

Molecular Consequences of Peripheral Influenza A Infection on Cell Populations in the Murine Hypothalamus

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Abstract

Infection with Influenza A virus (IAV) causes the well-known symptoms of the flu, including fever, loss of appetite and excessive sleepiness. These responses, mediated by the brain, will normally disappear once the virus is cleared from the system, but a severe respiratory virus infection may cause long-lasting neurological disturbances. These include encephalitis lethargica and narcolepsy. The mechanisms behind such long lasting changes are unknown. The hypothalamus is a central regulator of the homeostatic response during a viral challenge. To gain insight into the neuronal and non-neuronal molecular changes during an IAV infection, we intranasally infected mice with an H1N1 virus and extracted the brain at different time points. Using single-nucleus RNA sequencing (snRNA-seq) of the hypothalamus, we identify transcriptional effects in all identified cell populations. The snRNA-seq data showed the most pronounced transcriptional response at 3 days past infection, with a strong downregulation of genes across all cell types. General immune processes were mainly impacted in microglia, the brain resident immune cells, where we found increased numbers of cells expressing pro-inflammatory gene networks. In addition, we found that most neuronal cell populations downregulated genes contributing to the energy homeostasis in mitochondria and protein translation in the cytosol, indicating potential reduced cellular and neuronal activity. This might be a preventive mechanism in neuronal cells to avoid intracellular viral replication and attack by phagocytosing cells. The change of microglia gene activity suggest that this is complemented by a shift in microglia activity to provide increased surveillance of their surroundings.

eLife assessment

This **important** study combines experiments and computational approaches to understand the effects of influenza H1N1 infection on hypothalamic cells. The methodology and analysis are **solid** and raise questions around how a respiratory virus affects the central nervous system.

Introduction

In humans as well as other species, Influenza A virus (IAV) infection causes respiratory tract disease, evoking seasonal epidemics associated with increased morbidity and mortality rates (Macias et al., 2021 [↗](#); Paget et al., 2019 [↗](#)). Beyond seasonal epidemics, H1N1 IAV strains have been identified as the causal agent for the pandemics in 1918 and 2009, leading up to 50 million and 18,000 deaths worldwide, respectively (Simonsen et al., 2013 [↗](#); Taubenberger & Morens, 2008 [↗](#)). Beyond respiratory tract disease, IAV infections have also been associated with neurological consequences such as encephalitis lethargica and narcolepsy. Evidence from the Spanish flu pandemic in 1918 linked H1N1 influenza infections to epidemic encephalitis lethargica (Hoffman & Vilensky, 2017 [↗](#); Maurizi, 2010 [↗](#)). Similarly, increased numbers of encephalitis cases were reported during the 2009 H1N1 pandemic (Goenka et al., 2014 [↗](#); Gu et al., 2013 [↗](#); Surana et al., 2011 [↗](#)). A peak in new narcolepsy cases was also seen following the 2009 H1N1 pandemic, and while this was linked with the Pandemrix vaccine in Europe, in China it was unrelated to vaccinations and thus might instead be linked to infection with the virus itself (Ahmed et al., 2014 [↗](#); Han et al., 2011 [↗](#)). A better understanding of the direct and indirect effects of IAV infection on the brain is needed to shed light on these observations.

Some IAV strains are considered neurotropic, meaning that they hold the machinery to infiltrate the central nervous system (CNS) and replicate, but evidence suggest that even non-neurotropic influenza strains can infiltrate the brain and cause neurological disturbances (Ekstrand, 2012 [↗](#); Hodgson et al., 2012 [↗](#); Macias et al., 2021 [↗](#); Majde et al., 2007 [↗](#); Zielinski et al., 2013 [↗](#)). When non-neurotropic viruses are found in the CNS, they are either not able to replicate at all or not efficiently in the brain resident cells (Hosseini et al., 2018 [↗](#); Zielinski et al., 2013 [↗](#)). The H1N1 strain from the 2009 pandemic (H1N1 pdm09) is non-neurotropic (Sadasivan et al., 2015 [↗](#)), so the mechanism behind the neurological consequences seen after that H1N1 infection must be caused by indirect effects rather than direct viral infection of brain cells. It is well known that cytokines produced during a peripheral infection can reach the brain and cause functional disturbances (Dantzer & Kelley, 2007 [↗](#)). It has also been shown that microglia become activated during peripheral IAV infections with non-neurotropic strains. Whether this microglia activation is a neuroprotective mechanism or contributes to neuroinflammation and -degeneration remains elusive (Dusedau et al., 2021 [↗](#); Sadasivan et al., 2015 [↗](#); Wiley et al., 2015 [↗](#)).

The hypothalamus is a functional integrator of peripheral signals and regulates several physiological and behavioural functions via several specialised nuclei (Berthoud & Munzberg, 2011 [↗](#); Bonnavion et al., 2016 [↗](#); Stuber & Wise, 2016 [↗](#)). The different hypothalamic neuron populations control among other things feeding, body temperature, sleep-wake states, and circadian rhythms (Bonnavion et al., 2016 [↗](#); Timper & Bruning, 2017 [↗](#); Yousefvand, 2021). The lateral hypothalamic area (LHA) contains well described neuron populations like hypocretin (HCRT) or melanin-concentrating hormone releasing (MCH) neurons. These neurons are coupled to several biological functions including metabolism and the regulation of sleep and wake states (Oesch & Adamantidis, 2021 [↗](#)). Recent studies showed that the LHA is also involved during inflammatory processes and regulates pain perception (Bonnavion et al., 2016 [↗](#); Brown et al.,

2015 [↗](#); Fakhoury et al., 2020 [↗](#); Stuber & Wise, 2016 [↗](#)). The dorsomedial hypothalamus (DMH) receives circadian input from the suprachiasmatic nucleus to regulate downstream behavioural and physiological changes including food intake, locomotion, and core body temperature (Gooley et al., 2006 [↗](#)). Recent findings have linked neuronal subpopulations in the DMH to changes in thermoregulation and metabolism during immune activation with the bacterial cell wall component lipopolysaccharide (LPS) (Osterhout et al., 2022 [↗](#)). Several single-cell/nuclei-RNA sequencing (scRNA-seq, snRNA-seq) datasets have been published during the past years, shedding more light on the cell heterogeneity of the different hypothalamic regions, but little is known about how the transcriptome of these cell types change during environmental perturbations (Campbell et al., 2017 [↗](#); Chen et al., 2017 [↗](#); Mickelsen et al., 2019 [↗](#); Mickelsen et al., 2020 [↗](#); Romanov et al., 2017 [↗](#)).

In the current study, we used intranasal infections with H1N1 pdm09 in female mice and snRNA-seq to follow changes in the transcriptional landscape of cell populations in the medial hypothalamus. We find changes in all populations during the peak of disease. A change in cell state is seen in microglia at early infection and later strong effects are detected in astrocytes and oligodendrocytes. Interestingly, after the mice have recovered from the acute infection, all non-neuronal cell types return to their resting state, while in neurons we detect long lasting changes of the transcriptome even after full recovery.

Results

Mice were infected intranasally with influenza virus A/California/07/2009 (H1N1 pdm09) and monitored for a period of 23 days post infection for weight changes. Groups of mice were euthanized at 3, 7 and 23 days post infection (dpi) to collect lung, olfactory bulb (OB) and brain tissue for detection of viral abundance and snRNA-seq. The rationale for choosing these time points of sampling were as follows: (i) 3 dpi as a time point of initial disease onset before the start of weight loss – at this time point innate immunity is peaking; (ii) 7 dpi marks the point of strong weight loss and is where the adaptive immune response takes over, and before potential lethal outcome for some of the animals and (iii) 23 dpi as a recovery group where animals had completely regained their initial body weight (Uddback et al., 2016 [↗](#)).

IAV respiratory tract disease progression and viral infiltration into lung and olfactory bulbs

Infected mice started losing weight around 4 dpi and had the highest weight loss at 10 dpi (**Figure 1B** [↗](#)). Animals in the recovery group had regained their body weight at 23 dpi. Lungs and OB were tested for the abundance of the M1 gene of the virus, to test for viral infection of those tissues. Viral mRNA was detected at 3 and 7 dpi in the lungs but was not detected at 23 dpi (**Figure 1C** [↗](#)). In the OB, viral mRNA was detected in one animal at 3 dpi, in all animals at 7 dpi, and in four out of five animals at 23 dpi. Three of the animals did not lose weight due to the infection (7 dpi: 7.3; 23 dpi: 23.3 and 23.4) instead the presence of viral mRNA in lungs and/or OB confirmed a successful infection (Supplementary Figure 2).

Molecular and cellular composition of the medial hypothalamus snRNA-seq dataset

For snRNA-seq we dissected a hypothalamic region containing both the DMH and the LHA from brains obtained from infected mice and the control group (**Figure 1D** [↗](#), n=5 per group). Each sample was hashed individually, samples from one time point were pooled together and sequenced. After sequencing, samples were demultiplexed for the pooled samples at the different time points. We detected a total of 44,201 nuclei (henceforth referred to as cells). Subsequently, we

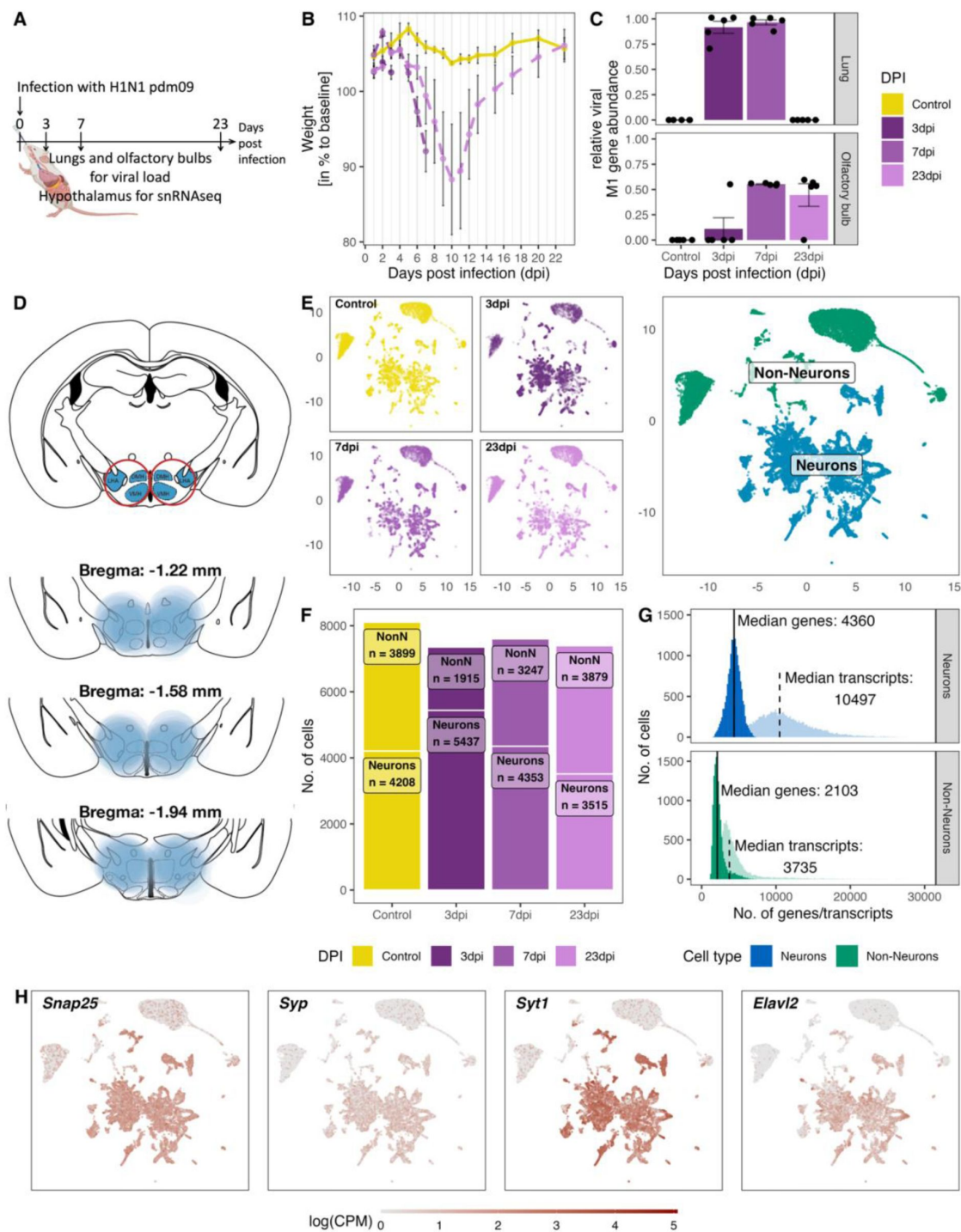


Figure 1.

Overview of the experiment, development of the infection, and the resulting snRNA-seq dataset.

A. Schematic representation of the experiment. **B.** Loss of body weight of mice during disease progression until full recovery ($n = 5$ per group). **C.** Viral M1 gene abundance in lung and olfactory bulb tissue at the 3 time points during disease progression and in controls. **D.** Location of microdissection punches of the hypothalamus mapped to the mouse brain atlas at distances from bregma of -1.22 , -1.58 and -1.94 . **E.** UMAP dimensional reduction of 30,452 cells, color-coded based on their sample group (different time points and control) membership (left) or their neuronal and non-neuronal identity (right). **F.** Bar graph showing the cell counts of neuronal and non-neuronal cells in the different sampling groups. **G.** Histograms depicting the distribution of transcript (lighter shading) and genes (solid shading) in all non-neuronal and neuronal cells. **H.** Normalized expression of neuronal marker genes (*Snap25*, *Syp*, *Syt1*, *Elavl2*) in all cells shown on a UMAP plot.

filtered for viable cells and removed duplicates as well as cells with high contamination of ambient mitochondrial RNA (mtRNA), which left us with a total 30,452 cells for downstream analyses. We separated the cells into neuronal and non-neuronal (NonN) cells by using known marker genes expressed in neurons (*Snap25*, *Syp*, *Syt1*, *Elavl2*; **Figure 1F**, H). As expected, NonN cells expressed in general only half the number of genes (median unique gene count: 2,103) compared to neuronal cells (median unique gene count: 4,360). Similarly, the median transcript count per cell in the NonN population was lower (3,735 transcripts/cell) compared to the median number transcripts in neuron (10,497 transcripts/cell, 4360 features/cell; **Figure 1F**). The ratio between NonN and neuronal cells was similar at all time points except 3 dpi, where we detected a greater number of neuronal cells than NonN cells (**Figure 1G**). In addition, we performed an case-control analysis developed for comparative scRNA-seq studies (Petukhov et al., 2022). Cluster based analysis revealed a shift in cell densities in non-neuronal populations only in the 3 dpi cluster (Supplementary Figure 15), possibly caused by the difference in captured nuclei at 3 dpi (**Figure 1F**). The cause of the difference in cell numbers stays unclear, however could be attributed to inconsistencies in the punching itself (Supplementary Figure 3) and the amount of tissue used for the nuclei isolation.

Identified NonN cell types

Among the non-neuronal cell types, we were able to identify 9 distinct populations, discriminated by their sets of cell type-specific markers (**Figure 2A**, B). We used published datasets of different regions of the hypothalamus to confirm the identity of the distinct cell clusters (Campbell et al., 2017; Chen et al., 2017; Mickelsen et al., 2019; Mickelsen et al., 2020; Romanov et al., 2017). Two of the clusters belonged to the oligodendrocyte (OL) lineage, including oligodendrocyte precursor cells (OPCs, NonN_3) and mature oligodendrocytes (MOLs, NonN_1). The main marker genes in these two clusters were *Lhfp13* and *Slc24a2* but they also expressed other typical marker genes as *Pdgfra*, *C1ql1*, *Fyn* (OPCs) or *Olig1*, *Anln*, *Plp1* in MOLs. Further, we resolved a cluster of astrocytes (NonN_2), which was defined by the differential expression of *Slc4a4* and which was also positive for *Sox9*, *Gja1* and *Agt*. The nearby cluster NonN_7 expressed similar characteristics as the astrocytes and exhibited a high expression of *Slit2*, *Col23a1*, and *Slc16a2* which have been described to be enriched in tanocytes (Chen et al., 2017) (**Figure 2B**, D, and Supplementary Table 2 & 4, Supplementary Figure 5). The identity of this cluster was confirmed by label transfer of other snRNA/scRNA-seq datasets from the hypothalamus, which showed a high prediction score of this cluster as tanocytes (Supplementary Table 2 and 3, Supplementary Figure 8). The distinct clusters NonN_5 (*Ranbp3l*+, *Cped1*+, *Slc6a20a*+) and NonN_9 (*Bnc2*+, *Slc47a1*+, *Slc26a7*+) were identified as vascular and leptomeningeal cells, respectively. The NonN_8 cluster expressing *Slco1a4* is expressing additionally *Ebf1*, *Flt1*, *Rgs5*, which are described as pericyte marker genes (Pagani et al., 2021; Zeisel et al., 2018). The cluster NonN_6 is a microglia cluster, with expression of *Hexb*, *C1qa* and *C1qc* - known microglia marker genes. Best predictions scores for cell type label transfer were found for non-neuronal cells, with mean prediction scores 0.8 comparing the HypoMap C7 cell type levels (Supplementary Figure 5) (Steuernagel et al., 2022). Lastly, even though the cluster NonN_4 (*Syt1*+) is expressing general neuron markers, we could not identify any discriminatory markers for inhibitory or excitatory neuron populations (**Figure 2D**). Discriminatory marker genes such as *Slc17a7*, *Crym*, *Ptk2b*, *Itcam5*, *Slit3*, *Crym* indicate a progenitor/neuroblast identity (Kostin et al., 2021; Zeisel et al., 2018). Aligning this dataset with a dataset from the developing dentate gyrus including cells from the neuroblast lineage, indicates the presence of an immature granule subcluster strengthening the idea of NonN_4 as a neuroblast cluster (Supplementary Figure 8) (Hochgerner et al., 2018). However, because of the general low confidence and weak alignment we have kept this cluster unassigned.

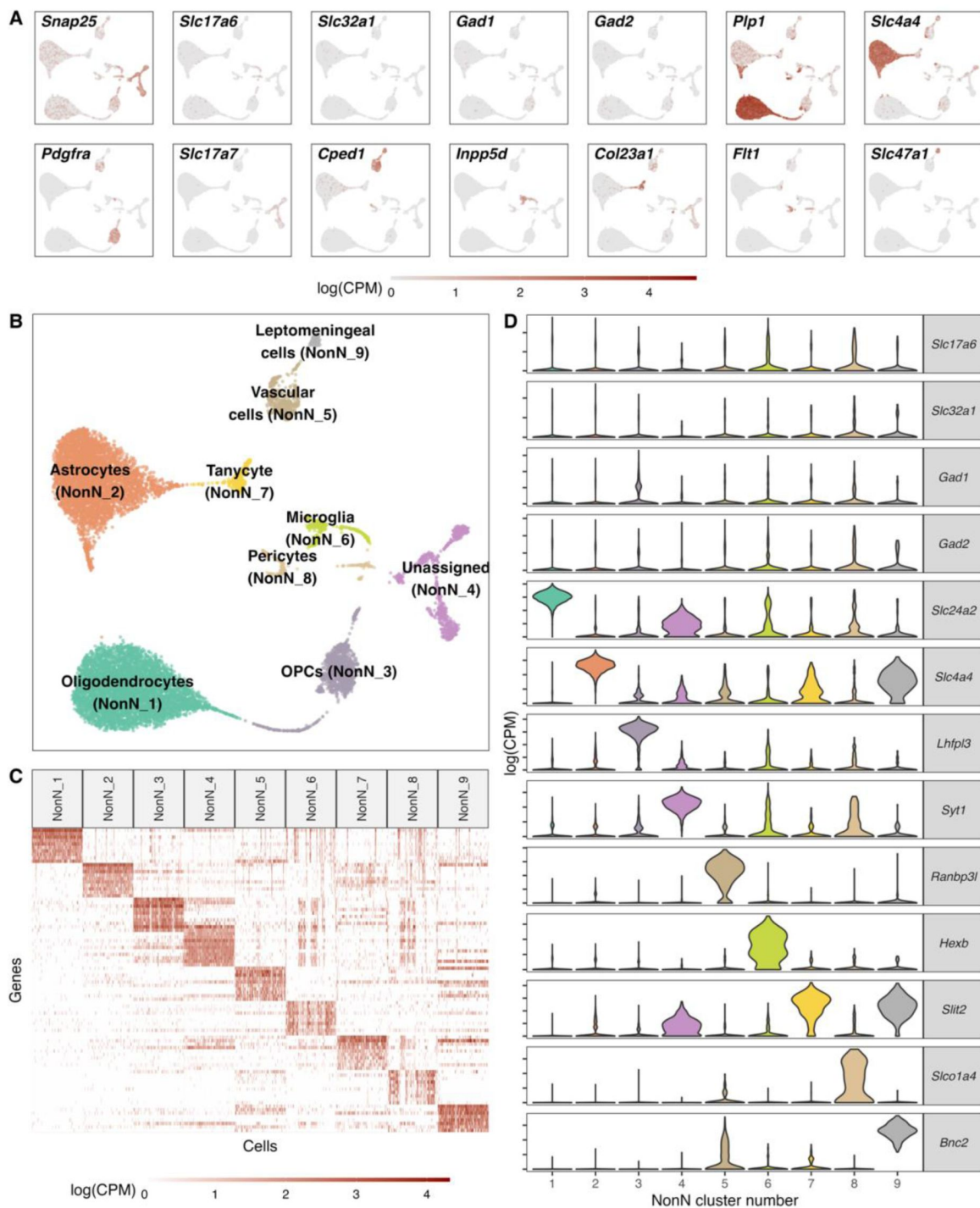


Figure 2.

Classification of non-neuronal cell types.

A. Normalized expression values of different neuronal and non-neuronal cell type markers (pan-neuronal marker - *Snap25*; Glutamatergic marker - *Slc17a6*; GABAergic markers - *Slc32a1*, *Gad1*, *Gad2*; Oligodendrocyte marker - *Plp1*; Astrocyte markers - *Slc4a4*; Oligodendrocyte precursor marker - *Pdgfra*; *Cped1* - Vascular and leptomeningeal cell markers - *Cped1*, *Slc47a1*, Microglia marker - *Inpp5d*, Tanycytic marker - *Col23a1*, Pericyte marker - *Flt1*) in all non-neuronal cells ($n = 12,940$) mapped on a UMAP. **B.** Unsupervised clustering of non-neuronal cell types shown in a UMAP embedding and color-coded and annotated by potential cell type annotations. **C.** Heatmap of normalized expression values showing discriminatory cell type markers of 9 non-neuronal cell populations. **D.** Violin plots showing distribution of normalized expression values of neurotransmitters and best discriminatory cell type markers.

Heterogeneous populations of inhibitory and excitatory neurons

To subcluster the neurons, we first examined the expression signature of neuronal markers: Vesicular glutamate transporter 2 (VGLUT2; encoded by *Slc17a6*); Vesicular GABA Transporter (VGAT, encoded by *Slc32a1*), glutamate decarboxylase 67 (GAD67; encoded by *Gad1*); and histidine decarboxylase (encoded by *Hdc*) and separated the neuronal cells in GABAergic and glutamatergic cells (**Figure 3A**). We identified a total of 26 GABAergic and 30 glutamatergic clusters. Within the glutamatergic cells we found also a small population of histaminergic (Glut_24; *Hdc*+) and dopaminergic neurons (Glut_27; *Th*+, *Ddc*+). The histaminergic neurons were left for visualisation and analysis purposes in this cell type, despite them often being described as GABAergic (**Figure 3B**). Among the glutamatergic cells, we were able to detect two small clusters of HCRT expressing neurons (Glut_29; *Hcrt*+) and MCH expressing neurons, (Glut_25; *Pmch*+). The cellular resolution aligns well with the composition of other cell types seen in other studies of the hypothalamus (Campbell et al., 2017; Chen et al., 2017; Mickelsen et al., 2019; Mickelsen et al., 2020; Romanov et al., 2017; Steuernagel et al., 2022). Among well described neuronal cell cluster, we found a good overlap with *Hcrt*+, *Pmch*+, *Hdc*+ and *Prkch*+ neurons (Supplementary Figure 6) and *Pomc*+, *Nkx2-4*+ GABAergic neurons (Supplementary Table 7). Additionally, we identified a *Argp*+/ *Npy*+ co-expressing cell cluster as a sub-cluster of GABA_1 (Supplementary Figure 14).

General transcriptional disease responses within the medial hypothalamus across all cells

To understand the nature of the molecular changes between the control group and the infected groups at different time points of disease progression, we first used a pseudo-bulk approach to compare their transcriptional signatures of all cells per time point against each other. A principal component analysis based on the pseudo-bulk dataset showed a clear clustering of the biological replicates per time point (**Figure 4A**). We found a clear separation of the samples taken at 3 dpi from the other time points. 7 dpi is clustering separately between 3 dpi and the control samples, whereas the samples from the recovery group at 23 dpi are spread between the 7 dpi and the control group cluster. This indicates a clear transcriptional change during the first 7 days of infection. The recovery group is mostly overlapping with the control group, but one sample clusters together with the 7 dpi group, suggesting a slower disease or recovery process in this animal (Animal 23.4, **Figure 4A**). The overall gene expression counts show a pattern of lower counts per cell at 3 dpi in both GABAergic and glutamatergic cell populations. In glutamatergic cells this is a lasting but less pronounced effect until 23 dpi (**Figure 4B**, Supplementary Table 5). Next, we performed a differential gene expression (DGE) analysis comparing the control samples to samples from different time points during disease progression, disregarding any differentiation between cell subpopulations. We tested, based on previous studies, gene sets known to be involved in neuroinflammatory and immune regulatory process within the central nervous system (CNS) for possible changes in gene expression (Dusedau et al., 2021; Hosseini et al., 2018; Sadasivan et al., 2015; Zielinski et al., 2013). We primarily detected a down-regulation of these gene categories, which was strongest at 3 dpi with only very few significant changes lasting until 23 dpi (**Figure 4C**).

The only up-regulated genes were detected at 3 and 7 dpi belonging to the category of neuron-related factors. Genes in the other categories were consistently down-regulated or not significantly regulated at all. This also included the virus-related cytokines and IFN-response genes (i.e. *Il1a*, *Ifih1*, **Figure 4C**) at 3 dpi. Toll-like receptors (TLRs) are central in initiating the innate immune response by orchestrating signalling pathways leading to the release of inflammatory cytokines (Kawai & Akira, 2011). In our dataset we see a general downregulation of TLRs (*Tlr2*, *Tlr3* and *Tlr4*, **Figure 4C**). These results show that a peripheral H1N1 pdm09 infection cause decreased gene expression in non-neuronal, GABAergic, and glutamatergic cell populations in the hypothalamus affecting several immune related processes.

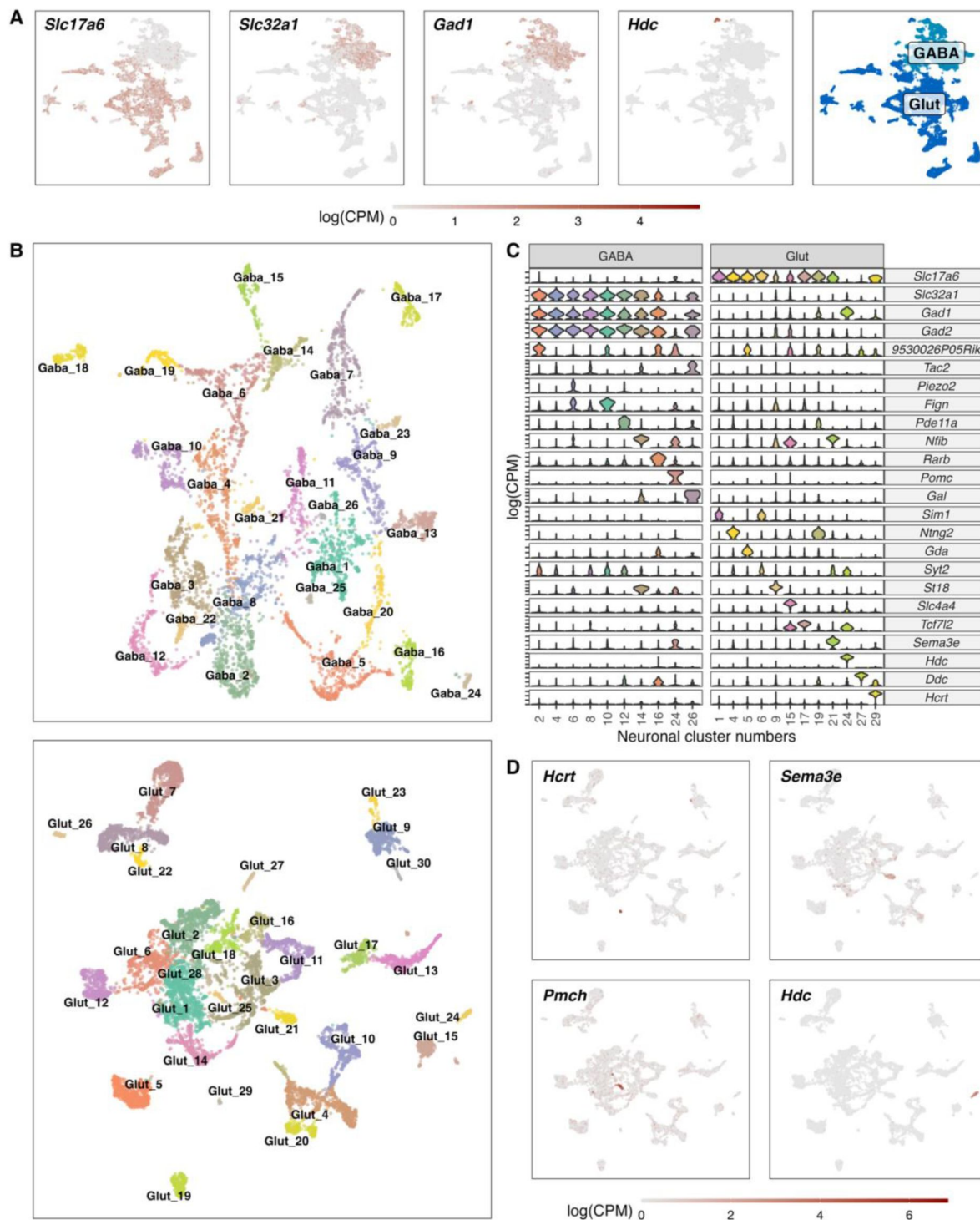


Figure 3.

Classification of GABAergic and glutamatergic neuronal cell types in the hypothalamus.

A. Normalized expression values of different neurotransmitters (Glutamatergic marker - *Slc17a6*; GABAergic markers - *Slc32a1*, *Gad1*, *Gad2*) in all neuronal cells (left) and a color-coded UMAP projection based on their GABAergic or glutamatergic identity (right) ($n = 17,512$). **B.** Un-supervised clustering of GABAergic (upper, $n = 6,032$) and glutamatergic (lower, $n = 11,481$) cells in UMAP plots. Cell type clusters are color-coded and annotated with labels. **C.** Violin plots showing normalized expression values of neurotransmitters and discriminating marker genes of selected GABAergic (left) and glutamatergic (right) cell type clusters. **D.** Glutamatergic UMAP plots showing normalized expression values of distinct markers for hypothalamic neuron populations (HCRT neurons - upper left, PMCH neurons - lower left, GnRH neurons - upper right, Histaminergic neurons - lower right).

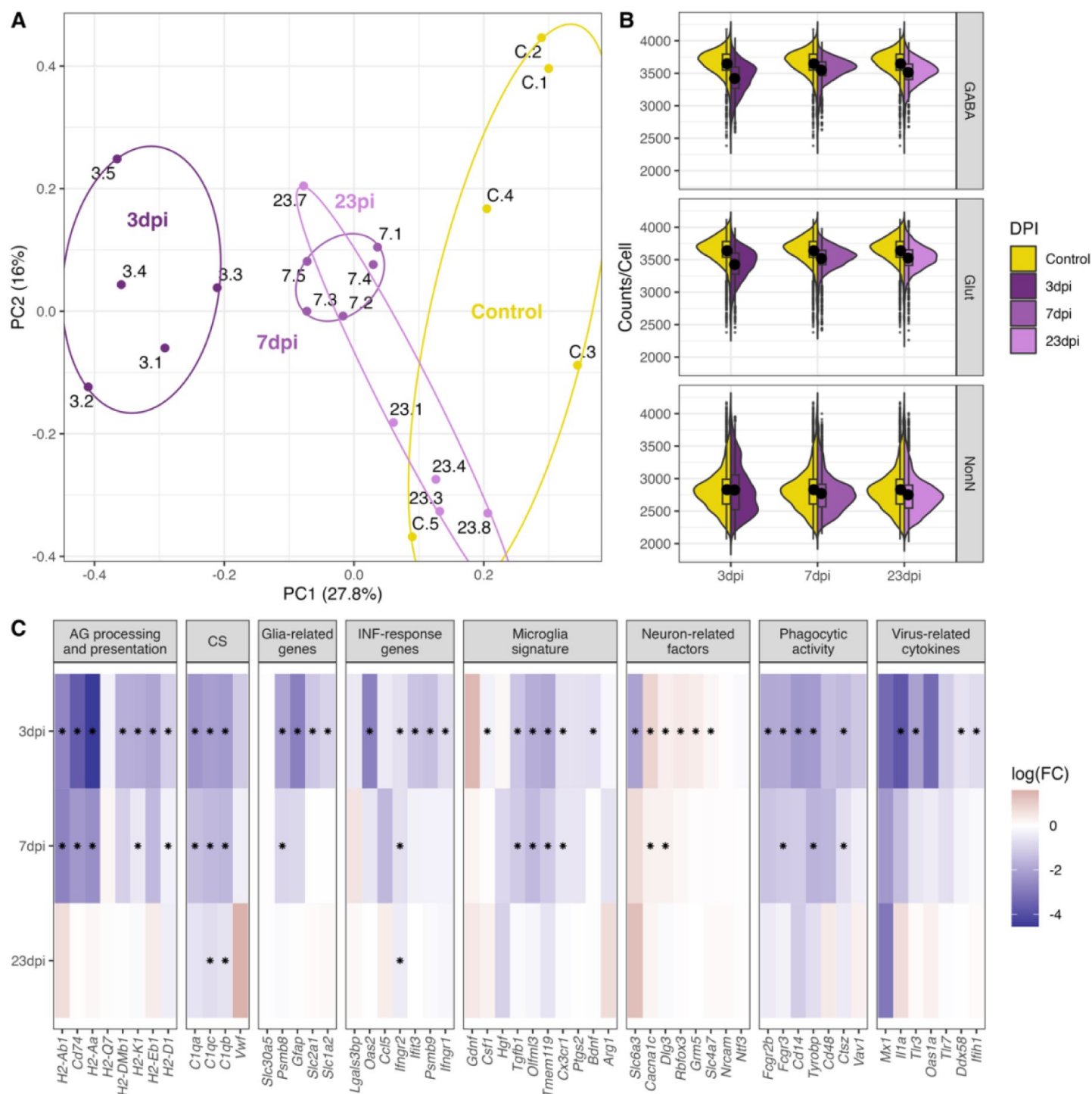


Figure 4.

General changes in the transcriptomic landscape of the hypothalamus during peripheral IAV infections.

A. Principal component plot of combined counts per sample. Each sample snRNA-library were down-sampled to a total of 300 cells (100 non-neuronal, 100 GABAergic, 100 glutamatergic cells) before normalized counts were aggregated before principal component analysis. Samples are color-coded according to their group membership and the numbers correspond to the individual animal IDs. **B.** Violin plot showing the distribution of counts per cell at the different time points. The three points of infections are compared to the control group. **C.** Heatmap showing log-transformed log-fold changes of known immune-related genes. Black stars indicate significant expression (FDR ≤ 0.05).

Specific molecular regulation in non-neuronal, glutamatergic, and GABAergic cell populations

Next, we resolved the molecular disease responses in better detail, by performing a differential gene expression (DGE) analysis on the three overall cell populations (GABAergic, glutamatergic, and NonN). Similarly, to the previous pseud-bulk approach, we detected the strong down-regulation of genes at 3 dpi in all the three distinct cell populations (**Figure 5A** [↗](#)). The overall number of down-regulated genes decreased at 7 and 23 dpi, but more differentially expressed genes were detected in the GABAergic cells at 23 dpi, than at 7 dpi (**Figure 5A** [↗](#) and **5B** [↗](#)). Of all highly regulated genes at 3 dpi (False discovery rate (FDR) ≤ 0.05 , log fold change (logFC) > 1 or logFC < -1), approximately a quarter (27%) of the genes were shared between the 3 cell populations, and 30% were shared between the two neuronal clusters. Around 10% of the genes expressed in the GABAergic and glutamatergic cell clusters at 3 dpi were found expressed only in these cells (**Figure 5B** [↗](#)). In contrast, 20% of the DEGs in the NonN cells at 3 dpi were unique to these cells only. At 7 dpi, only 4 genes were shared between all 3 clusters, and 59 genes between the two neuronal clusters (**Figure 5B** [↗](#)). At 23 dpi no more DEGs were detected in the NonN cells. Most differentially expressed genes (DEGs) at this time point belonged to the GABAergic cluster. To exclude if persistent transcriptional effects reported in neurons but not in non-neurons are caused by the different amount of captured transcripts between neurons and non-neurons (**Figure 1G** [↗](#)), we created a down-sampled dataset with similar transcripts per cell in neurons and non-neurons (Supplementary Figure 12B). Here we reported the same effect, of high down-regulation of genes in all cells (Supplementary Figure 12A) at 3 dpi, with a less pronounced pattern at 7 dpi. In the recovery group at 23 dpi, we found again a transcriptional response only in the neuronal cells but not in non-neuronal cells (Supplementary Figure 12A). To study what functional effects these detected changes in gene expression might have, we performed a gene ontology enrichment analysis. **Figure 5C** [↗](#) shows all the significantly regulated pathways. Of special interest, we detect an enrichment of genes involved in immune responses at 3 dpi in all three clusters, while this is restricted to the neuronal clusters at 7 dpi and only the Glut cluster at 23 dpi (**Figure 5C** [↗](#)). Further, enrichment of neuropeptide signalling, and hormone activity pathways are also present in all three clusters at 3 dpi, while lasting until 23 dpi only in the neuronal cells. These data suggest that long term effects in the hypothalamus after a peripheral IAV infection affects primarily neurons.

Dissecting immune system processes in microglia

We tested all identified NonN and neuronal cell populations for differential gene expression. A cell cluster was excluded from analysis if their cells were originating from less than 3 of the biological replicates and if they were composed of less than two cells per cluster per biological replicate. Similarly, we used for further downstream analysis only strongly expressed genes with high up-or down-regulation (FDR ≤ 0.05 , logFC > 1 or logFC < -1). As shown before, the strongest regulatory effect on the different clusters' transcriptomes were detected at 3 dpi, which lasted partially until 23 dpi (**Figure 6A** [↗](#)). Several GABAergic clusters down-regulated more genes at 23 dpi than at 7 dpi. These cell populations (GABA 2, 4, 6, 8, 10, 12) are aligning close to each other in the dimensional reduction (**Figure 3B** [↗](#)) and are overlapping in the centre of the UMAP. The glutamatergic neuron populations show in general a similar pattern, with the strongest transcriptional response at 3 dpi and less differentially regulated genes at 7 and 23 dpi.

As described in the previous paragraph, we identified immune response pathways as one of the most enriched gene ontologies (GO) terms in the dataset. Therefore, we selected only the genes annotated in these immune-related GO terms, leaving us with 22 genes. Interestingly, all these 22 immune related genes are regulated in microglia (NonN_6) at 3 and 7 dpi (**Figure 6C** [↗](#)). In other non-neuronal clusters less than half of this genes are differentially regulated at 3 dpi and none at 7 or 23 dpi. Similarly, neuronal cell populations showed fewer genes regulated at 3 dpi than microglia, however especially some GABAergic cell types show a transcriptional response of

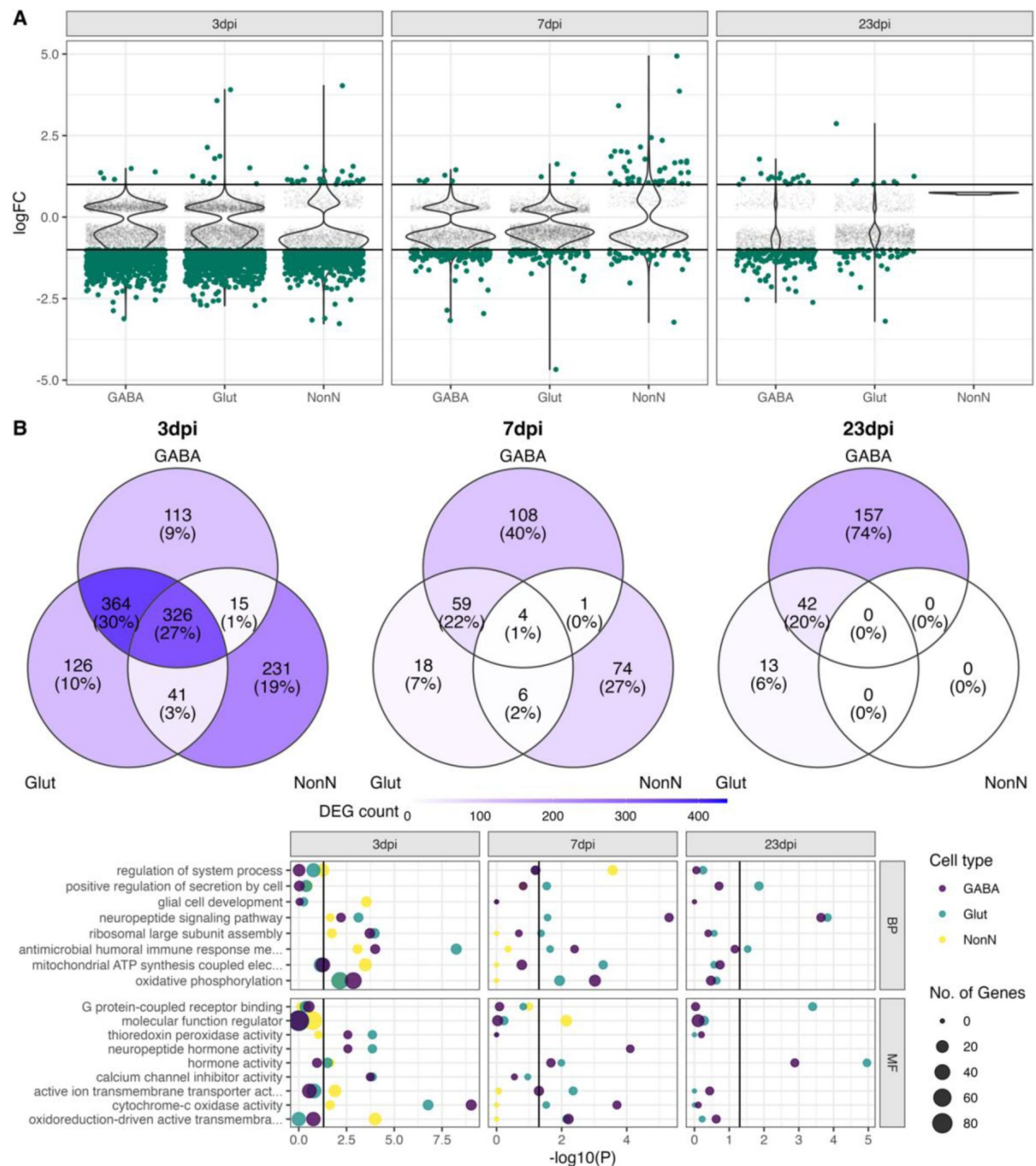


Figure 5.

Transcriptional changes in GABAergic and glutamatergic neurons of the hypothalamus during peripheral IAV infections.

A. Violin plots showing the log-transformed fold changes (logFC) per gene calculated for the GABAergic, glutamatergic, and non-neuronal cells at 3, 7, and 23 dpi. Dark-green dots represent significantly differential expressed genes (FDR ≤ 0.05) with a logFC greater than 1 or lower than -1. Small grey dots show significantly expressed genes with a logFC between -1 and 1. **B.** Venn diagrams depicting the overlap of highly differential expressed genes (logFC > 1 or logFC < -1) between the three main cell type clusters at each time point. **C.** Gene ontology analysis showing the most significant ontology terms for the category of Biological Process (BP, top) and Molecular Function (MF, bottom). The two highest significant ontology's for each time point and each cell type were chosen if it included more than 4 annotated genes. The size of the dots indicates the number of genes included in the ontology term and the colour show the cell type annotation.

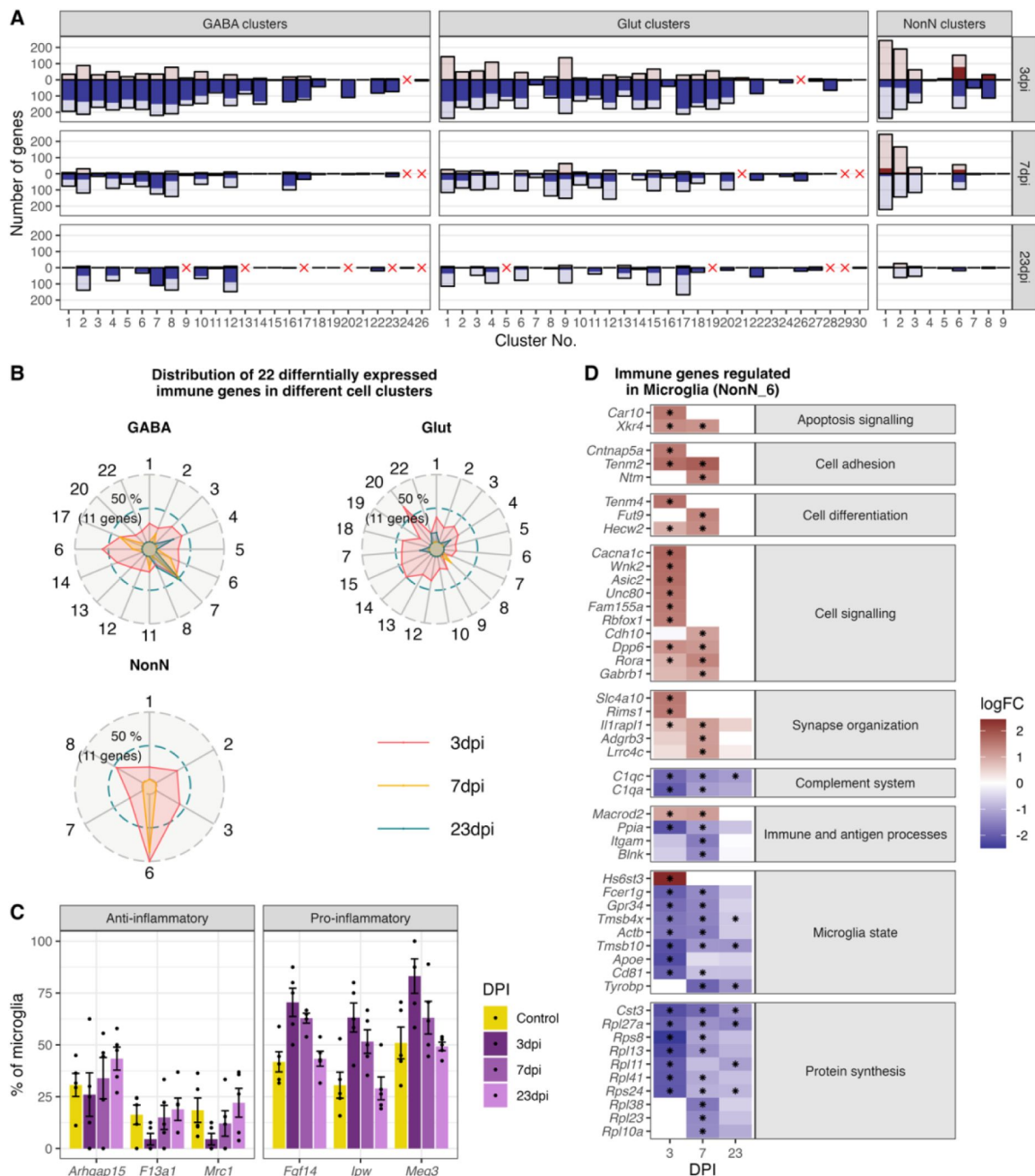


Figure 6.

Gene expression changes in different cell type clusters and microglia activation during peripheral IAV infections.

A. Bar plot showing the number of significantly differentially expressed genes in each cell type cluster at 3, 7 and 23 dpi. Dark solid blue and red bars show the number of highly differential expressed gene ($FDR \leq 0.05$, $\log FC \leq -1$ or $\log FC \geq 1$) per cell type cluster and time point. Lighter shaded bars depict the number of all significant regulated genes ($FDR \leq 0.05$, $\log FC \geq 0$ or $\log FC \leq 0$). Differentially expressed genes (DEGs) were only included for clusters containing at least 3 cells per sample and per time point in the cluster. A red cross indicates instances where these criteria were not met. **B.** Radar plot showing the number of significantly and strongly regulated genes ($FDR \leq 0.05$, $\log FC \geq 1$ or $\log FC \leq -1$; $n = 22$) involved in immune processes (based on their gene ontology annotations) per cell cluster at the different time points (red: 3 dpi, yellow: 7 dpi, blue: 23 dpi). **C.** Percentage of the number of cells expressing distinct anti-or pro-inflammatory signature genes in the microglia cluster. Cells with a raw count of 0 for a gene were assumed as non-expressing, all other cells were assumed to express this gene. Shown are percentages of microglia cells expressing a gene compared to all microglia at each time point across all samples. **D.** Relative changes in expression levels of disease associated genes within the microglia cluster. Data is shown as log-transformed fold changes ($\log FC$) of each time point compared with control group. Black stars depict significant differentially expressed genes ($FDR \leq 0.05$).

immune genes until 7 and even 23 dpi (i.e., *GABA_7*, 16 and 17). The here reported highest regulation of immune genes indicate that microglia are also in the medial hypothalamus the main immune orchestrating cell population.

We next aimed to describe the transcriptional changes in the microglia during disease progression. Raw DEG numbers emphasize the microglia cluster as the only cell cluster with a strong upregulation of genes at 3 dpi (75 genes), and a long-lasting down-regulation of genes until 23 dpi (52 and 12 genes at 7 and 23 dpi respectively; **Figure 6A** [↗](#), cluster NonN_6), indicating the importance of microglia for immune homeostasis in the medial hypothalamus. A closer look at the highest up-regulated genes reveals a strong induction of genes involved in cell adhesion, signalling and differentiation, synapse organization as well as apoptosis signalling. Down-regulated genes participate in the complement system, immune and antigen processing, microglia state genes and ribosomal genes contributing to protein synthesis (**Figure 6E** [↗](#)).

Typical markers like cell surface proteins or intracellular markers are often used to identify pro- (M1 state; i.e., *Il1b*, *Ifng*, *Tfna*, *Fcgr3*) or anti-inflammatory (M2 state; i.e., *Tgfb*, *Mrc1*, or *Arg1*) states of microglia (Jurga et al., 2020 [↗](#); Moehle & West, 2015 [↗](#)). These markers were not regulated during disease progression in our dataset. Discrimination of the activation state of microglia using snRNA-seq has been reported to be challenging, due to a depletion of certain marker genes in the nucleus especially in human tissue (Thrupp et al., 2020 [↗](#)). Because of this lack of typical M1 and M2 markers, we identified alternative DEGs in microglia at different the time points related to activation states. Two long non-coding RNA (lncRNA) genes: Maternally Expressed 3 (*Meg3*), Imprinted in Prader-Willi Syndrome (*Ipw*) and the gene Fibroblast Growth Factor 14 (*Fgf14*) which have been recently connected to immune activation and pro-inflammatory microglia states (Liu et al., 2021 [↗](#); Meng et al., 2021 [↗](#); Senatorov et al., 2019 [↗](#)), were found in our dataset enriched in the microglia at 3 and partly at 7 dpi (**Figure 6D** [↗](#)). In contrast, genes connected to an anti-inflammatory state (*F13a1*, *Mrc1*, *Arhgap14*), were expressed in less microglia cells during acute infection than in control and 23 dpi. The results show that microglia are also in the medial hypothalamus a main immune orchestrating cell population, shifting from their homeostatic state towards a pro-inflammatory microglia population during 3 and 7 dpi. This shift is completely resolved after recovery at 23 dpi.

Identification of Oligodendrocyte and Astrocytes subclusters at 7 dpi

In oligodendrocytes (NonN_1) and astrocytes (NonN_2) between 144 – 244 genes were up-or down-regulated ($\log_{2}FC > 0$ or $\log_{2}FC < 0$) at 3 and/or 7 dpi (**Figure 6A** [↗](#)). This was accompanied by a shift of the cells in the dimensional reduction plot of the not integrated data, especially at 7 dpi (**Figure 7A** [↗](#)). In the oligodendrocytes this was accompanied by an up-regulation of genes involved in stress-related and glucocorticoid pathways (**Figure 7B and C** [↗](#)). The same was not seen in the astrocyte sub-cluster. We identified significantly regulated GO terms associated with transport pathways in oligodendrocytes, with a strong up-regulation of genes in these terms at 3 and 7 dpi (**Figure 7D** [↗](#)). Only mitochondrial transport processes and transmembrane transport included down-regulated genes. Genes associated with GABAergic transport was up-regulated in oligodendrocytes uniquely at 7 dpi. Marker genes for the astrocytic 7-dpi cluster were identified as *Aldh1a1*, *Lrrc8a* (annotated here as ENSMUSG00000099041) and *Zfhx3*. *Zfhx3* showed a very distinct expression in this cluster (**Figure 7C** [↗](#)). For astrocytes, the GO term oxidoreductase activity was highly and uniquely enriched at 7 dpi (Supplementary Table 8). Accordingly, we found the oxidoreductase genes to be highly up-regulated in the 7 dpi cluster, with only a few being also regulated at 3 dpi and non at 23 dpi (**Figure 7E** [↗](#)). Together these results suggest that an IAV infection induces slowly evolving transcriptional changes in oligodendrocytes and astrocytes, leading to the appearance of distinct sub-populations of these cell types at 7 dpi. This altered cell stated disappear again after recovery at 23 dpi.

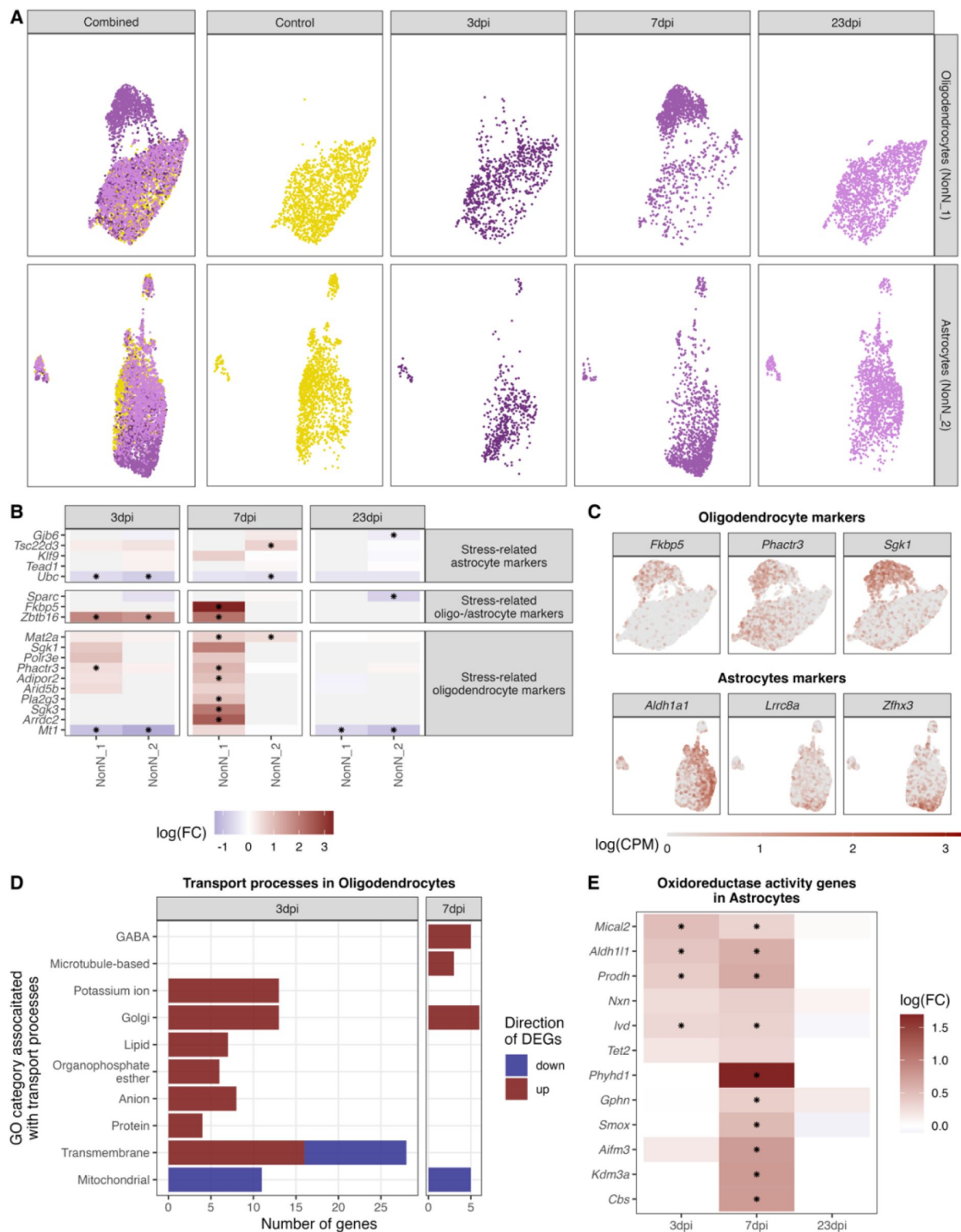


Figure 7.

Identification of distinct oligodendrocyte and astrocyte sub-clusters at 7 dpi.

A. Combined UMAP plot of all cells of the oligodendrocyte (upper right, $n = 5.023$) and astrocyte (lower right, $n = 4.542$) cluster in controls and at different time points. **B.** Relative gene expression changes (in comparison of to the mock-infected group) of stress-related oligodendrocyte and astrocyte markers within the oligodendrocyte (NN_1) and astrocyte (NN_2) subcluster. Data is shown as log-transformed fold changes, an asterisks indicates statistical significance ($FDR \leq 0.05$). **C.** Normalized expression values of selected marker genes at 7-dpi in UMAP plots of all oligodendrocytes or astrocytes. **D.** Number of differentially expressed genes ($FDR \leq 0.05$, $\log FC \leq -1$ or $\log FC \geq 1$) annotated to gene ontologies associated with transport process in the oligodendrocytes at 3 and 7 dpi. **E.** Expression dynamics of differential expressed oxidoreductase genes in the astrocyte cluster at 3, 7 and 23 dpi. A black star indicated significant differential expression ($FDR \leq 0.05$).

Long-term transcriptional response in neurons following as IAV infection

Even though the most prominent feature of the hypothalamic transcriptional response to H1N1 pdm09 was the downregulation of genes at 3 dpi across all cell clusters, some neuronal cell populations showed long-term differential expression of genes. As described in previous paragraphs, we detected overall changes in both GABAergic and glutamatergic neurons at 23 dpi (**Figure 5B**). When looking at neuronal subclusters, we found that 6 out of 26 GABAergic cell clusters and 17 out of 30 glutamatergic cell clusters contained significantly regulated genes at 23 dpi when compared to the control group (**Figure 6A**). The cluster based case-control analysis (Cocoa) showed a similar dysregulation of neuronal and non-neuronal cell populations at 3 and 7 dpi (Supplementary Figure 16). However, the long-lasting transcriptional changes described before, were not reproduced with this cluster-based analysis. Though, analysis of cluster-free expression shifts shows dysregulation of neuronal and non-neuronal cells at the acute phase of the infection (Supplementary Figure 18-20), however detects mainly transcriptional changes in neurons at 23 dpi (Supplementary Figure 17 & 208). Our initial GO term enrichment analysis (**Figure 5C**) showed several different pathways being affected at all timepoints, so we aimed at dissecting this further by analysing individual neuronal cell clusters. In this analysis we included glutamatergic and GABAergic cell clusters, with a high DGE profile and at least 10 genes exhibiting a logFC greater than 1 or less than -1. The 15 neuronal clusters fulfilling this criterion were: Gaba_2, 4, 6, 8, 10, 12 and Glut_1, 4, 6, 9, 11, 15, 17, 18 and 20. In general, we found the same regulatory processes to be regulated across many of the neuronal cell types. When looking at the pathways that remain altered at 23 dpi these include translational processes in the cytosol, metabolic processes in the mitochondria, as well as apoptotic processes (i.e., mitochondrial respiration and TCA cycle) (**Figure 8**, Supplementary Table 7 & 8).

Our analysis shows that many of the neuronal subpopulations respond with a similar pattern when the organism is challenged by an infection. To determine whether there would also be differences in response pattern between neuronal subtypes, we selected six hypothalamic neuronal subtypes with known regulatory functions in metabolism: Agrp⁺/Npy⁺, Lepr⁺, and POMC⁺ neurons; and sleep: Hdc⁺, Pmch⁺, and Hcrt⁺ neurons. With both feeding and sleep patterns changing during influenza, we would expect these neuronal subtypes to change their gene expression during the experiment. This is indeed what we find. While most of the subtypes respond with overall downregulation of gene expression (**Figure 8C-D**), the POMC⁺ neurons show a different pattern with equal and strongly both up and downregulated genes.

Taken together, neuronal populations were the only cell populations that were affected by the H1N1 pdm09 infection until recovery, with changes including inhibition of genes contributing to metabolic and translational processes in the neurons. While most neuronal subpopulations follow this same regulatory pattern, POMC⁺ neurons show bigger and more unique changes in gene regulation following a peripheral influenza infection.

Discussion

The compiled snRNA-seq dataset presented above, to our knowledge, represents the first comprehensive transcriptome profiling on single-nuclei level of the hypothalamus during a peripheral IAV infection. We detected 26 GABAergic, 30 glutamatergic and 9 non-neuronal cell types, aligning closely with previously published datasets of the hypothalamus (Campbell et al., 2017; Chen et al., 2017; Mickelsen et al., 2019; Mickelsen et al., 2020; Romanov et al., 2017; Zeisel et al., 2018). In addition, we deconvoluted the molecular changes of distinct cell populations in the hypothalamus during a peripheral IAV infection and detected a whole range of

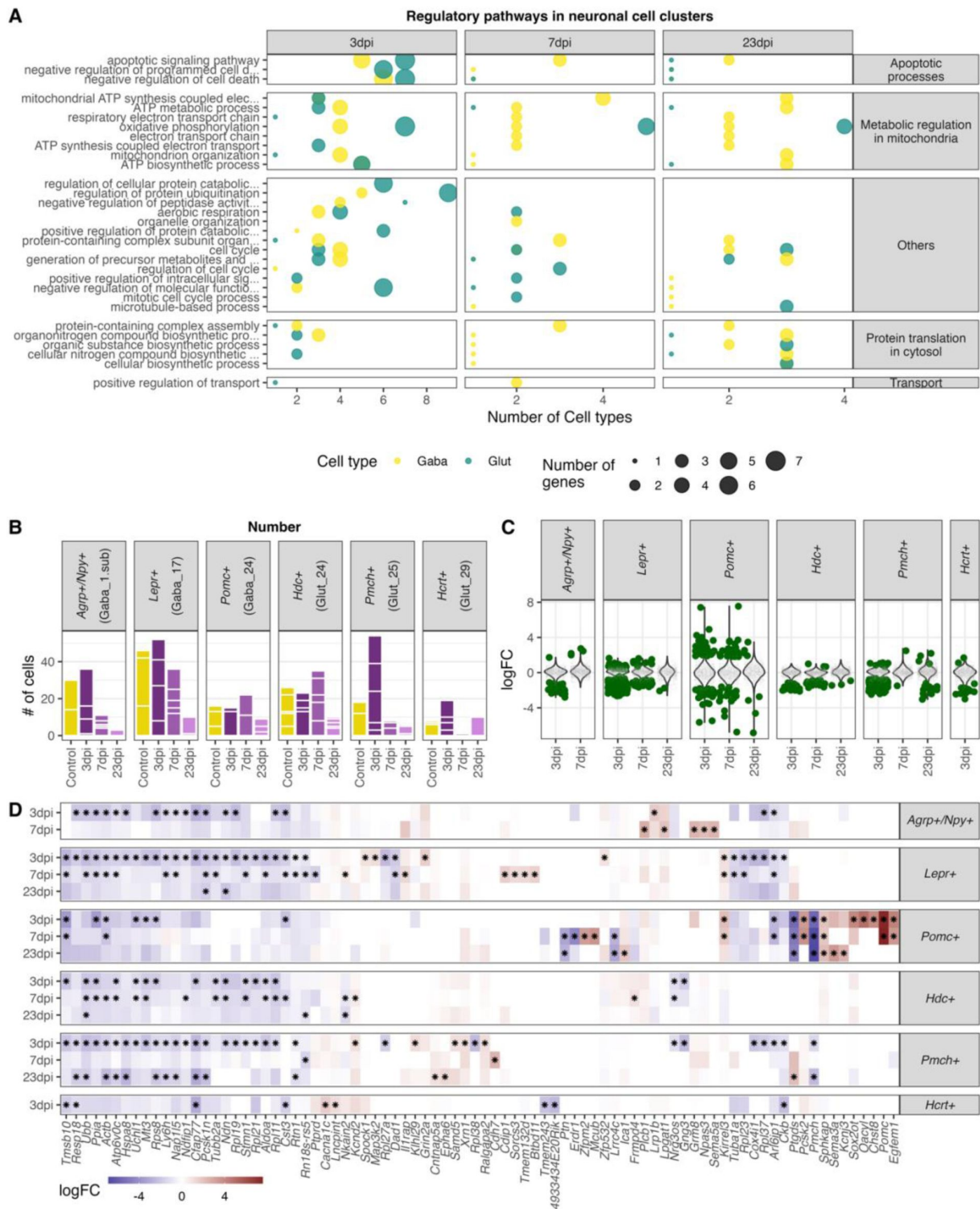


Figure 8.

Highly enriched gene ontology categories in neuronal cell populations at different time points of infections.

A. Dot-plot depicting the number of neuronal cell populations significantly enriched gene ontology categories. Only cell clusters with at least 10 significantly differentially expressed genes ($\log_{2}FC \leq -1$ or $\log_{2}FC \geq 1$, $FDR \leq 1$) in all three analysed time points were included in the here presented plot. Neuronal cell clusters included are (GABA_2, 4, 6, 8, 10, 12; Glut_1, 3, 4, 6, 9, 11, 13, 15, 17). Only significantly enriched ($P_{\text{lim}} \leq 0.05$) gene ontology terms were chosen and for each cluster per time point the 10 most enriched GO terms were included. **B.** Bar plot depicting the amount of captured nuclei in the individual animals at the different time points for known hypothalamic neuron populations. **C.** Differential expressed genes in selected neuron populations. Dark green dots show significantly differential expressed genes ($FDR \leq 0.05$). **D.** Heatmap showing the five highest and lowest DEGs ($FDR \leq 0.05$) in each neuronal population.

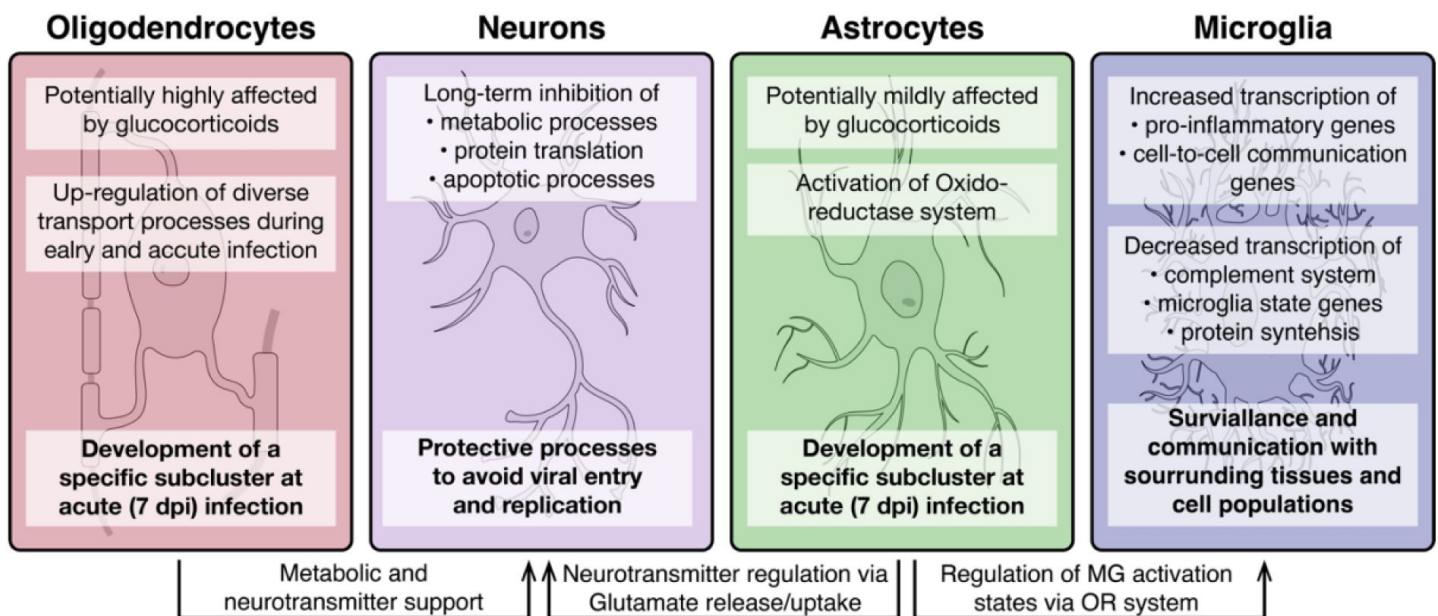


Figure 9.

Overview of molecular processes occurring in different cell types within the hypothalamus during a peripheral H1N1 IAV infection.

Schematic overview of the main molecular mechanisms of oligodendrocytes, astrocytes, microglia and neurons during acute to late immune responses of a peripheral IAV infection with the H1N1 pdm09 Influenza A virus.

transcriptional changes across both NonN and neuronal cell types. In addition, we believe our dataset is a valuable resource for the scientific community to explore unsolved questions of hypothalamic responses during peripheral infections.

We were able to report viral infection of the olfactory bulb using H1N1 pdm09. This is comparable to what has been seen in previous studies using the non-neurotropic H1N1 strain PR8 (Majde et al., 2007 [DOI](#); Zielinski et al., 2013 [DOI](#)). The same studies found replication intermediates of the viral nucleoprotein (NP1), suggesting the potential ability of the virus to replicate further within the OB. However, another mechanism has been suggested, in which the virus continuously replicate in the nasal epithelium and then gets transported into the OB (Zielinski et al., 2013 [DOI](#)). Other areas deeper in the CNS, like the hippocampal region, did not show any viral infiltration by PR8. (Hosseini et al., 2018 [DOI](#); Jurgens et al., 2012 [DOI](#)). Similar results were reported for infections with H1N1 pmd2009, where the virus could not be detected in the hippocampus, striatum or cortex, supporting the hypothesis that this virus is not replicating in the CNS (Sadasivan et al., 2015 [DOI](#)).

In contrast to our results, studies using the PR8 disease model reported elevated mRNA expression levels of cytokines related to viral infections 24 hours and 6 days post infection (Zielinski et al., 2013 [DOI](#)). We found in general a strong down-regulation of genes at 3 dpi, including also virus-related cytokines and IFN-response genes (i.e., *Tlr3*, *Il1a*, *Ifih1*) at 3 dpi with no differential regulation of these transcripts at 7 dpi. This indicate that we do not have neuroinflammatory processes in the hypothalamus in our model, or that the here chosen time point of 3 dpi might be too late to capture innate immune activation in the CNS.

Even though we used different published datasets to annotate the here-in identified cell types or compare gene expression changes in a disease context, the comparability of these datasets is challenging due to differences in the experimental setups. We identified a lack of RNA sequencing studies focusing solely on molecular changes in the hypothalamus during non-neurotropic IAV challenges. Different molecular changes based on viral strain or between brain regions are expected and further studies are needed to resolve this.

A prominent feature of our dataset is a long-lasting down-regulation of gene transcripts until 23 dpi in many neuronal cell populations. Gene set enrichment analysis showed these to be mainly related to processes in mitochondria (i.e. citric acid cycle (TCA), ATP synthesis) or to protein translation in the cytosol. This could be interpreted as a defensive mechanism, for the neurons to escape viral entry or replication in the cell and hence protect the cells from apoptosis. Different intermediates of the citric acid (TCA) cycle can be important for viral replication within the cell. In the context of IAV, acetyl-CoA and ATP are known to be beneficial for viral replication. Acetyl-CoA is also used for viral acetylation and ATP is needed for budding of the virus (Hatakeyama et al., 2018 [DOI](#); Hui & Nayak, 2001 [DOI](#); Sanchez-Garcia et al., 2021 [DOI](#)). The downregulation of genes contributing to the translational machinery could be another attempt to inhibit translation of viral proteins in case of a potential entry of the virus into the cell. Another explanation for our findings could be that the downregulation of metabolic and translational processes in the cell might be caused by the excessive weight loss over the disease course and the lack of energy resources. However, we did see these changes also after full weight regain at 23 dpi. In addition, our results indicate that the virus is present in the OB until complete recovery at 23 dpi, suggesting this as one potential trigger for the here reported long-lasting transcriptional changes on hypothalamic neuron populations. Further studies are needed to determine for how long the virus remains in the system and when/if the neurons return fully to the healthy control condition.

A subanalysis of *Agrp*⁺/*Npy*⁺, *Lepr*⁺, *POMC*⁺, *Hdc*⁺, *Pmch*⁺, and *Hcrt*⁺ neurons showed a particularly strong response of the *POMC*⁺ neurons compared to the other neuronal subtypes. *POMC*⁺ neurons have been shown before to be activated during inflammation in various models, a

mechanism that is speculated to be important for development of sickness behaviours (Jin et al., 2016 [DOI](#); Kang et al., 2020 [DOI](#)). This fits well with our observation that the *POMC* gene itself is strongly upregulated at 3 and 7dpi.

Microglia, the resident immune cells of the brain, can have based on their activation state different immunological functions and properties. Interestingly, our results demonstrate the activation of pro-inflammatory genes in the microglia population, whereas anti-inflammatory transcripts are depleted. Further, general microglia signature genes (i.e. *Hexb*, *Cx3cr1* and *Selp1g*) were found down-regulated at 3 and partly at 7 dpi, indicating a clear shift in microglia homeostasis at 3 dpi. This assumption is supported by the activation of genes contributing to cell adhesion, signalling and differentiation as well as synapse signalling. A pro-inflammatory microglia state (M1) is induced during an acute immune response and provide acute defence and repair mechanisms. For this, a tight control via constant cell-to-cell communication is necessary to avoid excessive long-term activation and eventual neurodegenerative consequences (Rodriguez-Gomez et al., 2020 [DOI](#)). In contrast, anti-inflammatory microglia (M2) are regulating the phagocytosis of pathogen and cell debris, inhibit inflammation and restore tissue homeostasis (Franco & Fernandez-Suarez, 2015 [DOI](#); Jurga et al., 2020 [DOI](#)). While our data does not support the presence of an acute inflammatory process in the hypothalamus, what we instead find is an increased alert state of the resident immune cells.

The most striking shift in gene expression in our dataset was detected in oligodendrocytes and astrocytes at 3 and 7 dpi, which caused the separation of the cell populations into distinct sub-populations at 7 dpi for both cell types. In oligodendrocytes, we detect a distinct expression of *Serum/glucocorticoid Regulated Kinase 1* (*Sgk1*) in the 7-dpi population. This protein belongs to the family of glucocorticoid receptors (GCR) and has been associated with several environmental perturbations, including its up-regulation in the CNS during stress or during fasting in obese individuals (Avey et al., 2018 [DOI](#); Hinds et al., 2017 [DOI](#); Miyata et al., 2015 [DOI](#); Nonogaki et al., 2006 [DOI](#)). Further, the activation of SGK1 in oligodendrocytes is believed to lead to morphological changes, potentially caused by abnormal swelling of cytoplasm in oligodendrocytes or arborization of their processes (Miyata et al., 2015 [DOI](#); Miyata et al., 2011 [DOI](#)). *Fkbp5*, encoding for FK506-binding protein 5, was another strongly induced transcript in oligodendrocytes at 7 dpi. It has previously been shown to be induced by glucocorticoids (Hahle et al., 2019 [DOI](#)) and involved in myelination in the prefrontal cortex (Choi et al., 2021 [DOI](#)). One of the main functions of oligodendrocytes is myelin formation around neuronal axons and the metabolic support of these to ensure neuronal signalling and information processing (Lee et al., 2012 [DOI](#); Moore et al., 2020 [DOI](#); Saab et al., 2016 [DOI](#)). In addition, oligodendrocytes are involved in orchestrating stress responses mediated by GCRs. This is relevant for our model, as glucocorticoids are released following immune activation. This release usually occurs in a bi-phasic pattern, with an initial peak at 2 – 3 dpi caused by pro-inflammatory cytokines and a second peak at 7 – 8 dpi driven by T-cell specific cytokines (Hermann et al., 1994 [DOI](#); Silverman et al., 2005 [DOI](#)). Since the disease specific subpopulation of oligodendrocytes, we detect at 7-dpi shows markers related to GC signalling, this sub-cluster is likely driven by T-cell specific cytokines. For the astrocytes, one of the most pronounced discriminatory factors for the 7-dpi specific population was the expression of the *Leucine-Rich Repeat-Containing Protein 8A* (*Lrrc8a*, also *Swell1*, annotated as ENSMUSG0000099041) transcript. *Lrrc8a* has putative regulatory transmembrane transporter functions and is one essential subunit of volume-regulated anion channels (VRAC). Hence, it plays an important role in cell volume changes and regulate synaptic transmission via glutamate release from astrocytes (Osei-Owusu et al., 2018 [DOI](#); Yang et al., 2019 [DOI](#)). The upregulation of *Lrrc8a* suggests changes in astrocytic morphology in the hypothalamus in our model, comparable to what has been reported in the hippocampus during peripheral IAV infections (Hosseini et al., 2018 [DOI](#)). However, the functional importance of this change in our study stays elusive and needs further investigation. In addition, we showed that the *zinc finger homeobox 3* (*Zfx3*) transcript is specifically up-regulated in astrocytes at 7 dpi. Its genomic locus has been associated with body mass index and it is further known to influence circadian rhythm and sleep (Balzani et al., 2016 [DOI](#);

Parsons et al., 2015; Turcot et al., 2018 [\[1\]](#)). The latter two are believed to be also regulated via astrocytic released glutamate (Brancaccio et al., 2017 [\[2\]](#)). Further, we identified a distinctive expressed gene in a part of the astrocyte population - *aldehyde dehydrogenase 1 A1* (*Aldh1a1*), which is contributing to retinol metabolism. The inhibition of *Aldh1a1* leads to reduced weight gain (Haenisch et al., 2018 [\[3\]](#)). *Aldh1a1* is known to mediate the synthesis of GABA in astrocytes (Kwak et al., 2020 [\[4\]](#)), which is then secreted by *Best1*-channels and GABA transporters (GAT, *Slc6a1* and *Slc6a11*) transporters (Mederos & Perea, 2019 [\[5\]](#)). These two transporters were found robustly expressed in the here analysed astrocyte population but not differentially expressed at any time point. The release of glutamate by astrocytes is regulated via the excitatory amino acid transporters (glutamate transporter 1; GLT-1, also known as *Slc1a2* and Glutamate Aspartate Transporter 1; GLAST, also known as *Slc1a3*) (Delgado, 2013 [\[6\]](#)), which we found highly expressed in the here analysed astrocytes (Supplementary Figure 11). Additionally, we identified an up-regulation of genes of the oxidoreductase system. Such modulations of the antioxidant gene expression system has been shown to regulate microglia activation states (Shih et al., 2006 [\[7\]](#)), so what we see might be a regulatory feature enabling astrocytes to fine tune the transitions between microglia homeostatic and inflammatory states. Taken together, the data suggest activation of two different transport processes in astrocytes. The expression of VRAC might facilitate glutamine release by astrocytes which can be taken up by neurons and converted to glutamate or GABA (McKenna & Ferreira, 2016 [\[8\]](#); Waagepetersen et al., 2003 [\[9\]](#)). This could be a supportive mechanism to maintain base level of neuronal signalling, which might be affected by the down-regulation of genes in the citric acid cycle in neurons and the accompanied loss of downstream products as neuropeptides. At the same time, the expression of transmembrane transporters might contribute to the swelling of astrocytes by the uptake of glutamate. Morphological changes in astrocytes are connected to appetite regulation by redirecting glutamate away from neurons and hence reducing glutamatergic signalling (Delgado, 2013 [\[6\]](#)). This might be a fundamental factor for sickness related weight loss and gain, categorizing the here identified 7-dpi astrocyte population as regulating neuroendocrine processes. Further studies are needed to understand and confirm the functional identity of the 7 dpi subclusters.

In summary, the data presented here shows how the cellular state of non-neuronal and neuronal cell populations in the hypothalamus change during peripheral IAV. Our data suggests the existence of tight co-regulation where one cell type affects the state of another. This connectivity on the one hand potentially ensures protection of the tissue by surveillant microglia but also counterbalances the microglia activity by astrocytic regulation of their antioxidant machinery. In addition, we found evidence that non-neuronal cell types like astrocytes and oligodendrocytes are potentially supporting neurons with their metabolic and signalling integrity, by enabling more nutrient transport as well as neuropeptide supply. Altogether, our study provides the first evidence of complex hypothalamic gene expression changes at the single cell level during a peripheral viral infection.

Methods

Animals and viruses

The experiments conducted in the study were approved by the Danish Animal Experimental Inspectorate (license number: 2020-15-0201-00585) in accordance with the Directive 2010/63/EU of the European Parliament and Council on the protection of animal used for scientific purposes. Female C57BL/6NTac mice were obtained from Taconic at 8-10 weeks old. Animals were kept in groups of 5, fed ad libitum and kept at a 12h day/night cycle. Virus stocks (H1N1 A/California/07/2009) were maintained at Prof. Jan P. Christensen Lab, University of Copenhagen. Virus stocks were propagated in Madin-Barby Canine Kidney cells (MDCK). Cell cultures were

infected at 0,01 MOI and harvested at 90-100 % CPE after 2-3 days. Supernatant was centrifuged to clear it of cell debris and supernatant was stored at -80°C for titration and infection of animals. Titration was done as previously described by Uddbäck and colleagues (Uddback et al., 2016 [DOI](#)).

Viral infections and tissue collection

10-week-old female C57BL/6N were inoculated intranasally with H1N1 A/California/07/2009 with a dose of 5×10^5 pfu in 30 µl per animal. 3 days, 7 days and 23 days post inoculation (dpi) the animals were sacrificed by cervical dislocation between 1:00-3:00 pm to minimize confounding effects from circadian rhythm or sleep/wake phases. The lungs, brains and olfactory bulbs were removed, frozen on dry ice and stored in at -80°C. The lungs and olfactory bulb were used to determine infection and the hypothalamus of the brain was used for snRNAseq.

Viral RNA Measurement by PCR

The olfactory bulbs and the right lung were homogenized using an ULTRA-Turrax T8.01 homogenizer in the RLT lysis buffer from the RNeasy mini and midi kit (Qiagen). The rest of the RNA extraction was performed according to the manufactured protocol from the RNeasy mini and midi kit to extract RNA from brain and lung tissue. The concentration and purity of RNA in the samples was measured by a Nanodrop spectrophotometer (NanoDrop 1000, Thermo Fisher Scientific, Delaware, USA). All the samples used in this experiment had a 1.7-2.1 260/280nm ratio. Next, the samples were diluted to 1 µg total RNA in each well and cDNA was synthesized by reverse transcription using the QuantiTect Reverse Transcription Kit.

Tissue preparation, nuclei isolation and hashing of nuclei

Mouse brains tissue were sectioned at 300 µm in a cryostat [Leica CM3050S] to identify the lateral hypothalamus (bregma -0.94 to -1.34) for punching. The sections were punched five times, and the tissue was transferred to a cooled Dounce-homogenizer containing Nuclei EZ lysis buffer to homogenize the tissue. The samples were filtered through a 40µm mini cell strainer followed by a 25%/29% iodixanol gradient to separate debris from the nuclei. To be able to distinguish samples later in post-sequencing analysis, the pellet was resuspended in FACS buffer containing sample-specific hash-tagged antibody recognizing a nucleus specific protein. A hashtag is an antibody with an oligonucleotide tail with PCR handle, a unique barcode and a poly A tail.

FACS Sorting and snRNA-sequencing

The sample were stained with DAPI (0.5µg/ml) and purified using the Sony SH800S cell sorter. Whole nuclei were gated using the DAPI fluorescence at 450 ± 25 nm, to avoid debris, and sorted directly into RT Reagent B (10X genomics) and further processed according to the company's guide and sequenced using a NovaSeq 6000 (Illumina). Commercially available anti-nuclear pore complex proteins antibodies from BioLegends, Inc. were used for hash-tagging. Per lane five different antibodies, to distinguish the different samples were used. Across the 5 sequencing lanes we used the same set of antibodies (TotalSeq™-A0451, Cat. No. 682205; TotalSeq™-A0452, Cat. No. 682207; TotalSeq™-A0453, Cat. No. 682209; TotalSeq™-A0454, Cat. No. 682211; TotalSeq™-A0455, Cat. No. 682213).

Single-nucleus RNA-sequencing raw data processing and quality control

Raw sequencing reads were processed using the 10x Genomics Cell Ranger version 3.0 pipeline and aligned to the *Mus musculus* (mm10) genome with default parameters. All libraries were converted to FASTQs using bcl2fastq. The FASTQ files were aligned to the mouse genome (assembly - GRCm38.p6) and subsequent read quantification was performed using the Salmon pipeline. Each

library underwent separate quality control steps as described in the Alevin pipeline (Srivastava, 2020). Hash-tagged antibody tag oligos (HTO) filtering removes nuclei with low tags or noisy cells. Furthermore, nuclei containing two or more cell barcodes (doublets) were removed.

Cells were tested for their mitochondrial RNA content and outliers with high mitochondrial RNA were removed using the quality control (QC) functions (perCellQCMetrics, PercentageFeatureSet, quickPerCellQC, isOutlier) from the R package *scuttle* (McCarthy et al., 2017). Counts were normalized using a log transformation of a genes counts divided by the total counts of a cell multiplied by 1e6. To remove variance attributed to batch and treatment effects for downstream visualization and clustering, samples were normalized with the ‘SCTransform’ method and integrated using the ‘FindIntegrationAnchors’ and ‘IntegrateData’ functions from Seurat (Version 4.1.0) (Hao et al., 2021). The integrated dataset was centred and scaled, PCA was carried out and for visualization we used UMAP dimensionality reduction.

Single-nucleus RNA-sequencing cell population identification

Cells were clustered into neuronal and non-neuronal cells using a Gaussian mixture model (GMM) based on the expression of pan-neuronal markers (*Syp*, *Galnt16*, *Rbfox3*, *Syn1*, *Dnm1*). Cells in clusters with high expression of these markers were classified as neurons and low expression as non-neuronal cells. The dataset was separated based on this classification and subsequently analysed independently. For identifying GABAergic and glutamatergic cells we used likewise a GMM based on known GABAergic and glutamate markers (*Slc17a6*, *Slc32a1*, *Gad1*, *Gad2*). Cells in clusters with high expression of GABAergic markers and low expression of the glutamatergic marker were defined as GABAergic cells and vice versa cells in cluster with high expression of the glutamatergic marker and low expression of the GABAergic markers were defined as glutamatergic. We divided the dataset further based on this classification. We identified a neuronal cluster expressing neither of the GABAergic nor glutamatergic markers. It also did not express any other neuron markers (histaminergic (HA), dopaminergic, ect.), and the cluster was therefore moved to the non-neuronal dataset.

Cluster identification was performed on the integrated dataset using the Seurat functions ‘FindNeighbors’ and ‘FindClusters’ using the Leiden algorithm. Non-neuronal cell populations and well known (i.e HCRT, HA, MCH, etc.) neuronal populations were identified and classified based of known marker genes. Additionally, we used published datasets from the hypothalamus (Campbell et al., 2017; Chen et al., 2017; Mickelsen et al., 2019; Mickelsen et al., 2020; Romanov et al., 2017; Steuernagel et al., 2022; Zeisel et al., 2018) to confirm cell type annotations by using the Seurat functions ‘FindTransferAnchors’ and ‘TransferData’. We accepted a prediction score ≥ 0.5 in at least 50 % of the cells of a cell cluster. Marker genes for the different cell populations were identified using the ‘FindMarkers’ function from the Seurat package and using the receiver operating characteristic (ROC) method.

Differential gene expression analysis and case-control based expression shifts (Cocoa)

To calculate gene expression fold changes in samples from infected mice compared to the control group, we used a pseudo-bulk approach. We calculated in total three different differential gene expression datasets; (I) Aggregating counts per gene across all cells without discriminating between cell types or clusters, (II) aggregating counts per gene for all GABAergic, glutamatergic, and non-neuronal cells and (III) aggregating counts per gene for each identified cluster. Differential gene expression analysis was performed used the *DESeq2* and *scran* R packages. Calculations were performed per time point. In order to account for biological variance we selected only cell types which were comprised of nuclei from at least 3 animals. Furthermore, we wanted to assure to only include cell types with a sufficient amount of nuclei (at least 9 nuclei/time point) and only included a cell type if it contained at least 3 nuclei from at least 3 individual animals. Further we selected only robustly expressed genes in a cell cluster for DGE analysis,

meaning a gene must be expressed in at least 95% of the cells of one of the tested time points versus the control to be included in the analysis. The counts were aggregated using the Seurat function ‘AggregateExpression’ and transformed to a DESeq2 data set with the function ‘DESeqDataSetFromMatrix’. We calculated normalization factors using the scran function ‘computeSumFactors’. The differential expression analysis was performed using a likelihood ratio test (LRT) and a Gamma-Poisson Generalized linear model (glmGamPoi) for fitting the dispersion to the mean intensities. P values were adjusted (for time points tested x number of cell populations x the number of genes) using the false discovery rate by Benjamini & Hochberg.

To dissect transcription responses for key neuronal cell populations (*Agrp/Npy+*, *Lepr+*, *Pomc+*, *Hcrt+*, *Pmch+* and *Hdc+*) we performed an additional differential gene expression analysis with adjusted parameters to identify DEGs also for small cell clusters (Supplementary Table 6). Here we accepted a cell cluster at a time point if it contained at least 2 cells from 2 individual animals.

We performed a case-control analysis based on the *Cocoa* package developed by Pethkhov et al. (Petukhov et al., 2022 [↗](#)), comparing the different infected time points (cases) against the control group. Therefore, we separated the Seurat object with all data into three different sub-dataset containing the 3 dpi and control, 7 dpi and control as well as 23 dpi and control samples. We performed cluster-based density and expression shifts and cluster-free analysis on default settings.

Gene set enrichment analysis

For gene ontology enrichment analysis (GOEA), we used the *topGO* package in R (Alexa & J., 2022). All significant differentially expressed genes (FDR ≤ 0.05) were defined as the gene universe. Only highly regulated genes (FDR ≤ 0.05, logFC > 1 or logFC < -1) were included in the GOEA and tested twice, first implementing a classical algorithm, and second using an algorithm eliminating (*elim* method) local dependencies between GO terms. For the t-test statistic we used in both cases a Fisher exact test. P-values returned by the *elimination* method are considered as not affected by multiple testing and hence can be interpreted as corrected (Alexa & J., 2022). Enrichment analysis for the Reactome database was performed with the ‘ReactomePA’ R package and its ‘enrichPathway’ function. We mapped the mice gene identifiers to human Entrez gene IDs for analysis. Genes were selected based on their expression in the same way as for GOEA. P values were adjusted for multiple testing using the Benjamini & Hochberg correction method.

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Data availability

Raw data filtered count matrices and metadata are available under accession number GEO: GSE226098. Code will be shared upon request.

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Reviewer #1 (Public Review):

This study uses single-cell genomics and gene pathway analysis to characterize the transcriptional effects of influenza H1N1 infection on hypothalamic cell types. The authors use droplet-based single-nuclei RNA-seq to profile genome-wide RNA expression in adult mouse hypothalamic cells at 3, 7, and 23 days after intranasal infection with the H1N1

influenza virus. Through state-of-the-art and rigorous computational methods, the authors find that many hypothalamic cell types, glia, and especially neurons, are transcriptionally altered by respiratory infection with a non-neurotropic influenza virus and that these alterations can persist for weeks and potentially affect cell type interactions that disrupt function. For instance, microglia shift towards a pro-inflammatory molecular phenotype at 3 days post-infection, while astrocytes and oligodendrocytes significantly alter their expression of oxidoreductase activity genes and transport genes, respectively, at 7 days post-infection. In addition, POMC neurons of the arcuate hypothalamus, which suppress appetite and increase metabolism, appear to be unusually sensitive to H1N1 infection, upregulating more genes than other hypothalamic neurons. The authors' thorough discussion of the findings raises interesting questions and hypotheses about the functional implications of the molecular changes they observed, including the physiological changes that can persist long after acute viral infection. Given the role of the hypothalamus in homeostasis, this work sheds light on potential mechanisms by which the H1N1 virus can disrupt cell function and organismal homeostasis beyond the cells that it directly infects.

Reviewer #2 (Public Review):

The new work from Lemcke et al. suggests that the infection with Influenza A virus causes such flu symptoms as sleepiness and loss of appetite through the direct action on the responsible brain region, the hypothalamus. To test this idea, the authors performed single-nucleus RNA sequencing of the mouse hypothalamus in controlled experimental conditions (0, 3, 7, and 23 days after intranasal infection) and analyzed changes in the gene expression in the specific cell populations. The key results are promising and spurring future research. After revision, the analysis was considerably improved. Alternative approaches were used for testing. Specifically, during the revision: 1) The annotation of cell types was considerably improved; 2) The authors performed an additional analysis comparing case-control studies (Cocoa), where they could partly confirm their earlier findings.

Author Response:

The following is the authors' response to the current reviews.

Reviewer #1 (Recommendations For The Authors):

The revision and rebuttal have addressed all concerns raised in the initial review. Upon review of the revised figures, however, it is unclear why Figure 8C shows many significant DEGs in POMC neurons (which according to Figure 8b is the "GABA_24" cluster), whereas Figure 6A shows few to no DEGs in the GABA_24 cluster. Same for Pmch neurons/Glut_25, which seem to be missing from Figure 6A.

Answer: In order to capture changes in these smaller cell population we performed an additional DEG analysis with modified and less strict parameters (compared to the first main analysis). We mention the different parameters in the methods part of the revised manuscript (Differential gene expression analysis and case-control based expression shifts (Cocoa)).

The following is the authors' response to the original reviews.

Reviewer #1 (Recommendations For The Authors):

Major issues

1. A key conclusion of this study is that neurons show longer lasting infection-related changes in gene expression than do non-neuronal cells, suggesting that neurons are more persistently affected, which could potentially underlie persistent effects of infection on behavior or physiology. However, the authors also report that over twice as many transcripts were captured in neurons than in non-neuronal cells, and that neurons and non-neurons were not equal in number. The number of transcripts and cells per cell type can affect the likelihood of detecting a differentially expressed gene when comparing cell types. Thus, the difference in infection related DEGs between non-neuronal cells and neurons may be due in part to differences in the numbers of transcripts and cells in each group. How would the number of infection related DEG's compare if the same number of transcripts were detected in neurons as in non-neuronal cells? In addition, is there any relationship between the number of infection related DEGs detected and the number of cells in the respective groups?

We performed an additional analysis, down sampling the transcripts per cells to similar numbers (~1600 transcripts/cell), showing a similar pattern as shown in the original calculation of DEGs. High downregulation of genes in GABAergic, Glutamatergic and Nonneuronal cells at 3 and 7 dpi, but long-lasting dis-regulation at 23 dpi only in the neuronal subtypes. The analysis results can be found in Supplementary figure12 and on page 11 in the results section.

1. The rationale for focusing on the LH and DMH is unclear. While these regions do play important roles in control of body weight and wakefulness, the authors do not report whether the cell types relevant to these functions are among those affected by infection. For instance, the authors mention HCRT and MCH neurons in the introduction but do not comment on whether these neurons show any significant changes after H1N1 infection in their analysis. Also, what about the POMC neurons or the Lepr+ DMH neurons? Knowing whether and how these body weight associated cell types are affected could help to connect the phenotypic (e.g., body weight) and molecular changes observed.

We have added an additional analysis of some well know hypothalamic subtypes. What is interesting is that the different neuronal subtypes respond to the infection differently. While most neurons show the strongest response at 3dpi, POMC+ neurons show consistent changes across all three time points. This could point to different neuronal subtypes paying different roles in the sickness response to the influenza infection. The new data has been added to Figure 8 together with new text in the result section and discussion (Page 17 & 20).

1. For discriminating neurons and non-neuronal cells based on their expression of neuronal marker genes, was this performed at the single-cell level or the cluster level? Similarly, was the discrimination of GABAergic and glutamatergic neurons done at the cell or cluster level?

The discrimination of the cell types was done on single cell level. This information has been added to the revised manuscript on page 25.

1. The authors mention that body weight did not change in some of the mice. Was there any difference in infection related DEGs between the mice that lost weight and the mice that didn't? Was there any correlation between the molecular and phenotypic (i.e., body weight) changes observed?

We agree that this could have been an interesting point to investigate, however, we can only say with certainty for 2 animals in the recovery group (23.7 and 23.8) that they didn't lose weight (Supplement figure 2). In Figure 4A we show that overall the different time points

group well together, with exception for animal 23.7 which seems to have a better overlap with 7 dpi, indicating that we possibly captured here a delayed disease response. However, to make any indepth analysis, we have to few animals without weight-loss.

5)The authors noted that the hypothalamic neurons continue to show infection-related changes in gene expression at 23dpi though body weight has returned to normal. In this H1N1 model, are there any persistent behavioral deficits at 23dpi that could be explained by the persistent changes in gene expression in DMH and LH neurons?

We did not test for long-lasting behavioural changes in these animals. Another study by Hosseini et al. (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6596076/>) focus on cognitive long term effects of viral infections. Even though they did not include the here used H1N1 model, they included the PR8 strain, but didn't report any long lasting behavioural or cognitive changes. So far only cognitive deficits during the acute phase of the infection caused by the PR8 H1N1 model have been shown. This would be a very interesting follow up study to perform, but this, we believe, is out of scope for the current manuscript.

1. In Figure 1F, the 3dpi sample appears to differ from the other samples in terms of its neuron/non-neuron composition. The authors point this out but offer no discussion or further analysis. Was this difference driven by one or more cell types? Is this difference likely to be technical (e.g., less white matter in sample = fewer oligodendrocytes), or could this be related to the infection (e.g., glial death or neurogenesis at 3dpi)?

We have added the location of the punching within the hypothalamus for the different groups to the supplements (Supplementary Figure 3). The differences in neuron/non-neuron composition could originate from differences in the punching location, but we do not have data to support this conclusion. The difference could also stem from biological alterations during the infection.

1. Since influenza viruses replicate in the cell nuclei, did the authors capture any H1N1 RNA in their single-nuclei RNA-seq samples?

We mapped the single nuclei data against the viral genes, but could not detect any of the viral genes in the data set. We are still optimizing detecting of low amounts of viral genes in snRNA-seq data and have not included this information in the manuscript. We believe, that the virus did not manage to migrate in the hypothalamus and infiltrate the cells in the here captured area.

Minor Issues

1. Page 1. The abstract ends with the sentence: This is complemented by increased activity of microglia monitoring their surroundings. Presumably, the authors are basing this statement on the functions of genes altered in microglia by infection. However, saying that microglia behavior has changed is a bit of a stretch here, since the results suggest a change in the molecular phenotype of microglia but do not demonstrate a change in their behavior.

We agree that the phrasing of the end of the abstract was not accurate and didn't reflect the outcome of the analysis. We adjusted the sentence to: "The change of microglia gene activity suggest that this is complemented by a shift in microglia activity to provide increased surveillance of their surroundings." Which should provide a better idea that the findings we present are a suggestion based on the transcriptomic changes in the cell population. (Page 1)

1. Page 8. The authors refer to *Th+*, *Ddc+* neurons as dopaminergic. However, adrenergic/noradrenergic neurons also express these genes. How do the authors know the neurons are not adrenergic/noradrenergic?

There are to our knowledge no nor-adrenaline/adrenaline producing neurons in the hypothalamus. In contrast dopaminergic neurons have indeed been identified in this area.

1. In the Methods section, *Slc17a6* and *Slc32a1* are not "pan-neuronal markers" since they are only expressed by subsets of neurons.

We removed the glutamatergic and GABAergic marker genes (*Slc17a6* and *Slc32a1*) from the list of neuronal markers. They are stated further down in the method section as glutamatergic and GABAergic markers. Find the changes on Page 24/25)

1. Was the hashtagging antibody custom or commercial? If commercial, what was the source, catalog #, lot #? If custom, the authors should describe how it was made and validated.

We used commercial antibodies for hash-tagging. We added the missing information to the manuscript and can be found on Page 24 of the revised manuscript.

1. In the data processing section of the Methods, *SCTransform* is mentioned twice. Was normalization with *SCTransform* applied twice?

The data was only normalized once using the *SCTransform* method. We adjusted the part of the method section to make it more clear (Page 24).

1. In the section on gene set enrichment analysis, the first sentence includes this text: "(is a reference needed?)." The answer is yes - Alexa A, Rahnenfuhrer J (2022). *topGO: Enrichment Analysis for Gene Ontology. R package version 2.50.0*.

The missing reference was added (Page 26).

1. Page 4: "leaved" should be corrected to "left"

The wrong wording was corrected.

1. Figure 2D - gene is labeled as *Slc31a1* on the figure and *Slc32a1* in the figure legend

We provided a new Figure plate with the right marker genes.

1. Official gene IDs should be italicized

We checked the gene IDs again, and italicized wrongly formatted gene IDs.

1. It is not clear whether the authors are planning to share their code. However, their code would be needed to reproduce their results, since the methods section provides a summary of what was done but lacks key details (e.g., parameters and software packages used during data processing and analysis)

Code will be shared on request. We added this also to the revised manuscript (Page

1.

Reviewer #2 (Public Review):

The new work from Lemcke et al suggests that the infection with Influenza A virus causes such flu symptoms as sleepiness and loss of appetite through the direct action on the responsible brain region, the hypothalamus. To test this idea, the authors performed single-nucleus RNA sequencing of the mouse hypothalamus in controlled experimental conditions (0, 3, 7, and 23 days after intranasal infection) and analyzed changes in the gene expression in the specific cell populations. The key results are promising.

However, the analysis (cell type annotation, integration, group comparison) is not optimal and incomplete and, therefore should be significantly improved.

More specifically:

- 1. The current annotation of cell types (especially neuronal but also applicable to the group of heterogeneous "Unassigned cells") did not make a good link to existing cell heterogeneity in the hypothalamus identified with scRNA seq in about 20 recently published works. All information about different peptidergic groups can not be extracted from the current version (except for a few). There are also some mistakes or wrong interpretations (eg, authors assigned hypothalamic dopamine cells to the glutamatergic group, which is not true). This state is feasible to improve (and should be improved) with already existing data.*

We repeated the cell label transfer with the newly published HypoMap and added additional information to the supplements about the cell type assignments. Additionally, we agree that the dopaminergic neurons do not belong to the group of glutamatergic neurons, however assigned them into this group based on the clustering. We changed the phrasing in the results, to make a better differentiation between the two groups (Page 8).

- 1. I am confused with the results shown in the label transfer (suppl fig 3 and 4; note, they do not have the references in the text) applied to some published datasets (authors used the Seurat functions 'FindTransferAnchors' and 'TransferData'). The final results don't make sense: while the dataset for the arcuate nucleus (Campbel et al) well covered the GABAergic neurons it is not the case for the whole hypothalamus datasets (Chen et al; Zeisel et al). Similarly, for glutamatergic neurons. Additionally, I could not see that the label transfer works well for PMCH cells which should be present in the dataset for the lateral hypothalamus (Mickelsen et al,2019).*

We performed the additional label transfer of the hypothalamus data. Here we accepted a prediction score of 0.5 and transferred a cell type label to our annotated cluster IDs, if at least 10% of cells within a cluster were annotated with the 0.5 prediction score. We found that well defined neuron population types like Hcrt+, Pmch+ and Hdc+ neurons as well as Pomc+ neurons were tagged with a high predictions scores (≥ 0.9 , Supplement Figures 6 and 7) and non-neuronal cell types (Supplement Figure 8) were well annotated. Additionally we identified an Agrp+ neuron population with the Gaba_1 neurons. This information has been added to the revised manuscript (Pages 6, 8).

- 1. There are newly developed approaches to check the shifts in the cell compositions and specific differential gene expression in the cell groups (e.g. Cacoa from Kharchenko lab, scCoda from Büttner et al; etc). Therefore, I did not fully understand why here the authors used the pseudo-bulk approaches for the data analysis (having such a valuable dataset with multiple hashed samples for each timepoint). Therefore it would be great to use at least one of those approaches, which were developed specifically for the scRNAseq data analysis. Or, if there are some reasons - the authors should argue why their approach is optimal*

We performed an additional analysis comparing case-control studies (Cocoa). We performed both modalities, cluster-based and cluster-free expression shifts and cell type compositions. We could partly confirm our findings using the pseudo-bulk approach. The cluster-specific density shift (Supplement Figure 15) identified only shifts in non-neuronal cell types between the Control group and 3 dpi. We believe, these composition shifts are caused by the lower number of non-neuronal cells in the 3 dpi time point. Cluster-specific expression shifts show similar results as in the pseudo-bulk approach, with significant expression shift identified at 3 and 7 dpi in neuronal and non-neuronal cell clusters (Supplement Figure 16). However, no significant expression shifts were identified in the recovery group at 23 dpi. Using the cluster-free expression shift approach, however we were able to identify a similar picture as described with the pseudo-bulk approach. In the recovery group at 23 dpi, we found mainly changed gene programs in neuronal cells, and no transcriptional changes in the non-neuronal cells (Supplement Figure 17-20). This new analysis has been added to the revised manuscript (Pages 4-6, 26) including supplementary figure and tables as stated.

1. When the authors describe the DGE changes upon experimental conditions (Figures 5 and 6), my first comment is again relevant: it is difficult to use the current annotation and cell type description as the reference for testing virus effects and shifts in the DGE in distinct neuronal subtypes.

The cell type annotations have been checked and additional label transfer has been performed. All figures in the manuscript have been updated.

I have to note that the experimental design is well done and logical. Therefore I believe that to strengthen the conclusions, the already obtained datasets can be used for improved analysis.

Reviewer #2 (Recommendations For The Authors):

I have some minor concerns:

1. For the quality check it would be good to see how different hashed samples for each timepoint cover the UMAP embeddings.

We added the UMAP embeddings to the supplement. (Supplement Figure 4)

1. In Fig 1e colors are not optimal - it is impossible to assess it.

We separated the UMAPs for the different time points to make it easier to assess. See updated Figure 1E.

1. In the methods authors started "Single-nucleus RNA-sequencing cell population identification" from the description of using a Gaussian mixture model (GMM). However, I could not clearly understand how this model was used and which kind of result it provided.

We used an GMM model with known markers for neurons and in a second step for glutamatergic and GABAergic cells to sub-cluster the cells and then selected based on high and low expression of the marker genes in the cluster into their respective classes. This information has been added to the method section (Page 24/25).

1. Could the authors better clarify why "they calculated normalization factors using the `scran` function `'computeSumFactors'`" when working with pseudobulk analysis?

This size factor normalization was recommended for single cell data by the authors of the DESeq2 packages.

<http://bioconductor.org/packages/devel/bioc/vignettes/DESeq2/inst/doc/DESeq2.html>

1. I didn't find logic in "a cell cluster was only included if it contained more than 2 nuclei in at least 3 individual animals" (page 24). Maybe I misinterpreted it.

The rationale for the selection methods was based on the findings that not all animals in the recovery group had the same effects in weight loss. The acute time points didn't show enough weight loss to decide if all animals in these groups lost the same amount and were equally sick. Hence, in order to have biological robustness we decided to only analyse clusters where cells from at least 3 animals at a specific time point contributed to a cell type. In order to have enough cells per cell type for the calculation of DEGs, we decided to only include a cell type at a specific time point if it contained at least 3 cells from one individual. This selection method limits the analysis to cell types with at least 9 cells per time point.