abDGCs modulate local circuit activity by contributing to the sparsity of mGC firing through feedback inhibition^{50–52} or through direct inhibition⁵³. This inhibitory influence is largely mediated by diverse interneuron populations in the DG, including parvalbumin-positive (PV+), somatostatin-positive (SOM+), and neuropeptide Y-positive (NPY+) interneurons, which play crucial roles in regulating the excitability of GCs. Given their importance in enforcing sparse coding, these interneurons could be key modulators of the SB–IB activity differences observed in our study, yet their precise contribution to blade-specific activation remains unclear.

Consistent with prior behavioral work implicating the DG in pattern separation⁵⁴, manipulations that ablate abDGCs impair discrimination when task difficulty increases⁵⁵, whereas genetic or environmental manipulations that enhance neurogenesis improve discrimination performance^{56,57}. In our study, only limited numbers of abDGCs were labeled during task performance, as expected given their low population size relative to mGCs. However, neurons that

were approximately seven weeks post-mitosis were more prominently labeled than those four weeks post-mitosis, consistent with the notion that older abDGCs are more integrated into the local circuit. Despite their small number, these neurons exerted a disproportionate influence on DG activity.

Using an acute, reversible chemogenetic inhibition of different abDGC cohorts during task performance³⁹, we observed that suppression of ≤ 7 -week-old abDGCs increased mGC activity, consistent with prior studies that chronically reduced neurogenesis³⁵. More importantly, inhibition of abDGCs during the LS task impaired performance in high-demand discrimination trials without affecting already-learned, low-demand trials within the same session. This manipulation disrupted the normal SB-biased pattern of mGC activation seen in controls, as well as the typical distribution of labeled mGCs along the hilar–molecular layer and apex–tip axes of the DG. These effects were specific to the inhibition of ≤ 7 -week-old abDGCs and were not observed when younger (≤ 4 -week-old) or older abDGC populations were targeted. These findings suggest that abDGCs exert their influence not through direct encoding, but through regulating inhibitory tone within the DG microcircuit, potentially via PV- or SOM-positive interneurons that shape granule cell recruitment.

Notably, our results indicate that behavioral performance is not determined solely by the magnitude of dentate gyrus activity. Inhibition of abDGCs increased overall mGC activation while disrupting the spatially organized, blade-biased pattern of activity, whereas direct inhibition of mGCs reduced overall activity without necessarily disrupting spatial organization in animals that remained capable of performing the task. These findings suggest that dentate gyrus function depends on the coordinated recruitment and composition of granule cell ensembles rather than on total activity levels alone. Distinct perturbations may therefore impair behavior by altering the identity, balance, and coordination of neurons participating in task-relevant ensembles.

Together, these results reveal complementary but distinct contributions of mGCs and abDGCs to DG function. mGCs provide the essential excitatory output required for encoding and task performance under high cognitive demand, whereas abDGCs modulate this activity to establish the structured, blade-biased distribution of mGC activation necessary for optimal discrimination. This modulation likely arises through transient inhibitory control during the critical period of abDGC maturation, allowing the DG to dynamically tune its excitability and spatial coding to match task difficulty. Our findings support a model in which efficient pattern separation relies on both the stable excitatory scaffold provided by mGCs and the adaptive, modulatory influence of maturing abDGCs on DG network organization. This aligns with and extends prior models by showing that it is the maturing cohort, not immature or fully mature abDGCs—that provides this modulatory control.

Limitations of the Study

The use of activity tagging has the inherent limitation that it provides a snapshot of activity integrated across a temporal window and therefore cannot resolve which specific trial types, behavioral epochs, or decision processes drive neuronal recruitment. This is countered by the less invasive nature of these experiments than ones that require implants for *in vivo* recording of neuron activity, which are themselves not fit for purpose because they cannot capture the spatial distribution in the subregions of the DG effectively. Future studies using *in vivo* measurements of activity will be required to support the present findings and find conclusive neural representations required for discrimination of similar patterns.

Additionally, the experimental design in DREADD experiments required extended duration of tamoxifen administration which resulted in broad labeling of adult-born granule cells spanning multiple stages of maturation (approximately 0–4 weeks and 0–7 weeks old), rather than a precisely birth-dated population. Consequently, the chemogenetic manipulations reflect the combined influence of a heterogeneous cohort of immature neurons, and do not resolve more finely the age-specific contributions of adult-born granule cells. Finally, the circuit mechanisms

underlying blade-biased activity remain unknown, and future studies will be required to determine how these spatial patterns arise and how they contribute to computations within the dentate gyrus.

Materials and methods

Ethics statement

All animal procedures were conducted in accordance with protocols approved by the Northwestern University Institutional Animal Care and Use Committee (IACUC). Male and female mice were housed 3–5 per cage on a reverse light/dark cycle of 12/12 hours within a controlled environment. They received food and water *ad libitum*. All behavioral testing was conducted during the animals' dark cycle and was conducted by experimenters blinded to genotype. Tail biopsies were collected for genotyping via PCR. All efforts were made to minimize the number of mice used and to reduce any potential pain or stress, in line with ethical guideline.

Animals

All mouse lines were maintained on a C57Bl/6 background. Mice were propagated by female breeders heterozygous for Cre recombinase and homozygous for either the Ai9 allele (Rosa-CAG-LSL-tdTomato-WPRE) or hM4Di allele (R26-LSL-Gi-DREADD) with male breeders homozygous for Ai9 or hM4Di. TRAP2;Ai9 mice were generated by crossing Fos^{2A-iCreER/+} (TRAP2) mice (Jax #030323)²³ with Ai9 (Jax #007909) mice to generate double-heterozygous mice. Ascl1-CreER^{T2};hM4Di mice were generated by crossing R26-LSL-Gi-DREADD mice (Jax #026219)³⁷ with mice carrying tamoxifen-inducible Cre recombinase under the control of the *Ascl1* promoter (Jax #:012882)⁵⁸ to generate double-transgenic progeny Ascl1-CreER^{T2};R26^{LSL}-hM4Di. Cre-negative littermates from heterozygote breedings were used as controls. Dock10-Cre;hM4Di mice were generated by crossing Dock10-Cre³³ mice with hM4Di mice to produce double-transgenic progeny Dock10-Cre;R26^{LSL}-hM4Di.

Birth-dating of abDGCs

6–8-week-old mice received three intraperitoneal (i.p.) injections of 5-Bromo-2'-deoxyuridine (BrdU, Sigma Aldrich #B9285) at 100 mg/kg (i.p.) at 4 hours intervals on a single day⁵⁹. Two weeks later, 5-Ethynyl-2'-deoxyuridine (EdU, Sigma Aldrich, #900584) was administered following the same injection protocol. EdU was used as an additional thymidine analog to label a distinct cohort of dividing cells. Because BrdU and EdU are incorporated into DNA during S-phase following systemic administration, the birth-dating timeline and survival intervals applied to BrdU labeling also apply to EdU labeling.

In this study, mature granule cells (mGCs) are operationally defined as dentate granule cells that were not labeled with BrdU or EdU and therefore are not classified as adult-born neurons within the defined labeling window. While the majority of these cells are expected to represent developmentally generated mature neurons, we cannot exclude the possibility that a small fraction may include younger, unlabeled neurons generated outside the labeling period.

Tamoxifen administration

IP injections: To time the labeling of TRAP+ cell in TRAP2 mice, 4-hydroxytamoxifen (4-OHT; Sigma Aldrich, #H7904) was administered intraperitoneally. 4-OHT was dissolved at 20 mg/mL in pure ethanol and stored at –20°C for up to several weeks. Before use, 4-OHT was redissolved in corn oil (Sigma, Cat #C8267) and heated to 90°C with shaking until the ethanol had evaporated yielding a final concentration of 10 mg/mL. The final 10 mg/mL 4-OHT solutions was used on the day it was prepared. All injections were delivered intraperitoneally at 50 mg/kg either at the end of the training phase of the day or 2 hours after running.

Oral administration: For conditional expression of hM4Di in Ascl1-CreER^{T2} cells, a tamoxifen diet (TD.130858, Envigo) was provided *ad libitum* for 4 or 7 weeks, depending on the experimental timeline. Body weight was carefully monitored daily. The chow contained 500 mg tamoxifen/kg,

delivering ~80 mg tamoxifen/kg body weight per day, assuming an average mouse weight of 20-25 g and a daily intake of 3-4 g.

Mice were monitored daily for signs of stress, discomfort, or adverse effects related to tamoxifen exposure.

Inhibitory DREADD-hM4Di activation

Deschloroclozapine (DCZ, MedChemExpress #HY-42110) was dissolved in 2% DMSO in 0.9% saline and administered by i.p. injections at a dose of 50 $\mu\text{g.kg}^{-1}$ ³⁴. Behavioral trials started 30 minutes after drug administration.

Behavioral procedures

Adult (8-10 weeks-old) male and female mice were used in behavioral studies. To acclimate the mice to handling before behavioral testing, all animals were handled by the experimenter for 2 min per day for 3 consecutive days.

Running wheels

To habituate the mice to running, wireless running wheels (ENV-047, Med Associates) were placed in their home cage for 5 days (maximum 5 mice per cage). Two days prior to BrdU injections, the wheels were removed from the cage. One week before the experiment, mice were individually housed. On the day of the experiment, a running wheel was added to each cage for a duration of 4 hours. After 2 hours, mice received a 4-OHT injection and were returned to their respective cages. Following an additional 2 hours of running, the wheels were removed from the cages. Three days later, wheels were reintroduced to the cage and remained until mice were euthanized for assessment of TRAP+ cells, 90 minutes after the beginning of the run. The effect of exposure to the wheels was quantified by measuring the total distance run by each mouse.

Automated touch screen operant behaviors

Testing was conducted in custom-built, touch screen-based automated operant device boxes designed specifically for mice, the Operant House (doi.org/10.1101/2025.01.24.634815 ²⁹). The apparatus consisted of a rectangular modular testing chamber (w14.5cm x l21.5cm x h18.5cm). A 60mW green LED was located above a port that gave access to a retractable water bottle at the back of the chamber. At the opposite end of the chamber, a flat-screen infrared touch monitor was controlled by custom scripts (Python). In front of the touchscreen was a mask with 2 horizontal lines of 5 squares. Mice performed a modified version of the “trial unique nonmatching-to-location” (TUNL) task ²⁹. Five days prior to testing and during the training phase, mice were placed on a water-restriction regime, with access to water for 10 minutes per day, and were maintained at 85-90% body weight during the entire duration of the experiment. Mice were trained each day, in two separate sessions of 30 trials or a maximum of 30 minutes.

Training phases – Shaping : Following 30 minutes of habituation to the operant chamber, mice were trained to touch a lit white square stimulus presented randomly in one of the response windows of the touch screen in order to receive a reward (2 seconds water nozzle access with saccharin water 0.01%). Mice were trained in Shaping until they successfully achieved 30 rewards within 30 min during 3 successive sessions.

Training phases – Large-only separation (low cognitive demand): After shaping, mice were trained in the TUNL task with a Large-only separation (L configuration). For this task, each trial was composed of two phases. During the sample phase, a white square was illuminated in one of the 10 possible locations on the screen ([Figure 1A](#) ²⁹, [S2A & B](#) ²⁹). Following a nose-poke to the sample square the stimulus disappeared, and the mouse received a reward only on 33% of the trials. After a 1s delay from the sample trial, the choice trial was initiated where two stimuli were presented one in the repeated sample location (incorrect) and the other in the new location (correct) separated spatially by 4 squares in the L configuration ([Figure 1A](#) ²⁹, [S2A & B](#) ²⁹). A touch to the correct sample location resulted in a reward and an 5s inter-trial interval (ITI) before the next trial

was initiated. An incorrect choice resulted in a 10 second punishment (roof lights ON) and then an ITI, followed by correction trials during the first two days of training. Correction trials followed the same procedures, except that the same sample and choice locations from the previous incorrect trial were presented again until the correct choice was made. The correction trials were not included in performance calculations. Mice progressed to the next training phase after completing ≥ 30 trials within 30 minutes and achieving $>70\%$ correct responses in at least two of four sessions across two consecutive days.

Training phases – Large + Small separation (high cognitive demand): Mice were then trained in sessions containing both large-separation (L) and small-separation (S) trials (LS configuration). Training followed the same pattern as previously described except that there were two separate possibilities for the presentation of the choice square which was either separated by 4 squares from the sample square during L trials, or the choice square was 0-1 square separated from the sample location during S trials (Figure 1A & S2B).

Each session consisted of 15 L and 15 S trials presented in alternating order. Mice were trained until reaching criterion defined as $\geq 70\%$ correct responses on S trials in two of four sessions across two consecutive days.

Approximately 25–30% of mice across experimental groups did not reach LS criterion within the allotted training period (Figure S2G).

Immunohistochemistry

Mice were deeply anesthetized with isoflurane and then perfused transcardially with PBS containing 0.02% sodium nitrite followed by 2% paraformaldehyde (PFA) in 0.1 M sodium acetate buffer (pH 6.5) for 3 min, and 2% PFA in 0.1 M sodium borate buffer (pH 8.5) for 10 min⁶⁰. Brains were dissected and post-fixed in 2% PFA 0.1 M sodium borate buffer (pH 8.5), for 24–48 hours and stored at 4°C in PBS until sectioning. 50 μ m floating sections were collected on a Leica Vibratome VT1000 and stored at 4°C in PBS containing 0.02% sodium azide. For c-Fos analysis, perfusions were performed 90 minutes after the end of the behavioral training. Immunohistochemistry was conducted at room temperature using standard procedures. Free-floating sections were incubated for 30 min in a solution of 25 mM glycine to decrease the background signal and then rinsed two times in Tris-Buffered Saline 1X (TBS). Sections were then blocked for 1 hour in the saturation solution containing TBS-Triton (0.25% Triton X-100 (Fisher, BP151-100), 3% normal donkey serum (NDS; Jackson ImmunoResearch, 017-000-121) and 0.1% bovine serum albumin (BSA; Sigma, A9085)). The sections were then incubated overnight in the saturation solution containing the primary antibody (rabbit anti c-Fos (1/1500, Cell Signaling), sheep anti-BrdU (1/2500, NovusBio), rabbit anti RFP (1/5000, Abcam), rabbit anti HA-Tag (1:500, #3724, Cell Signaling)). For c-Fos staining, no serum and BSA were used. For BrdU staining, sections were incubated for 10 mins in a solution of DNase I (Sigma #D5025, 15 KU/mL dissolved in 0.05 M NaCl), MgCl₂ (6 mM) and CaCl₂ (10 mM) to cleave DNA then rinsed 2 x 5 min in TBS before the saturation part. For EdU staining, sections were incubated in solution from the kit ClickITTM (#C10340 ThermoFisher) during 30 min then rinsed 2 x 5 min TBS before the saturation part. No antibodies were necessary for the rest of the immunostaining. The next day, sections were washed 2x5 min in TBS before being incubated for 1 hour in secondary antibody solution (donkey anti-rabbit Alexa 647 or 488 (1/1000, Invitrogen), donkey anti-sheep Alexa 488 (1/1000, Invitrogen)). Finally, sections were washed 2x5 min in TBS and cover-slipped with ProLong Diamond mounting media 275 containing DAPI (#P36962, ThermoFisher Scientific).

Imaging and neuron quantification

Analyses were conducted in the dorsal DG, as prior studies have demonstrated its involvement in spatial memory and the fine-grained contextual differentiation required for pattern separation. The dorsal hippocampus is crucial for encoding and distinguishing spatial information, which is central to this process. In contrast, the ventral DG is more strongly associated with emotional processing and affective memory rather than high-resolution spatial encoding needed for pattern separation.

Confocal images were obtained with a Zeiss microscope (LSM700) with a 20x objective with the experimenter blind to the genotype and condition following previous protocols⁵⁹. One section in every ten serial sections were analyzed through the dorsal hippocampus (stereotaxic coordinates: -1.20 to -2.30 relative to bregma according to Paxinos' brain atlas). Two to three 10 μm z-stack acquisitions, were made to picture the whole dentate gyrus in each section. 3D reconstructions were used to univocally verify co-localization (e.g. activated mature neurons or immature neurons). All acquisitions were carried out in sequential scanning mode to prevent cross-bleeding between channels. Stacks of images were then reconstituted using Fiji software, allowing the experimenter to count the number of cells. The surface area of the dorsal hippocampus, DG, SB, IB and CA3 were traced in mapping software (NeuroLucida Zeiss) using a 10x objective. The volume was determined by multiplying the surface area by the distance between sections (500 μm). Densities of BrdU+, EdU+, TRAP+ and c-Fos+ cells were estimated by multiplying the total number of labeled cells by 10. For immunohistochemical analysis, a subset of approximately 5 mice from the behavioral cohorts of ~15 mice were randomly selected. If the results exhibited high variability, one or two additional mice were included to improve data precision. Selection remained random throughout. This approach was chosen to balance the workload associated with immunohistochemistry while ensuring sufficient statistical power; given that behavioral experiments require larger sample sizes to achieve robust significance only a subset could be included in this analysis.

Distribution of TRAP+ and c-Fos+ Cells in the DG

We followed methods presented in literature⁶¹. Using stacks of images obtained from confocal microscopy, the localization of TRAP+ and c-Fos+ cells in the DG was assessed using Fiji software. *Distance from the subgranular zone (SGZ)*: The contours of each blade were manually outlined and the distance from the center of each cell body to the SGZ was manually measured. Cells located in the apex region were excluded from the counting.

Distance from the apex: Excluding the apex region, the size of each suprapyramidal blade was measured as well as the distance of each cell from the apex. These distances were then expressed as percentage of total blade length.

Statistical analysis

Data are expressed as mean \pm SEM. Statistical analyses were performed using Statview and Origin. Histochemical and other experimental data were assessed for normality using the Shapiro-Wilk test. For normally distributed datasets, comparisons between two groups were performed using unpaired t-tests, while one-way or two-way ANOVAs were used for experiments with multiple factors, followed by appropriate post hoc tests (Tukey or Bonferroni) when significant. For datasets that deviated from normality, non-parametric tests (Mann-Whitney U test or Kruskal-Wallis test) were applied, with Dunn's post hoc test and Bonferroni correction as appropriate. Differences were considered significant at $p < 0.05$.

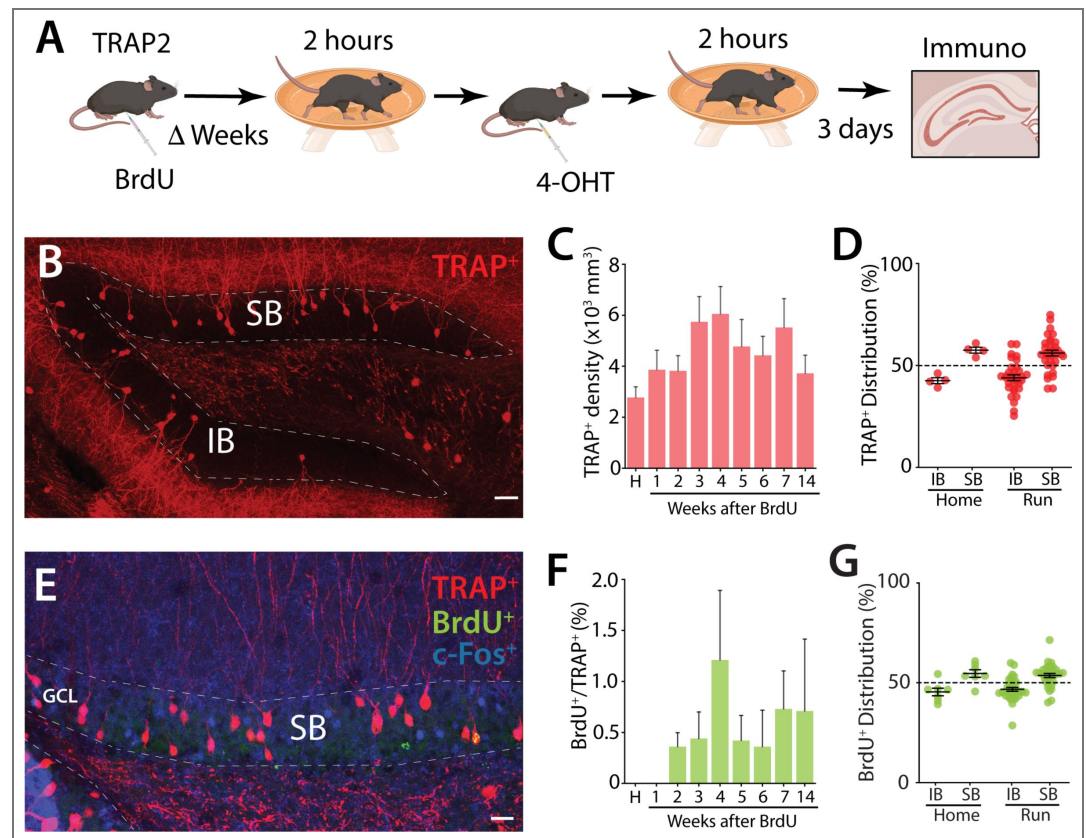


Figure Sup 1. Activity labeling of GCs after running. (A) Schematic representation of the running paradigm. Prior to the behavioral experiment, mice (males $n = 30$, females $n = 3$) were injected with BrdU. After Δ weeks, the mice were allowed to run for 2 hours before receiving a 4-OHT injection, followed by an additional 2 hours of running. Mice were perfused 3 days later for immunohistochemistry of brain sections. (B) Example section of immunohistochemistry of TRAP⁺ cells in the dorsal DG illustrating the distinct blades of the DG: SB and IB. Calibration bar: 40 μ m. (C) Density of TRAP⁺ GCs in mice after running compared to home cage controls (H). Each group represents mice that ran after variable time intervals following BrdU injections. (D) Blade specific distribution of TRAP⁺ cells in the infrapyramidal (IB) and suprapyramidal (SB) blades of the dorsal DG in TRAP2 mice that ran for 4 hours, compared to home cage mice. (IB: home: 42.5 ± 1.43 %, 4 mice; run: 43.9 ± 1.42 %, 33 mice; ns; Mann-Whitney). (E) Example section from mice labeled for BrdU (green), TRAP⁺ GCs (red) and endogenous c-Fos (blue) in TRAP2 mice. Calibration bar: 20 μ m. (F) Percentage of BrdU⁺ cells (Δ 1-14 weeks old) also labeled with TRAP in the dorsal DG of TRAP2 mice that ran for 4 hours, compared to home cage controls (H). (G) Blade specific distribution of BrdU⁺ cells in the infrapyramidal (IB) and suprapyramidal (SB) blades of the dorsal DG in TRAP2 mice that ran for 4 hours, compared to home cage mice.

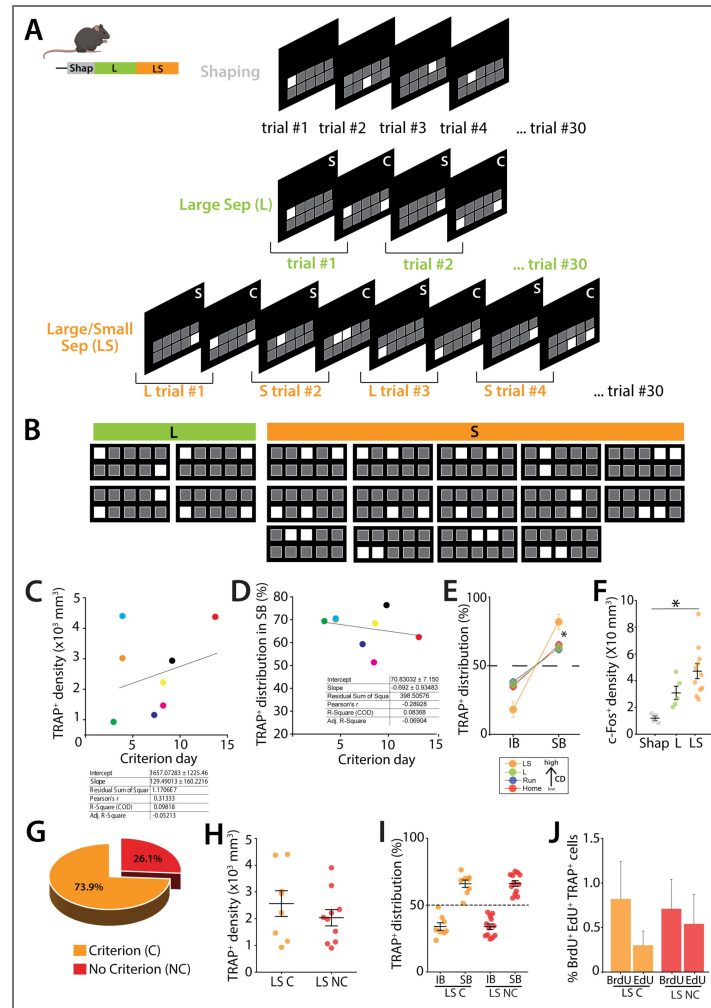


Figure Sup 2. Description of the TUNL protocol and analysis of TRAP+ GCs in mice not meeting the criterion in the pattern separation task.

(A) Overview of successive trials in the Shaping, L and LS steps. Mice must complete 30 trials at each step. The shaping phase involves touching the illuminated square to receive a reward. In the L and LS steps, there are in two phases: the sample (S) phase, where mice must touch the single illuminated square on the panel and the choice test (C), where two squares are illuminated: the previously touched square and a new one. Mice must nose poke the new square to receive the reward. In the L phase, trials consist only of L panels only (see Figure S2B – L). In the LS phase, L and S trials are alternated (see Figure S2B for S panels). (B) Recommended stimulus pairs for the TUNL protocol. Large pairs (green) consist of 2 illuminated squares separated by 4 squares that are turned OFF. Small pairs (orange) consist of two illuminated squares separated by 0 to 1 square turned OFF. (C) Correlation between the performance (= number of days to reach criterion) and the TRAP+ density or TRAP+ distribution in SB (D). Both regression analysis (right tables) show that the slope is not significantly different from zero. (E) Comparison of TRAP+ distribution in IB and SB of the dorsal DG in mice from the home cage (Home), running (Run), large configuration (L) and large and small separation (LS) groups. In the control groups (Home, Run and L) the TRAP+ cells are predominantly distributed in the SB, with a slight preference for the SB over IB. However, in the high cognitive demand (CD), LS task the distribution of TRAP+ cells become more balanced, with a marked preference for the SB (SB >>IB), indicating task-specific modulation of activity across the blades. (F) Density of c-Fos-labeled GCs in mice undergoing LS compared to control mice engaged in either shaping (Shap) or the Large configuration task only (L) (one-way ANOVA, $F(2,17) = 8.05$, $p = 0.0035$; Tukey post-hoc test. Groups Shap vs LS were significantly different). (G) Percentage of TRAP2 mice reaching the criterion (C) compared to those that did not within 14 days of training (NC). (H) Density of TRAP+ neurons in mice undergoing LS for mice that reached criterion (LS C) with mice that did not reach 70% criterion within 14 days (LS NC) (Unpaired t-test, $t(16) = 0.96$, $p = 0.35$). (I) Blade specific distribution of TRAP+ GCs in the SB and the IB of the dorsal DG (IB: LS C: $34 \pm 2.79\%$, 8 mice; LS NC: $33.8 \pm 2.12\%$, 12 mice; ns; Mann-Whitney). (J) Percentage of TRAP+ cells also birth-dated with BrdU (39 days \pm 1.23) or Edu cells (25 days \pm 1.37 days) in the dorsal DG of TRAP2 mice that reached criterion (C) and those failed (NC) (BrdU: LS C: $0.71 \pm 0.333\%$, 7 mice; LS NC: $0.822 \pm 0.45\%$, 6 mice; ns; Edu: LS C: $0.542 \pm 0.33\%$, 9 mice; LS NC: $0.304 \pm 0.16\%$, 7 mice; ns; Mann-Whitney).

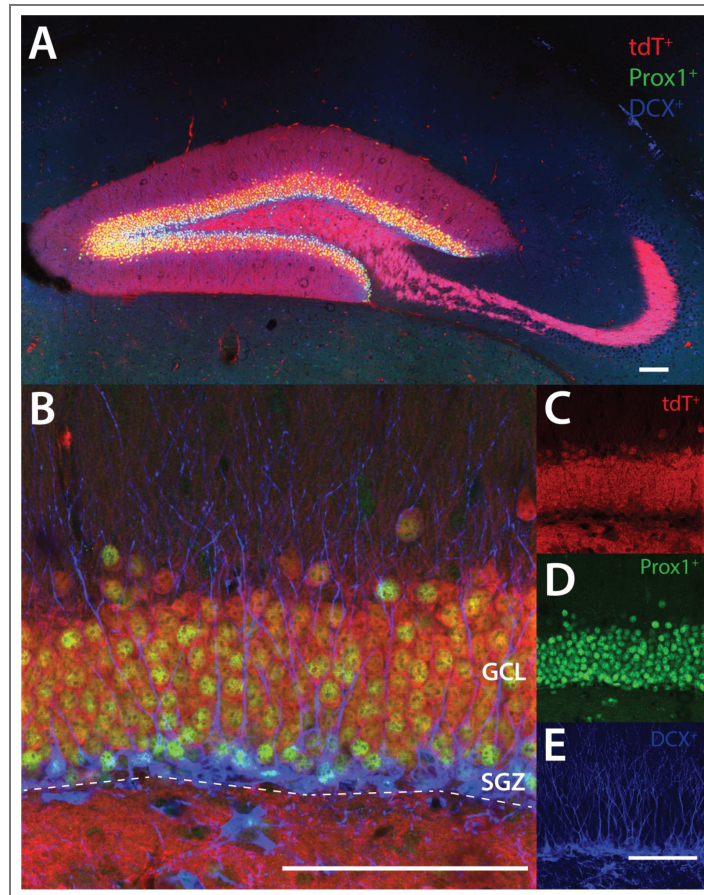


Figure Sup 3. Dock10 is not expressed in -positive immature newborn neurons but is expressed in Prox1-positive neurons.

(A) Representative section from a Dock10-cre: Ai9 mouse following tdTomato (tdT), Prox1, and DCX immunohistochemistry in the dorsal DG. Calibration bar: 100 μ m. (B) 20X magnification of the suprapyramidal layer of the dentate gyrus, showing tdTomato (Dock10) in red, Prox1 in green, and DCX in blue. Calibration bar: 100 μ m. (C) tdT (Dock10) labeling. (D) Prox1 (green) labeling. (E) DCX (blue) labeling. Calibration bar: 100 μ m.

Data availability

All datasets for immunohistochemistry will be made available by depositing into a suitable platform.

Acknowledgements

We thank members of the Contractor lab for helpful discussion and Ms. Astrid Castellanos for assistance during the performance of this study. This work was supported by NIH/NINDS 5R01NS115471 and NIH/NIMH R01MH130428 to AC.

Additional information

Author Contributions

C.C.M.C., J.N.A., and A.C. designed experiments. C.C.M.C. performed all behavioral testing, immunohistochemistry, imaging and analysis. S.O. developed the touch screen automated task and wrote the code to operate the procedures. C.C.M.C. and A.C. prepared the manuscript. All authors approved the final version.

Funding

Funder	Grant reference number	Author
HHS National Institutes of Health (NIH)	R01NS115471	Anis Contractor
HHS National Institutes of Health (NIH)	R01MH130428	Anis Contractor

Author ORCID iDs

Charlotte CM Castillon: <https://orcid.org/0009-0001-5688-1893>

Anis Contractor: <https://orcid.org/0000-0002-5131-2536>

References

1. Hainmueller T., Bartos M (2020) Dentate gyrus circuits for encoding, retrieval and discrimination of episodic memories. *Nat Rev Neurosci* **21**:153-168 <https://doi.org/10.1038/s41583-019-0260-z> | [PubMed](#)
2. Stefanini F., Kushnir L., Jimenez J.C., Jennings J.H., Woods N.I., Stuber G.D., Kheirbek M.A., Hen R., Fusi S (2020) A Distributed Neural Code in the Dentate Gyrus and in CA1. *Neuron* **107**:703-716.e704. <https://doi.org/10.1016/j.neuron.2020.05.022> | [PubMed](#)
3. Hunsaker M.R., Rosenberg J.S., Kesner R.P (2008) The role of the dentate gyrus, CA3a,b, and CA3c for detecting spatial and environmental novelty. *Hippocampus* **18**:1064-1073 <https://doi.org/10.1002/hipo.20464> | [PubMed](#)
4. Leutgeb J.K., Leutgeb S., Moser M.B., Moser E.I (2007) Pattern separation in the dentate gyrus and CA3 of the hippocampus. *Science* **315**:961-966 <https://doi.org/10.1126/science.1135801> | [PubMed](#)
5. Marr D (1969) A theory of cerebellar cortex. *J Physiol* **202**:437-470 <https://doi.org/10.1113/jphysiol.1969.sp008820> | [PubMed](#)
6. Marr D (1971) Simple memory: a theory for archicortex. *Philos Trans R Soc Lond B Biol Sci* **262**:23-81 <https://doi.org/10.1098/rstb.1971.0078> | [PubMed](#)
7. Rolls E.T (2016) Pattern separation, completion, and categorisation in the hippocampus and neocortex. *Neurobiol Learn Mem* **129**:4-28 <https://doi.org/10.1016/j.nlm.2015.07.008> | [PubMed](#)
8. Borzello M., Ramirez S., Treves A., Lee I., Scharfman H., Stark C., Knierim J.J., Rangel L.M (2023) Assessments of dentate gyrus function: discoveries and debates. *Nat Rev Neurosci* **24**:502-517 <https://doi.org/10.1038/s41583-023-00710-z> | [PubMed](#)

9. **Amaral D.G.**, Scharfman H.E., Lavenex P (2007) The dentate gyrus: fundamental neuroanatomical organization (dentate gyrus for dummies). *Prog Brain Res* **163**:3-22 [https://doi.org/10.1016/S0079-6123\(07\)63001-5](https://doi.org/10.1016/S0079-6123(07)63001-5) | PubMed
10. **Chawla M.K.**, Guzowski J.F., Ramirez-Amaya V., Lipa P., Hoffman K.L., Marriott L.K., Worley P.F., McNaughton B.L., Barnes C.A (2005) Sparse, environmentally selective expression of Arc RNA in the upper blade of the rodent fascia dentata by brief spatial experience. *Hippocampus* **15**:579-586 <https://doi.org/10.1002/hipo.20091> | PubMed
11. **Erwin S.R.**, Sun W., Copeland M., Lindo S., Spruston N., Cembrowski M.S (2020) A Sparse, Spatially Biased Subtype of Mature Granule Cell Dominates Recruitment in Hippocampal-Associated Behaviors. *Cell Rep* **31**:107551 <https://doi.org/10.1016/j.celrep.2020.107551> | PubMed
12. **Snyder J.S.**, Radik R., Wojtowicz J.M., Cameron H.A (2009) Anatomical gradients of adult neurogenesis and activity: young neurons in the ventral dentate gyrus are activated by water maze training. *Hippocampus* **19**:360-370 <https://doi.org/10.1002/hipo.20525> | PubMed
13. **Scharfman H.E.**, Sollas A.L., Smith K.L., Jackson M.B., Goodman J.H (2002) Structural and functional asymmetry in the normal and epileptic rat dentate gyrus. *J Comp Neurol* **454**:424-439 <https://doi.org/10.1002/cne.10449> | PubMed
14. **Altman J.**, Das G.D (1965) Autoradiographic and histological evidence of postnatal hippocampal neurogenesis in rats. *J Comp Neurol* **124**:319-335 <https://doi.org/10.1002/cne.901240303> | PubMed
15. **Kaplan M.S.**, Hinds J.W (1977) Neurogenesis in the adult rat: electron microscopic analysis of light radioautographs. *Science* **197**:1092-1094 <https://doi.org/10.1126/science.887941> | PubMed
16. **Cameron H.A.**, Woolley C.S., McEwen B.S., Gould E (1993) Differentiation of newly born neurons and glia in the dentate gyrus of the adult rat. *Neuroscience* **56**:337-344 [https://doi.org/10.1016/0306-4522\(93\)90335-d](https://doi.org/10.1016/0306-4522(93)90335-d) | PubMed
17. **Kempermann G.**, Kuhn H.G., Gage F.H (1997) More hippocampal neurons in adult mice living in an enriched environment. *Nature* **386**:493-495 <https://doi.org/10.1038/386493a0> | PubMed
18. **Folsz O.**, Trouche S., Croset V (2023) Adult-born neurons add flexibility to hippocampal memories. *Front Neurosci* **17**:1128623 <https://doi.org/10.3389/fnins.2023.1128623> | PubMed
19. **Dranovsky A.**, Picchini A.M., Moadel T., Sisti A.C., Yamada A., Kimura S., Leonardo E.D., Hen R (2011) Experience dictates stem cell fate in the adult hippocampus. *Neuron* **70**:908-923 <https://doi.org/10.1016/j.neuron.2011.05.022> | PubMed
20. **Jinno S** (2011) Topographic differences in adult neurogenesis in the mouse hippocampus: a stereology-based study using endogenous markers. *Hippocampus* **21**:467-480 <https://doi.org/10.1002/hipo.20762> | PubMed
21. **Ramirez-Amaya V.**, Marrone D.F., Gage F.H., Worley P.F., Barnes C.A (2006) Integration of new neurons into functional neural networks. *J Neurosci* **26**:12237-12241 <https://doi.org/10.1523/JNEUROSCI.2195-06.2006> | PubMed
22. **Alves N.D.**, Patricio P., Correia J.S., Mateus-Pinheiro A., Machado-Santos A.R., Loureiro-Campos E., Morais M., Bessa J.M., Sousa N., Pinto L (2018) Chronic stress targets adult neurogenesis preferentially in the suprapyramidal blade of the rat dorsal dentate gyrus. *Brain Struct Funct* **223**:415-428 <https://doi.org/10.1007/s00429-017-1490-3> | PubMed
23. **DeNardo L.A.**, Liu C.D., Allen W.E., Adams E.L., Friedmann D., Fu L., Guenther C.J., Tessier-Lavigne M., Luo L (2019) Temporal evolution of cortical ensembles promoting remote memory retrieval. *Nat Neurosci* **22**:460-469 <https://doi.org/10.1038/s41593-018-0318-7> | PubMed
24. **Castillon C.**, Lunion S., Desvignes N., Hanauer A., Laroche S., Poirier R (2018) Selective alteration of adult hippocampal neurogenesis and impaired spatial pattern separation performance in the RSK2-deficient mouse model of Coffin-Lowry syndrome. *Neurobiol Dis* **115**:69-81 <https://doi.org/10.1016/j.nbd.2018.04.007> | PubMed

25. Chatzi C., Zhang Y., Hendricks W.D., Chen Y., Schnell E., Goodman R.H., Westbrook G.L (2019) Exercise-induced enhancement of synaptic function triggered by the inverse BAR protein, Mtss1L. *eLife* **8** <https://doi.org/10.7554/eLife.45920> | [PubMed](#)
26. Deng W., Mayford M., Gage F.H (2013) Selection of distinct populations of dentate granule cells in response to inputs as a mechanism for pattern separation in mice. *eLife* **2**:e00312 <https://doi.org/10.7554/eLife.00312> | [PubMed](#)
27. Lamothe-Molina P.J., Franzelin A., Beck L., Li D., Auksutat L., Fieblinger T., Laprell L., Alhbeck J., Gee C.E., Kneussel M., *et al.* (2022) DeltaFosB accumulation in hippocampal granule cells drives cFos pattern separation during spatial learning. *Nat Commun* **13**:6376 <https://doi.org/10.1038/s41467-022-33947-w> | [PubMed](#)
28. Talpos J.C., McTighe S.M., Dias R., Saksida L.M., Bussey T.J (2010) Trial-unique, delayed nonmatching-to-location (TUNL): a novel, highly hippocampus-dependent automated touchscreen test of location memory and pattern separation. *Neurobiol Learn Mem* **94**:341-352 <https://doi.org/10.1016/j.nlm.2010.07.006> | [PubMed](#)
29. Oomen C.A., Hvoslef-Eide M., Heath C.J., Mar A.C., Horner A.E., Bussey T.J., Saksida L.M (2013) The touchscreen operant platform for testing working memory and pattern separation in rats and mice. *Nat Protoc* **8**:2006-2021 <https://doi.org/10.1038/nprot.2013.124> | [PubMed](#)
30. Williams P.A., Larimer P., Gao Y., Strowbridge B.W (2007) Semilunar granule cells: glutamatergic neurons in the rat dentate gyrus with axon collaterals in the inner molecular layer. *J Neurosci* **27**:13756-13761 <https://doi.org/10.1523/JNEUROSCI.4053-07.2007> | [PubMed](#)
31. Pilz G.A., Carta S., Stauble A., Ayaz A., Jessberger S., Helmchen F (2016) Functional Imaging of Dentate Granule Cells in the Adult Mouse Hippocampus. *J Neurosci* **36**:7407-7414 <https://doi.org/10.1523/JNEUROSCI.3065-15.2016> | [PubMed](#)
32. Danielson N.B., Kaifosh P., Zaremba J.D., Lovett-Barron M., Tsai J., Denny C.A., Balough E.M., Goldberg A.R., Drew L.J., Hen R., *et al.* (2016) Distinct Contribution of Adult-Born Hippocampal Granule Cells to Context Encoding. *Neuron* **90**:101-112 <https://doi.org/10.1016/j.neuron.2016.02.019> | [PubMed](#)
33. Kohara K., Pignatelli M., Rivest A.J., Jung H.Y., Kitamura T., Suh J., Frank D., Kajikawa K., Mise N., Obata Y., *et al.* (2014) Cell type-specific genetic and optogenetic tools reveal hippocampal CA2 circuits. *Nat Neurosci* **17**:269-279 <https://doi.org/10.1038/nn.3614> | [PubMed](#)
34. Nagai Y., Miyakawa N., Takuwa H., Hori Y., Oyama K., Ji B., Takahashi M., Huang X.P., Slocum S.T., DiBerto J.F., *et al.* (2020) Deschloroclozapine, a potent and selective chemogenetic actuator enables rapid neuronal and behavioral modulations in mice and monkeys. *Nat Neurosci* **23**:1157-1167 <https://doi.org/10.1038/s41593-020-0661-3> | [PubMed](#)
35. Ikrar T., Guo N., He K., Besnard A., Levinson S., Hill A., Lee H.K., Hen R., Xu X., Sahay A (2013) Adult neurogenesis modifies excitability of the dentate gyrus. *Front Neural Circuits* **7** <https://doi.org/10.3389/fncir.2013.00204> | [PubMed](#)
36. Drew L.J., Kheirbek M.A., Luna V.M., Denny C.A., Cloidt M.A., Wu M.V., Jain S., Scharfman H.E., Hen R (2016) Activation of local inhibitory circuits in the dentate gyrus by adult-born neurons. *Hippocampus* **26**:763-778 <https://doi.org/10.1002/hipo.22557> | [PubMed](#)
37. Zhu H., Aryal D.K., Olsen R.H., Urban D.J., Swearingen A., Forbes S., Roth B.L., Hochgeschwender U (2016) Cre-dependent DREADD (Designer Receptors Exclusively Activated by Designer Drugs) mice. *Genesis* **54**:439-446 <https://doi.org/10.1002/dvg.22949> | [PubMed](#)
38. Yang S.M., Alvarez D.D., Schinder A.F (2015) Reliable Genetic Labeling of Adult-Born Dentate Granule Cells Using Ascl1 CreERT2 and Glaxt CreERT2 Murine Lines. *J Neurosci* **35**:15379-15390 <https://doi.org/10.1523/JNEUROSCI.2345-15.2015> | [PubMed](#)
39. Tunc-Ozcan E., Peng C.Y., Zhu Y., Dunlop S.R., Contractor A., Kessler J.A (2019) Activating newborn neurons suppresses depression and anxiety-like behaviors. *Nat Commun* **10**:3768 <https://doi.org/10.1038/s41467-019-11641-8> | [PubMed](#)

40. Lacefield C.O., Itskov V., Reardon T., Hen R., Gordon J.A. (2012) Effects of adult-generated granule cells on coordinated network activity in the dentate gyrus. *Hippocampus* **22**:106-116 <https://doi.org/10.1002/hipo.20860> | PubMed
41. Ge S., Yang C.H., Hsu K.S., Ming G.L., Song H. (2007) A critical period for enhanced synaptic plasticity in newly generated neurons of the adult brain. *Neuron* **54**:559-566 <https://doi.org/10.1016/j.neuron.2007.05.002> | PubMed
42. Kennedy W.M., Gonzalez J.C., Lee H., Wadiche J.I., Overstreet-Wadiche L. (2024) T-Type Ca(2+) Channels Mediate a Critical Period of Plasticity in Adult-Born Granule Cells. *J Neurosci* **44** <https://doi.org/10.1523/JNEUROSCI.1503-23.2024> | PubMed
43. Guzman S.J., Schlogl A., Espinoza C., Zhang X., Suter B.A., Jonas P. (2021) How connectivity rules and synaptic properties shape the efficacy of pattern separation in the entorhinal cortex-dentate gyrus-CA3 network. *Nat Comput Sci* **1**:830-842 <https://doi.org/10.1038/s43588-021-00157-1> | PubMed
44. Besnard A., Sahay A. (2021) Enhancing adult neurogenesis promotes contextual fear memory discrimination and activation of hippocampal-dorsolateral septal circuits. *Behav Brain Res* **399**:112917 <https://doi.org/10.1016/j.bbr.2020.112917> | PubMed
45. Berdugo-Vega G., Lee C.C., Garthe A., Kempermann G., Calegari F. (2021) Adult-born neurons promote cognitive flexibility by improving memory precision and indexing. *Hippocampus* **31**:1068-1079 <https://doi.org/10.1002/hipo.23373> | PubMed
46. Berdugo-Vega G., Dhingra S., Calegari F. (2023) Sharpening the blades of the dentate gyrus: how adult-born neurons differentially modulate diverse aspects of hippocampal learning and memory. *EMBO J* **42**:e113524 <https://doi.org/10.15252/embj.2023113524> | PubMed
47. van Praag H., Schinder A.F., Christie B.R., Toni N., Palmer T.D., Gage F.H. (2002) Functional neurogenesis in the adult hippocampus. *Nature* **415**:1030-1034 <https://doi.org/10.1038/4151030a> | PubMed
48. Zhao C., Teng E.M., Summers R.G., Ming G.L., Gage F.H. (2006) Distinct morphological stages of dentate granule neuron maturation in the adult mouse hippocampus. *J Neurosci* **26**:3-11 <https://doi.org/10.1523/JNEUROSCI.3648-05.2006> | PubMed
49. Toni N., Teng E.M., Bushong E.A., Aimone J.B., Zhao C., Consiglio A., van Praag H., Martone M.E., Ellisman M.H., Gage F.H. (2007) Synapse formation on neurons born in the adult hippocampus. *Nat Neurosci* **10**:727-734 <https://doi.org/10.1038/nn1908> | PubMed
50. Temprana S.G., Mongiat L.A., Yang S.M., Trinchero M.F., Alvarez D.D., Kropff E., Giacomini D., Beltramone N., Lanuza G.M., Schinder A.F. (2015) Delayed coupling to feedback inhibition during a critical period for the integration of adult-born granule cells. *Neuron* **85**:116-130 <https://doi.org/10.1016/j.neuron.2014.11.023> | PubMed
51. Johnston S.T., Shtrahman M., Parylak S., Goncalves J.T., Gage F.H. (2016) Paradox of pattern separation and adult neurogenesis: A dual role for new neurons balancing memory resolution and robustness. *Neurobiol Learn Mem* **129**:60-68 <https://doi.org/10.1016/j.nlm.2015.10.013> | PubMed
52. Dieni C.V., Gonzalez J.C., Overstreet-Wadiche L. (2019) Multifaceted circuit functions of adult-born neurons. *F1000Res* **8** <https://doi.org/10.12688/f1000research.20642.1> | PubMed
53. Luna V.M., Anacker C., Burghardt N.S., Khandaker H., Andreu V., Millette A., Leary P., Ravenelle R., Jimenez J.C., Mastrodonato A., et al. (2019) Adult-born hippocampal neurons bidirectionally modulate entorhinal inputs into the dentate gyrus. *Science* **364**:578-583 <https://doi.org/10.1126/science.aat8789> | PubMed
54. Tuncdemir S.N., Lacefield C.O., Hen R. (2019) Contributions of adult neurogenesis to dentate gyrus network activity and computations. *Behav Brain Res* **374**:112112 <https://doi.org/10.1016/j.bbr.2019.112112> | PubMed
55. Clelland C.D., Choi M., Romberg C., Clemenson G.D., Fragniere A., Tyers P., Jessberger S., Saksida L.M., Barker R.A., Gage F.H., et al. (2009) A functional role for adult hippocampal neurogenesis in spatial pattern separation. *Science* **325**:210-213 <https://doi.org/10.1126/science.1173215> | PubMed

56. Sahay A., Scobie K.N., Hill A.S., O'Carroll C.M., Kheirbek M.A., Burghardt N.S., Fenton A.A., Dranovsky A., Hen R (2011) Increasing adult hippocampal neurogenesis is sufficient to improve pattern separation. *Nature* **472**:466-470 <https://doi.org/10.1038/nature09817> | PubMed
57. Creer D.J., Romberg C., Saksida L.M., van Praag H., Bussey T.J (2010) Running enhances spatial pattern separation in mice. *Proc Natl Acad Sci U S A* **107**:2367-2372 <https://doi.org/10.1073/pnas.0911725107> | PubMed
58. Kim E.J., Ables J.L., Dickel L.K., Eisch A.J., Johnson J.E (2011) Ascl1 (Mash1) defines cells with long-term neurogenic potential in subgranular and subventricular zones in adult mouse brain. *PLoS One* **6**:e18472 <https://doi.org/10.1371/journal.pone.0018472> | PubMed
59. Castillon C., Gonzalez L., Domenichini F., Guyon S., Da Silva K., Durand C., Lestaevel P., Vaillend C., Laroche S., Barnier, J.V., et al. (2020) The intellectual disability PAK3 R67C mutation impacts cognitive functions and adult hippocampal neurogenesis. *Hum Mol Genet* **29**:1950-1968 <https://doi.org/10.1093/hmg/ddz296> | PubMed
60. Sloviter R.S., Dichter M.A., Rachinsky T.L., Dean E., Goodman J.H., Sollas A.L., Martin D.L (1996) Basal expression and induction of glutamate decarboxylase GABA in excitatory granule cells of the rat and monkey hippocampal dentate gyrus. *The Journal of Comparative Neurology* **373**:593-618 [https://doi.org/10.1002/\(sici\)1096-9861\(19960930\)373:4<593::aid-cne8>3.3.co;2-s](https://doi.org/10.1002/(sici)1096-9861(19960930)373:4<593::aid-cne8>3.3.co;2-s)
61. Perederiy J.V., Luikart B.W., Washburn E.K., Schnell E., Westbrook G.L (2013) Neural injury alters proliferation and integration of adult-generated neurons in the dentate gyrus. *J Neurosci* **33**:4754-4767 <https://doi.org/10.1523/JNEUROSCI.4785-12.2013> | PubMed

Peer reviews

Reviewer #1 (Public review):

This manuscript investigates how dentate gyrus (DG) granule cell subregions, specifically suprapyramidal (SB) and infrapyramidal (IB) blades, are differentially recruited during a high cognitive demand pattern separation task. The authors combine TRAP2 activity labeling, touchscreen-based TUNL behavior, and chemogenetic inhibition of adult-born dentate granule cells (abDGCs) or mature granule cells (mGCs) to dissect circuit contributions.

This manuscript presents an interesting and well-designed investigation into DG activity patterns under varying cognitive demands and the role of abDGCs in shaping mGC activity. The integration of TRAP2-based activity labeling, chemogenetic manipulation, and behavioral assays provides valuable insight into DG subregional organization and functional recruitment. However, several methodological and quantitative issues limit the interpretability of the findings. Addressing the concerns below will greatly strengthen the rigor and clarity of the study.

Major points:

- (1) Quantification methods for TRAP+ cells are not applied consistently across panels in Figure 1, making interpretation difficult. Specifically, Figure 1F reports TRAP+ mGCs as density, whereas Figure 1G reports TRAP+ abDGCs as a percentage, hindering direct comparison. Additionally, Figure 1H presents reactivation analysis only for mGCs; a parallel analysis for abDGCs is needed for comparison across cell types.
- (2) The anatomical distribution of TRAP+ cells is different between low- and high-cognitive demand conditions (Figure 2). Are these sections from dorsal or ventral DG? Is this specific to dorsal DG, as it is preferentially involved in cognitive function? What happens in ventral DG?
- (3) The activity manipulation using chemogenetic inhibition of abDGCs in AsclCreER; hM4 mice was performed; however, because tamoxifen chow was administered for 4 or 7 weeks, the labeled abDGC population was not properly birth-dated. Instead, it consisted of a

heterogeneous cohort of cells ranging from 0 to 5-7 weeks old. Thus, caution should be taken when interpreting these results, and the limitations of this approach should be acknowledged.

(4) There is a major issue related to the quantification of the DREADD experiments in Figure 4, Figure 5, Figure 6, and Figure 7. The hM4 mouse line used in this study should be quantified using HA, rather than mCitrine, to reliably identify cells derived from the *Ascl* lineage. mCitrine expression in this mouse line is not specific to adult-born neurons (off-targets), and its expression does not accurately reflect hM4 expression.

(5) Key markers needed to assess the maturation state of abDGCs are missing from the quantification. Incorporating DCX and NeuN into the analysis would provide essential information about the developmental stage of these cells.

Minor points:

- (1) The labeling (Distance from the hilus) in Figure 2B is misleading. Is that the same location as the subgranular zone (SGZ)? If so, it's better to use the term SGZ to avoid confusion.
- (2) Cell number information is missing from Figures 2B and 2C; please include this data.
- (3) Sample DG images should clearly delineate the borders between the dentate gyrus and the hilus. In several images, this boundary is difficult to discern.
- (4) In Figure 6, it is not clear how tamoxifen was administered to selectively inhibit the more mature 6-7-week-old abDGC population, nor how this paradigm differs from the chow-based approach. Please clarify the tamoxifen administration protocol and the rationale for its specificity.

Comments on revisions:

I appreciate the authors' careful and thorough revisions. They have addressed all of my previous concerns satisfactorily, and the manuscript is now significantly strengthened. I have no further concerns.

<https://doi.org/10.7554/eLife.109611.2.sa3>

Reviewer #2 (Public review):

In this study, the authors investigate how increasing cognitive demand shapes activity patterns in the dorsal dentate gyrus (DG). Using a touchscreen-based TUNL task combined with TRAP/c-Fos tagging, birth-dating of adult-born granule cells (abDGCs), and chemogenetic inhibition, they show that higher task demand increases mature granule cell (mGC) recruitment and enhances suprapyramidal (SB) versus infrapyramidal (IB) blade bias. Functionally, mGC inhibition reduces overall activity and impairs performance without disrupting blade bias, whereas inhibition of {less than or equal to}7-week-old abDGCs increases mGC activity, abolishes blade bias, and impairs discrimination under high-demand conditions. These findings suggest that effective pattern separation depends not only on overall DG activity levels but also on the spatial organization of recruited ensembles.

The integration of touchscreen TUNL with temporally controlled activity tagging and birth-dated cohorts is technically strong. Quantification of SB-IB bias and radial/apical distributions adds anatomical precision beyond bulk activity measures. The comparison between mGC and abDGC inhibition is conceptually compelling and supports dissociable functional roles. Overall, the data convincingly demonstrate that increasing cognitive demand amplifies blade-biased DG recruitment and that mGCs and abDGCs differentially contribute to both behavioral performance and network organization.

However, how abDGCs are integrated into the mGC network under high cognitive demand remains unresolved. Additional experiments are needed to clarify how abDGCs shape spatial recruitment patterns and whether they directly inhibit or indirectly regulate mGC activity to maintain high performance.

Furthermore, the authors frame "high cognitive demand" as a multidimensional construct encompassing broad behavioral challenge. It would strengthen the work to delineate how local abDGC-mGC circuit interactions regulate specific task components in real time. This will require higher temporal resolution approaches, as TRAP and c-Fos labeling integrate activity over prolonged windows and primarily reflect sustained engagement rather than moment-to-moment computations.

The central conclusion that dentate function depends on coordinated spatial recruitment rather than total activity magnitude is supported by the data, although mechanistic interpretations should be tempered given methodological limitations.

Overall, this work advances models of adult neurogenesis by emphasizing a critical-period modulatory role of abDGCs in organizing DG network activity during high-demand discrimination. The combined behavioral and circuit-level framework is likely to be influential in the field.

<https://doi.org/10.7554/eLife.109611.2.sa2>

Reviewer #3 (Public review):

This study examines the role of dentate gyrus neuronal populations, reflecting neurogenesis and anatomical location (suprapyramidal vs infrapyramidal blade), in a mnemonic discrimination task that taxes the pattern separation functions of the dentate. The authors measure dentate gyrus activity resulting from cognitive training and test whether adult neurogenesis is required for both the anatomical patterns of activity and performance in the cognitive task. The authors find that more cognitively challenging variants of the task evoked more dentate activity, but also distinct patterns of activity (more activity in the suprapyramidal blade, less in the infrapyramidal blade). Using chemogenetic approaches they silence mature vs immature dentate gyrus neurons and find that only mature neurons (either the general population or specifically mature adult-born neurons), and not immature adult-born neurons, are required for the difficult version of the task. Inhibition of mature adult-born neurons furthermore increased overall activity in the dentate and reduced the biased pattern of activity across the blades, consistent with evidence that adult-born neurons broadly regulate dentate gyrus activity.

Comments on revisions:

I appreciate the efforts the authors have taken to revise this manuscript. I have only minor concerns with this revised version of the manuscript:

Methods state that significance is defined as $P < 0.05$ but some results are interpreted as significant when $P = 0.05$. Either the alpha value needs to change or the interpretation needs to change.

I believe the statistical results for group and blade effects for the ANOVAs, in Figs 2,3 & 4, appear to be switched (blade should be significant, not group).

I appreciate that sometimes there is not a perfect overlap between immunohistochemical signals, but I continue to believe that the spatially-non-overlapping TRAP and EDU signals in Fig 3 is caused by these 2 markers being in different cells. A Z-stack or orthogonal projection could verify/disprove this concern.

<https://doi.org/10.7554/eLife.109611.2.sa1>

Author Response:

The following is the authors' response to the original reviews.

Public Reviews:

Reviewer #1 (Public review):

This manuscript investigates how dentate gyrus (DG) granule cell subregions, specifically suprapyramidal (SB) and infrapyramidal (IB) blades, are differentially recruited during a high cognitive demand pattern separation task. The authors combine TRAP2 activity labeling, touchscreen-based TUNL behavior, and chemogenetic inhibition of adult-born dentate granule cells (abDGCs) or mature granule cells (mGCs) to dissect circuit contributions.

This manuscript presents an interesting and well-designed investigation into DG activity patterns under varying cognitive demands and the role of abDGCs in shaping mGC activity. The integration of TRAP2-based activity labeling, chemogenetic manipulation, and behavioral assays provides valuable insight into DG subregional organization and functional recruitment. However, several methodological and quantitative issues limit the interpretability of the findings. Addressing the concerns below will greatly strengthen the rigor and clarity of the study.

Major points:

(1) Quantification methods for TRAP+ cells are not applied consistently across panels in Figure 1, making interpretation difficult. Specifically, Figure 1F reports TRAP+ mGCs as density, whereas Figure 1G reports TRAP+ abDGCs as a percentage, hindering direct comparison. Additionally, Figure 1H presents reactivation analysis only for mGCs; a parallel analysis for abDGCs is needed for comparison across cell types.

In Figure 1G and 1H we report TRAP+ abDGCs as a percentage rather than density because we are analyzing colocalization of the two markers, which are very sparse in this population. Given the very low number of double-labeled abDGCs, calculating density would not be practical. In the revised manuscript we have clarified the rationale for using these measures. As noted in the current text, we did not observe abDGCs co-expressing TRAP and c-Fos; we have made this point more explicit to guide interpretation of these data.

(2) The anatomical distribution of TRAP+ cells is different between low- and high-cognitive demand conditions (Figure 2). Are these sections from dorsal or ventral DG? Is this specific to dorsal DG, as it is preferentially involved in cognitive function? What happens in ventral DG?

The sections shown in Figure 2 were obtained from the dorsal dentate gyrus (see Methods, "Histology and imaging": stereotaxic coordinates -1.20 to -2.30 mm relative to bregma, Paxinos atlas). From a feasibility standpoint, it is not possible to analyze the entire longitudinal extent of the hippocampus with these low-throughput histological approaches. We therefore focused on the dorsal DG, for which there is a strong functional rationale. A large body of work indicates that the dorsal hippocampus, and specifically the dorsal DG, is preferentially involved in spatial memory and in the fine contextual discrimination that underlies pattern separation. The dorsal hippocampus is critical for encoding and distinguishing similar spatial representations, a core component of the high-cognitive demand task used here. In contrast, the ventral DG is more strongly associated with emotional regulation and affective memory processing and is less implicated in high-

resolution spatial encoding. For these reasons, the present study was designed to assess TRAP⁺ cell distributions specifically in the dorsal DG.

(3) The activity manipulation using chemogenetic inhibition of abDGCs in AsclCreER; hM4 mice was performed; however, because tamoxifen chow was administered for 4 or 7 weeks, the labeled abDGC population was not properly birth-dated. Instead, it consisted of a heterogeneous cohort of cells ranging from 0 to 5-7 weeks old. Thus, caution should be taken when interpreting these results, and the limitations of this approach should be acknowledged.

We agree that prolonged tamoxifen administration results in labeling a heterogeneous population of abDGCs spanning approximately 0 to 5–7 weeks of age, rather than a precisely birth-dated cohort. This is a limitation of this approach and we have included discussion of this in more detail in the revised manuscript.

(4) There is a major issue related to the quantification of the DREADD experiments in Figure 4, Figure 5, Figure 6, and Figure 7. The hM4 mouse line used in this study should be quantified using HA, rather than mCitrine, to reliably identify cells derived from the Ascl lineage. mCitrine expression in this mouse line is not specific to adult-born neurons (off-targets), and its expression does not accurately reflect hM4 expression.

We agree that mCitrine is not a marker that allows localization of hM4Di as it is well known that the mCitrine can be independently expressed in a Cre independent manner in this mouse. As suggested, we have removed the figure that showed the mCitrine and have performed immunohistochemical localization of the DREADD with an antibody against the HA tag. This is now shown in Figure 5.

(5) Key markers needed to assess the maturation state of abDGCs are missing from the quantification. Incorporating DCX and NeuN into the analysis would provide essential information about the developmental stage of these cells.

The goal of this study was to examine activity patterns of adult-born versus mature granule cells, rather than to assess maturation state. The adult-born neurons analyzed were 25–39 days old, an age at which point most cells have progressed beyond the DCX⁺ stage and are expected to express NeuN based on prior work. We therefore do not think that including DCX or NeuN quantification would provide additional information relevant to the aims or interpretation of this study.

Minor points:

(1) The labeling (Distance from the hilus) in Figure 2B is misleading. Is that the same location as the subgranular zone (SGZ)? If so, it's better to use the term SGZ to avoid confusion.

We have updated Figure 2B, the Methods, and the main text to more explicitly localize this which is the boundary between the subgranular zone (SGZ) and the hilus.

(2) Cell number information is missing from Figures 2B and 2C; please include this data.

We have now added the cell number information to the figure legends. In Figures 2B and 2C, each point corresponds to a single cell, with an equal number of mice per group. The total number of TRAP⁺ cells per mouse is shown in Figure 1F, which reports TRAP⁺ cell densities by group.

(3) Sample DG images should clearly delineate the borders between the dentate gyrus and the hilus. In several images, this boundary is difficult to discern.

We made the DG-hilus boundaries clearer in the sample images to improve visualization and interpretation.

(4) In Figure 6, it is not clear how tamoxifen was administered to selectively inhibit the more mature 6-7-week-old abDGC population, nor how this paradigm differs from the chow-based approach. Please clarify the tamoxifen administration protocol and the rationale for its specificity.

We apologize for the confusion here. The protocol used in Figure 6 is the same tamoxifen chow-based approach as in Figure 5, differing only in the duration of tamoxifen exposure. Mice in Figure 5 received tamoxifen chow for 7 weeks, whereas mice in Figure 6 received it for 4 weeks, restricting labeling to a younger and narrower cohort of adult-born DGCs. Thus, the population targeted in Figure 6 is younger than that in Figure 5 and does not correspond to mature 6–7-week-old neurons. By contrast, the experiment in Figure 4 targets a more mature population, consisting predominantly of ~5-week-old adult-born neurons as well as mature granule cells, which are Dock10-positive and express Cre endogenously, allowing selective manipulation of this later-stage population.

We have corrected the paragraph accordingly and clarified the age range of the labeled populations in the revised manuscript.

Reviewer #2 (Public review):

Summary

In this manuscript, the authors combine an automated touchscreen-based trial-unique nonmatching-to-location (TUNL) task with activity-dependent labeling (TRAP/c-Fos) and birth-dating of adult-born dentate granule cells (abDGCs) to examine how cognitive demand modulates dentate gyrus (DG) activity patterns. By varying spatial separation between sample and choice locations, the authors operationally increase task difficulty and show that higher demand is associated with increased mature granule cell (mGC) activity and an amplified suprapyramidal (SB) versus infrapyramidal (IB) blade bias. Using chemogenetic inhibition, they further demonstrate dissociable contributions of abDGCs and mGCs to task performance and DG activation patterns.

The combination of behavioral manipulation, spatially resolved activity tagging, and temporally defined abDGC perturbations is a strength of the study and provides a novel circuit-level perspective on how adult neurogenesis modulates DG function. In particular, the comparison across different abDGC maturation windows is well designed and narrows the functionally relevant population to neurons within the critical period (~4-7 weeks). The finding that overall mGC activity levels, in addition to spatially biased activation patterns, are required for successful performance under high cognitive demand is intriguing.

Major Comments

(1) *Individual variability and the relationship between performance and DG activation.*

The manuscript reports substantial inter-animal variability in the number of days required to reach the criterion, particularly during large-separation training. Given this variability, it would be informative to examine whether individual differences in performance correlate with TRAP+ or c-Fos+ density and/or spatial bias metrics. While the authors report no correlation between success and TRAP+ density in some analyses, a more systematic correlation across learning rate, final performance, and DG activation patterns (mGC vs abDGC, SB vs IB) could strengthen the interpretation that DG activity reflects task engagement rather than performance only.

As mentioned, we previously reported no correlation between task success and TRAP+ density. We have now performed additional analyses examining correlations with learning rate, final performance, and DG activation patterns (mGC vs abDGC, SB vs IB), and found no significant relationships. Therefore, as we did not find any positive correlations the original interpretation that DG activity primarily reflects task engagement rather than performance level seems the most parsimonious.

(2) Operational definition of "cognitive demand".

The distinction between low (large separation) and high (small separation) cognitive demand is central to the manuscript, yet the definition remains somewhat broad. Reduced spatial separation likely alters multiple behavioral variables beyond cognitive load, including reward expectation, attentional demands, confidence, engagement, and potentially motivation. The authors should more explicitly acknowledge these alternative interpretations and clarify whether "cognitive demand" is intended as a composite construct rather than a strictly defined cognitive operation.

We agree that reducing spatial separation between stimuli likely engages multiple behavioral and cognitive processes beyond a single, strictly defined operation. We have now clarified this point in the manuscript and explicitly state that our use of the term “cognitive demand” reflects a multidimensional behavioral challenge rather than a singular cognitive process (see Discussion).

(3) Potential effects of task engagement on neurogenesis.

Given the extensive behavioral training and known effects of experience on adult neurogenesis, it remains unclear whether the task itself alters the size or maturation state of the abDGC population. Although the focus is on activity and function rather than cell number, it would be useful to clarify whether neurogenesis rates were assessed or controlled for, or to explicitly state this as a limitation.

While the primary goal of this study was to examine activity and functional recruitment of adult-born granule cells, we also quantified the survival of birth-dated neurons at the end of behavioral training. Density measurements of BrdU⁺ and EdU⁺ cells revealed no differences across experimental groups, indicating that engagement in the pattern separation task, across low to high cognitive demand conditions, did not significantly alter survival of adult-born neurons. In addition, we examined the spatial distribution of BrdU⁺ and EdU⁺ neurons between the suprapyramidal and infrapyramidal blades of the dentate gyrus. The proportion of newborn neurons was consistent across all groups, with approximately 60% located in the suprapyramidal blade and 40% in the infrapyramidal blade. These findings indicate that behavioral training did not alter the baseline distribution of adult-born neurons. We have now clarified these points in the manuscript (See Results).

(4) Temporal resolution of activity tagging.

TRAP and c-Fos labeling provide a snapshot of neural activity integrated over a temporal window, making it difficult to determine which task epochs or trial types drive the observed activation patterns. This limitation is partially acknowledged, but the conclusions occasionally imply trial-specific or demand-specific encoding. The authors should more clearly distinguish between sustained task engagement and moment-to-moment trial processing, and temper interpretations accordingly. While beyond the scope of the current study, this also motivates future experiments using in vivo recording approaches.

We agree and have made changes to the manuscript to discuss these points (see Discussion and Limitations).

(5) *Interpretation of altered spatial patterns following abDGC inhibition.*

In the abDGC inhibition experiments, Cre+ DCZ animals show delayed learning relative to controls. As a result, when animals are sacrificed, they may be at an intermediate learning stage rather than at an equivalent behavioral endpoint. This raises the possibility that altered DG activation patterns reflect the learning stage rather than a direct circuit effect of abDGC inhibition. Additional clarification or analysis controlling for the learning stage would strengthen the causal interpretation.

We agree that differences in learning stage could in principle confound the interpretation of DG activation patterns. However, although Cre+ DCZ-treated mice exhibited delayed learning, they ultimately reached the same performance criterion as control animals. Thus, adult-born DGC inhibition did not prevent learning but increased the time required to reach criterion, indicating that these neurons are beneficial for learning efficiency rather than strictly necessary for task acquisition. Importantly, all animals were sacrificed only after reaching the predefined success criterion. Therefore, the immunohistochemical analyses were performed at the same behavioral endpoint for Cre+ DCZ and control groups, even though the number of training days differed. Consequently, the observed differences in DG activation reflect circuit recruitment at equivalent task mastery rather than differences in learning stage.

(6) *Relationship between c-Fos density and behavioral performance.*

The study reports that abDGC inhibition increases c-Fos density while impairing performance, whereas mGC inhibition decreases c-Fos density and also impairs performance. This raises an important conceptual question regarding the relationship between overall activity levels and task success. The authors suggest that both sufficient activity and appropriate spatial patterning are required, but the manuscript would benefit from a more explicit discussion of how different perturbations may shift the identity, composition, or coordination of the active neuronal ensemble rather than simply altering total activity levels.

We agree that our findings highlight that successful performance is not determined solely by the overall level of dentate gyrus activity, but rather by the composition and spatial organization of the active neuronal ensemble. In our study, inhibition of abDGCs increased overall mGC activity while disrupting the spatially organized, blade-biased activation pattern and impaired performance. In contrast, direct inhibition of mGCs reduced global excitability but preserved the relative spatial organization of active neurons in animals that continued to perform the task. These findings suggest that different perturbations alter task performance by shifting the identity and coordination of the active neuronal ensemble, rather than simply increasing or decreasing total activity levels. We have now expanded the Discussion to more explicitly address how dentate gyrus computations may depend on the structured recruitment of granule cell ensembles and how distinct manipulations differentially disrupt this organization.

Reviewer #3 (Public review):

Summary:

The authors used genetic models and immunohistochemistry to identify how training in a spatial discrimination working memory task influences activity in the dentate gyrus subregion of the hippocampus. Finding that more cognitively challenging variants of the task evoked more and distinct patterns of activity, they then investigated whether newborn neurons in particular were important for learning this task and regulating the spatial activity patterns.

Strengths:

The focus on precise anatomical locations of activity is relatively novel and potentially important, given that little is known about how DG subregions contribute to behavior. The authors also use a task that is known to depend on this memory-related part of the brain.

Weaknesses:

Statistical rigor is insufficient. Many statistical results are not stated, inappropriate tests are used, and sample sizes differ across experiments (which appear to potentially underlie null results). The chemogenetic approach to inhibit adult-born neurons also does not appear to be targeting these neurons, as judged by their location in the DG.

Please refer to the updated statistical analyses in response to the recommendations below.

Recommendations for the authors:**Reviewing Editor Comments**

*Please note that reviewers agreed that appropriate revisions are needed to increase the strength of evidence for the paper's claims. Concerns were raised about a lack of statistical rigor in the statistical analyses used. Results of statistical tests were not consistently provided (i.e., statistic applied, value of statistic, degrees of freedom, p-value), and seemingly inappropriate statistical tests were used in some instances. Also, some comparisons had lower statistical power than others. When clarifying the statistical approaches used in the manuscript, we also encourage you to consider reading this article that outlines common statistical mistakes (Makin TR, Orban de Xivry JJ. Ten common statistical mistakes to watch out for when writing or reviewing a manuscript. *Elife*. 2019 Oct 9;8:e48175. doi: 10.7554/eLife.48175.), such as the importance of not basing conclusions on a significant p-value for one pair-wise comparison vs a non-significant p-value for another pairwise comparison (i.e., groups that are being compared should be included in the same statistical analysis, and interaction effects should be reported when appropriate). We hope that you find this information to be helpful should you decide to submit a revised manuscript to eLife.*

Reviewer #1 (Recommendations for the authors):

(1) Standardize TRAP+ quantification across Figure 1.

Please report TRAP+ cell numbers using consistent metrics (e.g., density or percentage) to enable comparison across cell types. In addition, extend the TRAP+ reactivation analysis in Figure 1H to include abDGCs so that reactivation dynamics can be compared directly between mGCs and abDGCs.

Reply in Public Review

(2) Clarify whether dorsal or ventral DG was analyzed in Figure 2.

The differing anatomical distributions of TRAP+ cells under low- and high-demand conditions raise important questions about DG axis specificity. Please indicate whether analyses were performed in dorsal DG, ventral DG, or both, and provide data or justification accordingly.

Reply in Public Review

(3) Acknowledge limitations of the tamoxifen-chow labeling strategy in *AsclCreER*; *hM4* experiments.

Since tamoxifen chow administered over 4-7 weeks labels a heterogeneous abDGC population spanning a broad age range, this approach does not generate birth-dated cohorts. This limitation should be clearly addressed in the text and interpretations, particularly related to cell age-dependent effects, should be tempered.

Reply in Public Review

(4) Revise DREADD quantification using HA rather than mCitrine.

The hM4 mouse line requires HA immunostaining to accurately identify Ascl-lineage cells expressing the DREADD receptor. Because mCitrine is not specific to adult-born neurons and does not reliably reflect hM4 expression, quantification based on mCitrine should be revised.

Reply in Public Review

(5) Include markers to assess abDGC maturation state.

Adding quantification of DCX and NeuN would help define the developmental stage of abDGCs in key experiments and improve the interpretation of cell-age-dependent effects.

Reply in Public Review

(6) Clarify DG layer boundaries and terminology in Figure 2.

If the metric labeled "Distance from the hilus" corresponds to the subgranular zone (SGZ), using SGZ terminology would prevent confusion. Additionally, please provide clearer delineation of DG and hilus borders in sample images.

Reply in Public Review

(7) Provide missing cell number data for Figures 2B and 2C.

Reply in Public Review

(8) Clarify the tamoxifen administration protocol in Figure 6.

Please describe how the protocol selectively targets 6-7-week-old abDGCs and how it differs from the chow-based approach. This will help readers understand the intended specificity of the manipulation.

Reply in Public Review

Reviewer #2 (Recommendations for the authors):

(1) EdU birth-dating timeline

The manuscript would benefit from a clearer description of the EdU birth-dating timeline, ideally with a schematic similar to that provided for BrdU in Supplementary Figure 1.

We appreciate the suggestion. However, we did not include a separate schematic for EdU because its use and birth-dating logic are identical to BrdU (both are thymidine analogs administered systemically and incorporated during S-phase). Therefore, the timeline shown in Supplementary Figure 1 applies equally to both markers. We have clarified this point in the Methods section to avoid confusion.

(2) Clarity of TUNL task description.

The description of the TUNL task, particularly for readers unfamiliar with touchscreen-based paradigms, is difficult to follow without consulting prior literature. A simplified

schematic or a clearer step-by-step explanation in the main text or supplementary material would improve accessibility.

We note that the main steps of the TUNL protocol are illustrated in Figure 1A, Supplementary Figure 2A and 2B. Nevertheless, we agree that the description in the text can be made clearer for readers less familiar with touchscreen-based tasks. Thus, we have now revised the Methods section to provide a clearer step-by-step description of the TUNL.

(3) Influence of outliers in Figure 1G.

In Figure 1G, the reported trend that ~1% of 25-39-day-old abDGCs are TRAP+ during LS trials appears to be driven by a small number of outliers. This should be acknowledged, and the wording of the conclusion moderated to reflect the variability in the data.

We agree with the reviewer that the apparent outliers reflect the inherent sparsity of TRAP labeling in this population. In absolute terms, this corresponds to between 0 and 2 TRAP⁺ 25–39-day-old abDGCs per mouse, such that the presence or absence of a small number of labeled cells can appear as outliers when expressed as a percentage. We have revised the text to acknowledge this (see Results).

(4) Presentation of learning curves.

Rather than focusing primarily on "days before criterion" (DBC), it would be helpful to show full learning curves across the entire training period. This would provide a clearer picture of acquisition dynamics and inter-animal variability.

We agree that learning curves can be informative in many behavioral paradigms. However, in our protocol, mice do not undergo the same number of training days because training stops individually once each animal reaches criterion. As a result, plotting full learning curves would produce trajectories of different lengths, making group comparisons difficult and visually cluttered. For this reason, we aligned animals based on days before criterion (DBC), which allows direct comparison of learning dynamics relative to task acquisition. We also consider the cumulative probability representation to be the most appropriate way to summarize learning progression across animals in this context which are also included in the figures.

(5) Clarification of Figure 3B labeling

In Figure 3B, the identity of the orange-labeled group above the LS condition is unclear. Clarification in the figure legend would improve interoperability.

Figure 3B includes two experimental groups. One group performed both the large- and small-separation conditions; this group is shown in orange and labeled LS. Within this group, the upper orange trace corresponds to performance in the large-separation condition, while the lower orange trace corresponds to performance in the small-separation condition. The second group is a control group that performed only the large-separation configuration, and therefore only a single green trace is shown. We agree that this distinction was not sufficiently clear and have revised the figure legend and text to clarify the identity of each trace.

Reviewer #3 (Recommendations for the authors):

(1) Please label figures and, even better, put the legends on the same page.

(2) Just to confirm, in establishing the task, mice performed above 70% for the small separation trials in one of the sessions on 2 consecutive days, for each criterion? Performance seems to be below 70%.

Yes. To meet the criterion, each mouse had to reach $\geq 70\%$ correct performance in at least one of the two daily sessions on two consecutive days. We then averaged the performance across both sessions for each of those days. As a result, if one session was $\geq 70\%$ but the other was lower, the daily average could fall below 70%. The values shown in the figure correspond to these daily averages, further averaged across mice.

(3) mGC needs to be explicitly defined. Am I assuming any non-birthdated GC is an mGC according to the authors? (which means it is unknown whether they are in fact mature, though likely most of them are).

In this study, “mature granule cells” (mGCs) refer operationally to granule cells that are not birth-dated with BrdU or EdU and therefore are not classified as adult-born neurons within the defined labeling window. We agree that this population is not directly age-defined, and that while the majority are expected to be mature based on their birth timing relative to the labeling period, we cannot exclude the possibility that a small fraction may include younger, unlabeled neurons. We have now explicitly defined this usage of mGCs in the Methods and clarified this point in the text to avoid ambiguity.

(4) Methods state that Kruskal-Wallis tests were used when more than 3 groups were compared, but I don't see these stats presented (e.g., for trap data in Figure 1, blade x task TRAP expt in Figure 3 (should be 2-way RM anova here and elsewhere), etc) or any corrections for multiple comparisons. I appreciate that the mean rates of TRAPed abGCs are higher in the S and LS groups than in the shaping group, but most mice do not have any BrdU+ cells that are also TRAPed, and there are no statistics here to support the claim. I don't think there is enough sampling to accurately quantify activation of abGCs. Also, no stats to support the claim that TRAPing increases at the "tip of the SB after the more demanding LS task".

We agree with this comment. We have now systematically tested all datasets for normality (by group) and applied parametric tests when the data met normality assumptions, and non-parametric tests otherwise. The statistical analyses have been revised accordingly. We added the appropriate tests (including two-way ANOVA where relevant, such as for blade \times group comparisons) and now report full statistics in the figure legends and results sections. For the TRAP analyses in adult-born DGs, we explicitly acknowledge the very low number of BrdU⁺/TRAP⁺ cells, which limits statistical power and, in some cases, precludes robust statistical testing. These limitations are now clearly stated in the Results and Discussion, and the corresponding interpretations have been tempered. For all Kruskal–Wallis tests, post hoc pairwise comparisons were performed using Dunn’s test, with Bonferroni correction for multiple comparisons, as now specified in the Methods section. We also expanded the Methods to describe the statistical workflow in detail. In addition, we have added the previously missing statistical analysis for Figure 2C. Comparisons were performed between the 0–50% and 50–100% portions of the blade, where 0% corresponds to the apex and 100% corresponds to the distal tip of the blade.

(5) Figure 3I: I can't figure out which effect is statistically significant here (what does the asterisk signify?). Why no individual data points in this graph?

We agree that the absence of individual data points reduced interpretability, and we have now updated the figure to include individual data points to better illustrate data distribution and variability.

(6) The gradient of activity (shap < S < LS) could be due to how long they've been trained on a given stage (e.g. less activity during shaping because they have habituated, and neurons encoding that task phase have already been selected)

We agree that task duration and habituation could, in principle, influence activity levels. Under this interpretation, higher activity would primarily reflect task novelty rather than cognitive demand. However, our data do not support this explanation. Specifically, we found no correlation between the number of training days required to reach criterion and c-Fos-positive or TRAP-positive cell density within a given stage. Thus, animals that reached criterion rapidly did not show higher activity levels than animals that required more days of training and were presumably more habituated to the task demands. This suggests that the observed activity gradient (shaping < S < LS) is not driven by exposure duration or habituation, but rather reflects differences in cognitive demand across task stages.

(7) The TRAP+ EDU+ cell in Figure 3 looks odd because the BrdU signal is (a lot) larger than the TRAP signal, but BrdU is in the nucleus and should be smaller.

We agree that the example in Figure 3 is not optimal. In dividing cells, BrdU/EdU signals can sometimes appear broader or closely apposed, which may affect their apparent size.

(8) For the Ascl-HM4Di experiment, HM4Di appears to be expressed in all of the areas of the granule cell layer where abGCs are NOT located (i.e. no expression in the deep cell layer, near the sgz). This is problematic because it suggests perhaps abGCs are not inhibited as expected.

As noted in our response to Reviewer #1, we did not use the mCitrine to localize the DREADD receptor as it has been demonstrated that mCitrine expression is expressed in a Cre-independent manner and not correlated with hM4Di expression. In the revised manuscript we include a representative image where we performed immunostaining using an HA antibody to directly visualize hM4Di and confirm its expression in adult-born granule cells (Figure 5).

(9) Line 267: "6-7 week old neurons by themselves do not influence either the performance of mice in the task". I don't think this is fair because this experiment wasn't designed with as much power to detect an effect. The group trends are in the same direction, but there are many fewer mice in this experiment (n=6/group) than in the <7w experiment (n=11/group), where the effect just reached statistical significance.

We are sorry for this confusion which came from an incorrect version. The experiment shown in Figure 6 does not target 6–7-week-old neurons specifically. It uses the same tamoxifen chow-based protocol as Figure 5, but with a shorter exposure (4 weeks vs. 7 weeks), thereby labeling a younger and more restricted cohort of adult-born DGCs. By contrast, Figure 4 targets a more mature population, consisting predominantly of ~5-week-old adult-born neurons as well as mature granule cells (Dock10+).

We have corrected the paragraph accordingly and clarified the age range of the labeled populations in the revised manuscript.

<https://doi.org/10.7554/eLife.109611.2>