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Heparan sulfate dependent phase separation of CCL5 and its chemotactic activity

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Abstract

Secreted chemokines form concentration gradients in target tissues to control migratory directions and patterns of immune cells in response to inflammatory stimulation; however, how the gradients are formed is much debated. Heparan sulfate (HS) binds to chemokines and modulates their activities. In this study, we investigated the roles of HS in the gradient formation and chemoattractant activity of CCL5 that is known to bind to HS. CCL5 and heparin underwent liquid–liquid phase separation (LLPS) and formed gradient, which was confirmed using CCL5 immobilized on heparin-beads. The biological implication of HS in CCL5 gradient formation was established in CHO-K1 (wild type) and CHO-677 (lacking HS) cells by Transwell assay. The effect of HS on CCL5 chemoattractant activity was further proved by Transwell assay of human peripheral blood cells. Finally, peritoneal injection of the chemokines into mice showed reduced recruitment of inflammatory cells either by mutant CCL5 (lacking heparin binding sequence) or by addition of heparin to wild type CCL5. Our experimental data propose that co-phase separation of CCL5 with HS establishes a specific chemokine concentration gradient to trigger directional cell migration. The results warrant further investigation on other heparin binding chemokines and allows for a more elaborate insight into disease process and new treatment strategies.

eLife assessment

How the triplicate interaction between chemokines with both GAGs and G protein-coupled receptors (GPCR) works and how gradients are created and potentially maintained in vivo are poorly understood. The authors provide **solid** evidence to show phase separation can drive chemotactic gradient formation. The paper is a **useful** advance in the field of chemokine biology.

Introduction

Gradients of signaling molecules are ubiquitous in embryonic development and cellular activities, modulating cell migration, proliferation, and survival(Griffith, Sokol et al., 2014, Yamamoto, Kambayashi et al., 2022, Yu, Burkhardt et al., 2009). Chemokines, belonging to a ~45-member family of small (8-12 kDa) proteins, are signaling molecules that can induce distinct cellular chemotaxis in response to inflammatory stimuli in a concentration-dependent gradient(Weber, Hauschild et al., 2013). Chemokines play critical roles in regulating cell migration in a wide range of biological activities, e.g. developmental, homeostatic and inflammatory/pathological processes(Douglas, Dyer et al., 2015). It is established that chemokines are secreted from source cells and immobilized on glycosaminoglycans (GAGs) including heparan sulfate (HS) in the extracellular matrix(Proudfoot, Handel et al., 2003). The interaction of GAG-chemokine forms an immobilized gradient to provide direction to cell movement(Proudfoot, Johnson et al., 2017). Understanding how these chemokine gradients are formed and maintained is fundamental to identifying how they direct cell migration and proliferation. Straightforward molecular diffusion is often proposed as a possible mechanism(Kicheva, Pantazis et al., 2007, Schier & Needleman, 2009, Yu et al., 2009). However, it is assumed that more complicated mechanisms are involved, which remains to be elucidated.

CCL5 (also known as RANTES, for 'regulated on activation normal T cell expressed and secreted') (Roscic-Mrkic, Fischer et al., 2003) is an inflammatory chemokine that recruits a wide variety of leukocytes, including monocytes, granulocytes, and T cells, as well as mast cells and dendritic cells, through chemokine gradients(Roy, Mondal et al., 2015, Weber et al., 2013). CCL5 is composed 68 amino acids that reversibly self-assembles into high-MW oligomers, up to >600 kDa. This highly basic protein binds heparin with high affinity(Mulloy, 2005) and the oligomerization of CCL5 for the formation of gradient is modulated by GAGs on cell surface (Proudfoot et al., 2003).

Liquid–liquid phase separation (LLPS) driven by weak interactions between multivalent biomolecules was shown to be an important mechanism by which mesoscale structures of the condensates can form within the cell(Xue, Gong et al., 2019) and on the cell surface(Xue, Zhou et al., 2022). Within phase-separated condensates, biomolecules are often mobile and interchanged between the dense and light phases. Based on reported information and our previous study, we have studied the implications of HS on CCL5 gradient formation and its chemotactic activity. Both in vitro and in vivo experiments demonstrate that co-phase separation of CCL5 with HS establishes specific chemokine concentration gradients for chemotactic activity.

Results

Co-phase separation of CCL5 and heparin

To illustrate the phase separation property of CCL5 in liquid, recombinant protein expressed in *E. coli* (**Figure 1—figure supplement 1A**)(Proudfoot, Power et al., 1996) was labeled with organic dye Cy3 (CCL5-Cy3) and mixed with heparin at different ratios. After 30 min formation of droplets was examined by fluorescence under confocal microscopy. At the ratio of 20:1 CCL5-Cy3 to heparin, few tiny droplets were detected, indicating phase separation. With increased ratio of CCL5-Cy3: heparin to 1:20 (20 μM CCL5-Cy3:1 μM heparin), large and round droplets were formed (**Figure 1A and B**). Further increases in heparin concentration (ratio 1:50) prevented droplet formation as a "reentrant" behavior. Since heparin is a linear repetitive structure with rich negative charges, the LLPS seems following a similar mechanism as RNA in the phase separation of many RNA binding proteins(Ghosh, Mazarakos et al., 2019). Longer incubation revealed that

the droplets were dynamic and some became fused to form larger droplets (**Figure 1C**) that displayed a quick recovery speed in the fluorescence recovery after photobleaching (FRAP) (**Figure 1D**). The heparin-induced phase separation of CCL5 was attenuated at high salt concentrations as illustrated by incubation at different KCl concentrations in KMEI buffer (**Figure 1—figure supplement 2A and B**). In comparison, a mutant (A22K-CCL5) with higher positive charge density showed stronger interaction with heparin(Brandner, Rek et al., 2009) and formed aggregates instead of phase separation with heparin in KMEI buffer (**Figure 1—figure supplement 2C and D**). These results indicated that heparin-induced LLPS of CCL5 was based on weak electrostatic interactions between the negatively charged polysaccharide and the basic protein of CCL5, while strong ionic interaction has abolished this effect.

An earlier study showed complex formation of CCL5 with a heparin disaccharide, which proposed the BBXB motif of CCL5 playing essential role in the interaction between the chemokine and the disaccharide (Shaw, Johnson et al., 2004). In light with this finding, we mutated ^{44}RKNR^{47}, a conserved positively charged BBXB motif of CCL5, to hydrophobic alanine motif ^{44}AANA^{47} (Proudfoot, Fritchley et al., 2001), which is expected to reduce the interaction with heparin. As anticipated, the phase separation of the mutant ^{44}AANA^{47}-CCL5 in solution containing heparin formed much smaller and fewer droplets in comparison to WT CCL5 (**Figure 1E, F**). The electrostatic interaction feature was further demonstrated by molecular docking analysis using a tetrasaccharide structure of heparin (**Figure 1—figure supplement 3A**). The results showed the docking of heparin tetrasaccharide into human CCL5, forming crucial electrostatic contacts at the CCL5 dimer interface, where it packed tightly against Arg 44, Lys45, Arg47 **(Figure 1—figure supplement 3A)**. In comparison, binding free energy of heparin to the ^{44}AANA^{47}-CCL5 mutant was significantly higher (**Figure 1—figure supplement 3B**).

Heparin dependent phase separation is one essential step for CCL5 gradient formation

To elucidate the intrinsic connection between the heparin-dependent phase separation of CCL5 and its chemotactic activity, we developed an in vitro diffusion assay in which purified His-CCL5-EGFP (400 ng/μL) (**Figure 1—figure supplement 1B**) was immobilized on heparin-beads or on Ni-NTA-beads. In 96-well plates, the beads were embedded in the center of Matrigel(Helen P. Makarenkova, 2009) and incubated for 12 hr. Diffusion of the His-CCL5-EGFP was monitored by measuring fluorescence intensity along a line interval passing through the bead localized center. As shown in **Figure 2A**, His-CCL5-EGFP diffused a considerable distance away from the heparin-beads, forming a long and shallow gradient, whereas His-CCL5-EGFP did not diffuse from the Ni-NTA beads (**Figure 2B**). In comparison, ^{44}AANA^{47}-CCL5 binding to the heparin-beads was significantly weaker and did not diffuse (**Figure 2A**, right panel). To exclude the potential effect of EGFP, we repeated the experiment using Cy3-tagged CCL5. In a similar manner, the heparin-beads bound CCL5-Cy3 diffused in Matrigel but not the Ni-NTA beads bound CCL5-Cy3 as revealed by 3D imaging (**Figure 2—figure supplement 1 and Figure 2—figure supplement 2**). These results suggest that heparin beads tethered CCL5 via phase separation, which enables rapid exchange with the external environment leading to diffusion and gradient formation; while the Ni-NTA beads bound CCL5 lacks phase separation, therefore no diffusion.

Next, to test whether the formed chemokine gradients of CCL5-heparin beads contribute to the chemotactic activity, we established an in vitro chemotaxis assay using Transwell (Proudfoot et al., 2003) in which CCL5 either in heparin-beads or in Ni-beads were placed in the lower chamber (**Figure 2C**) and THP-1 cells were placed in the upper chamber. Again, CCL5-heparin-beads showed robustly higher chemotactic activity (**Figure 2D**), while CCL5-Ni-NTA beads essentially did not induce chemotaxis. In comparison, ^{44}AANA^{47}-CCL5-heparin-beads showed relatively weaker chemotactic activity than WT-CCL5-heparin-beads (**Figure 2D**). The dramatic difference

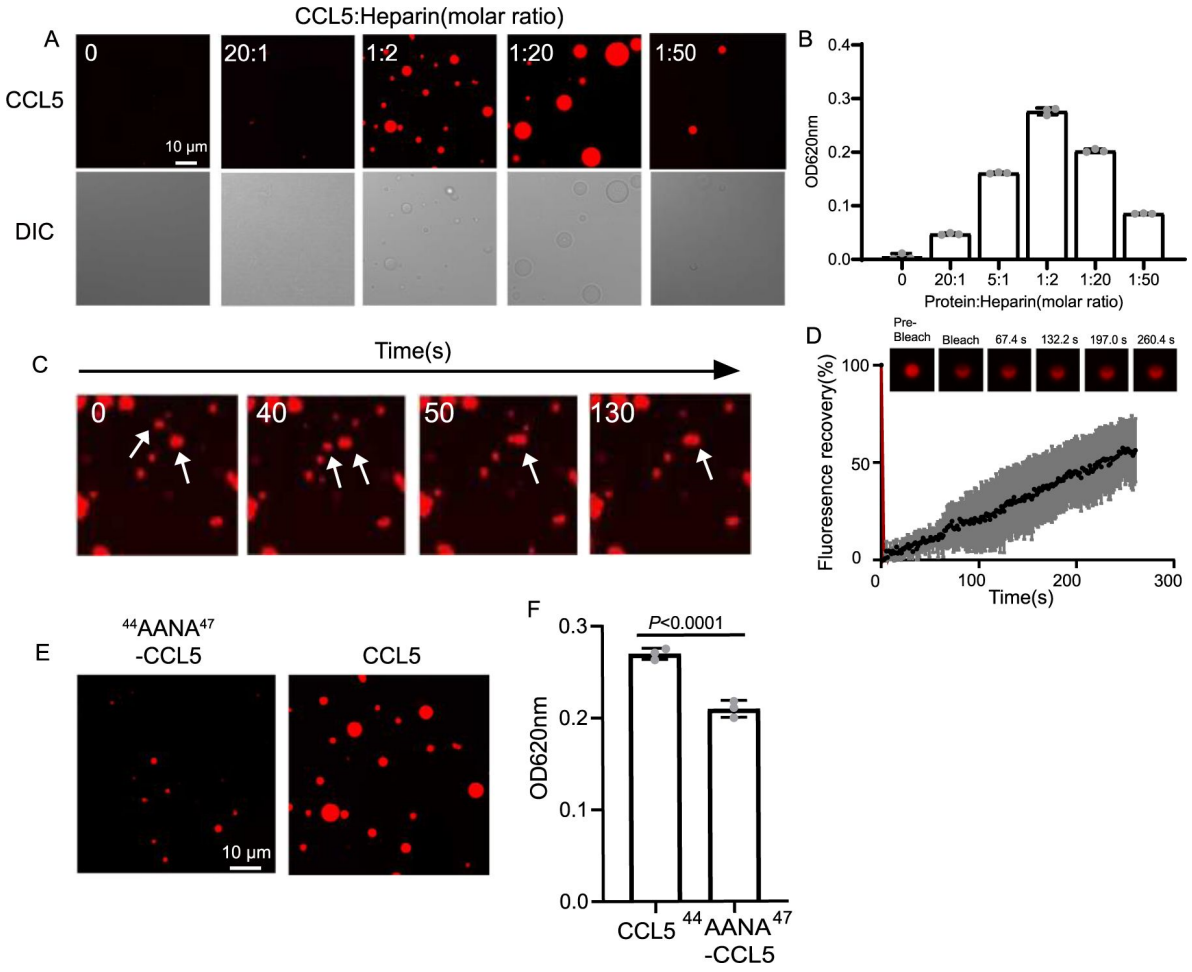

Figure 1

CCL5 phase separates with heparin in solution.

(A) Confocal images of assembling status of 20 μM CCL5-Cy3 mixed with heparin at different ratios and 5% PEG. Scale bar=10 μm. (B) Turbidity changes with increase of heparin concentration were measured at 620 nm (**Figure 1**-Source Data 1). (C) Fusion of phase-separated droplets formed at the ratio of CCL5-Cy3: heparin=1:2. The white arrows indicate the dynamic fusion of two adjacent droplets. (D) Representative FRAP results of droplets formed by CCL5-Cy3: heparin=1:2 depicted in **Figure 1A**, showing the intensity of fluorescence pre-and after photobleaching (**Figure 1**-Source Data 2). The images of representative droplets in different recovery stages are shown. (E) Confocal images of assembling status of ^{44}AANA^{47}-CCL5 or CCL5 in the presence of heparin. Scale bar=10 μm. (F) Comparison of CCL5 and ^{44}AANA^{47}-CCL5 turbidity in the presence of heparin (**Figure 1**-Source Data 3). Data are mean ± s.d. n=3 (for B, F). Normal distribution was assessed by the Shapiro-Wilk (SW) normality test. *P* values were determined by unpaired two-tailed t-tests.

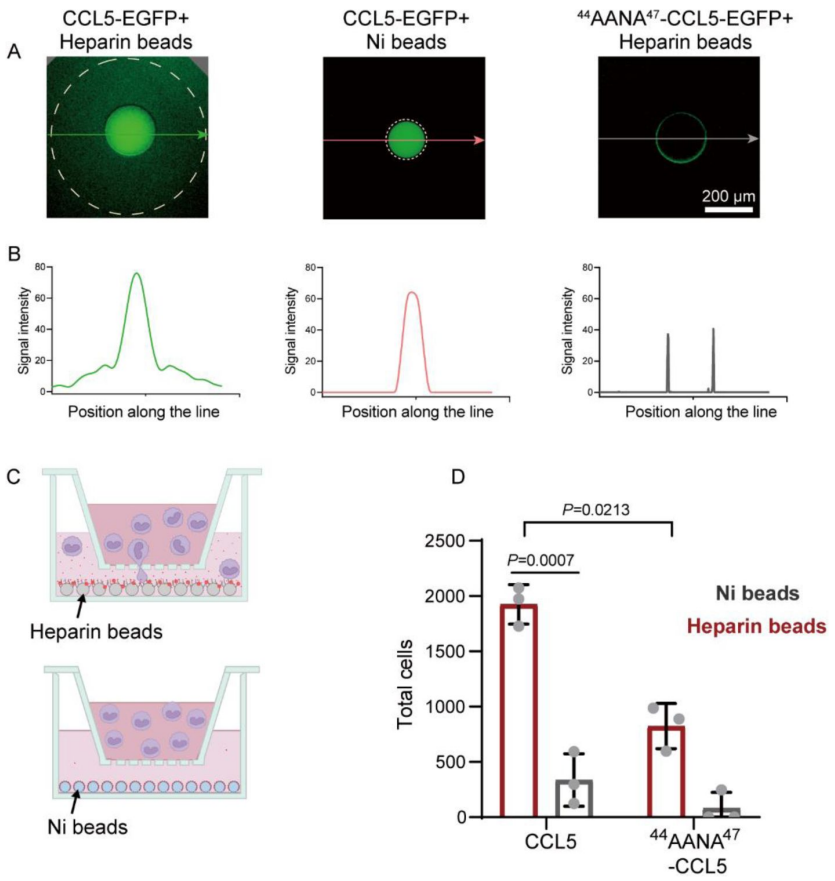

Figure 2

Co-phase separation of CCL5 and heparin establishes chemokine gradient

(A) CCL5-EGFP or ^{44}AANA^{47}-CCL5-EGFP were bound to heparin beads or Ni-NTA beads, respectively, and were placed in Matrigel in 96-well plate. After incubation for 12 hr images were taken to quantify the fluorescence intensity. (B) Quantification of the fluorescence signals along the lines with arrows indicated in **Figure 2A** (**Figure 2**-Source Data 1). (C) Illustration of *in vitro* chemotaxis assay. (D) Heparin beads or Ni-NTA beads bound with CCL5 or ^{44}AANA^{47}-CCL5 were placed in the lower chamber. THP-1 cells (3×10^5 cells) were added to upper chamber. After 4 hr, THP-1 in the lower chamber was collected and counted (**Figure 2**-Source Data 2). Data are mean ±s.d. n=3. Normal distribution was assessed by the Shapiro-Wilk (SW) normality test. *P* values were determined by unpaired two-tailed t-tests.

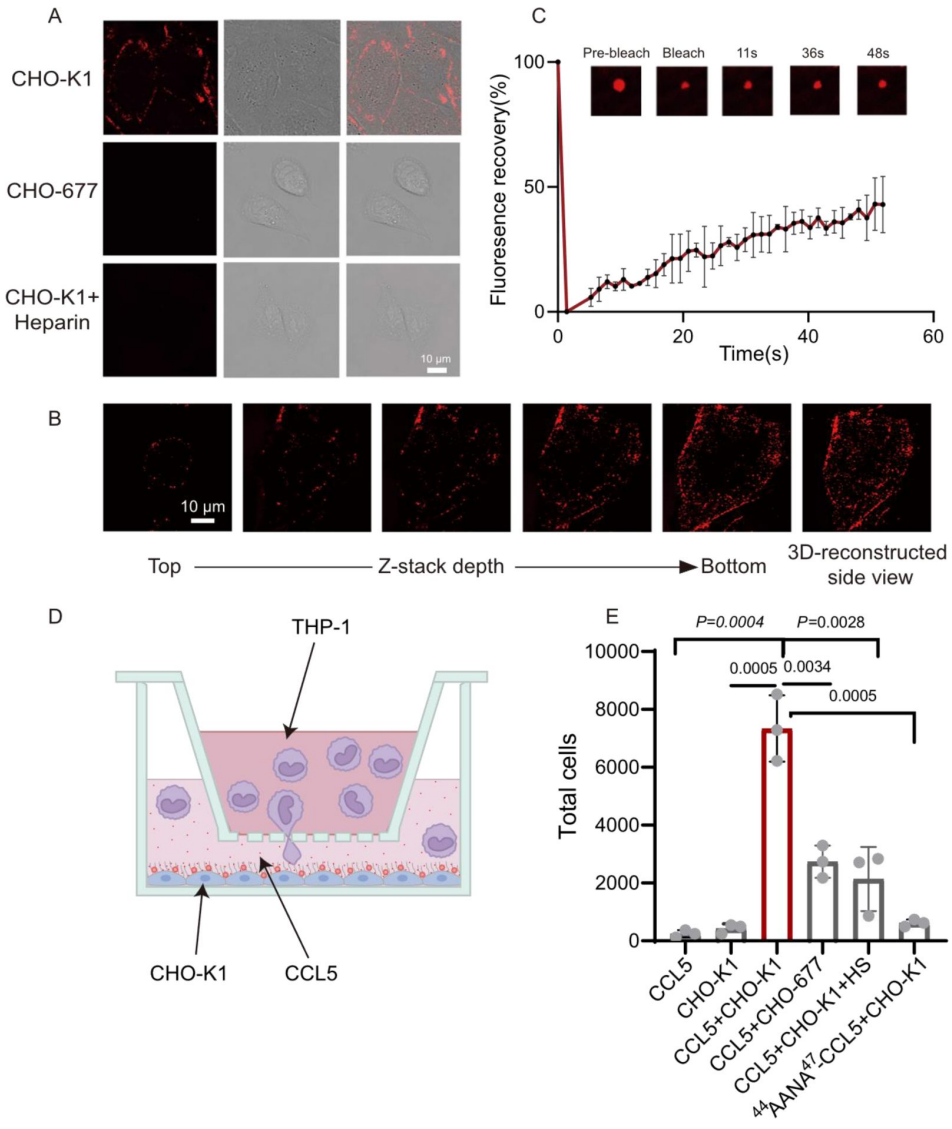

Figure. 3

CCL5 phase separates with heparan sulfate on the cell surface

(A) Microscopy images of CCL5-Cy3 on the surface CHO-K1 and CHO-677 as well as CHO-K1 treated with 1 mg/mL heparin. From left to right, the images are fluorescent-field, bright-field and overlay of two illuminations. Scale bar=10 μm. (B) Z-stack scanning of CCL5-Cy3 phase separation on CHO-K1 cell surface. The cell was imaged by confocal microscope with the Z-stack method. Scale bar=10 μm. (C) FRAP of the condensates formed by CCL5-Cy3, showing the intensity of fluorescence pre-and after photobleaching (**Figure 3** -Source Data 1). The size of the representative droplets in different recovery stages are shown above of the graph. (D) Graphical illustration of the cell-based chemotaxis assay. CHO cells (1×10^5 cells/well) were plated onto the lower chamber for 24 h (fully attached) and CCL5 or ^{44}AANA^{47}-CCL5 or heparin were added as indicated. THP-1 cells (3×10^5) were placed on upper chambers. After 4 hr, a small volume of medium in the lower chamber was aspirated to count THP-1 transmigrated through the membrane. (E) Quantification of THP-1collected from the lower chamber (**Figure 3** -Source Data 2). Data are mean ±s.d. n=3. Normal distribution was assessed by the Shapiro-Wilk (SW) normality test. *P* values were determined by unpaired two-tailed t-tests.

in the chemotactic activity of CCL5 between immobilization on heparin-beads or Ni-NTA beads indicate that the chemotactic function is achieved by establishment of a functional gradient rather than just immobilization.

Heparan sulfate dependent CCL5 phase separation and its chemotactic activity

Having seen the functional roles of heparin in CCL5 gradient formation in solution and Matrigel, we wanted to find out whether heparan sulfate (HS) on the cell surface also can phase separate with CCL5 using a pair of well-established cell lines, CHO-K1 (wild type) and CHO-677 (mutant lacking HS). Co-incubation of CCL5-Cy3 with the cells resulted in strong signals on the cell surface of CHO-K1 revealed by confocal microscopy, indicating formation of puncta surrounding the cells (**Figure 3A**). However, CHO-677 completely lacked the signals. The finding was further verified by Z-stack scanning (**Figure 3B**). Fluorescence recovery of about 40% condensates after photobleaching (FRAP) indicates there is a liquidity of the condensates on CHO-K1(**Figure 3C**). These results indicated that CCL5 formed phase separation with HS on the CHO K1 cell surface, which was not possible in CHO-677 cells lacking HS.

In order to know whether the HS-dependent CCL5 phase separation is essential for the chemotactic activity of CCL5, we established a cell-based chemotaxis assay (**Figure 3D**). CHO-K1 cells along with CCL5 was placed in the lower chamber for 1 hr and THP-1 cells was seeded in the upper chamber, separated by a porous membrane. After incubation for 4 hr, THP-1 cells transmigrated into the lower chamber was quantified. The results revealed that both CCL5 alone and CHO-K1 alone had low chemotactic activity; in contrast, CHO-K1 in the presence of CCL5 showed significantly higher chemotactic activity (**Figure 3E**). In the same line, the chemotactic activity of CCL5 was much lower when incubated with CHO-677 in the lower chamber. As it was found that higher proportion of heparin reduced phase separation of CCL5 (**Figure 1A**), we tested whether this is the case in cells. Indeed, addition of heparin in the incubation of CHO-K1 abolished the phase separation of CCL5-Cy3 on the cells surface (**Figure 3A**, lower panel), accordingly attenuated the chemotactic activity of CCL5 (**Figure 3E**). When ^{44}AANA^{47}-CCL5 was incubated with CHO-K1 cell, no obvious phase-separated condensates appeared on the cell surface (**Figure 3—figure supplement 1**), and consequently, ^{44}AANA^{47}-CCL5 showed reduced chemotactic activity when incubated with CHO-K1 (**Figure 3E**) in comparison with WT CCL5. Thus, we may conclude that HS is essential on the CHO cell surface for phase separation of CCL5 and its chemotaxis function.

Based on the observation that CCL5 forms a concentration gradient on heparin-beads where heparin-driven phase separation enabled CCL5 diffuse, we hypothesized a similar scenario of HS on cells, e.g., CCL5 was immobilized or condensed by HS on the cell surface through phase separation, and gradually diffuse to form a gradient to guide cell migration. To test this conjecture, we transfected the construct of CCL5-EGFP into CHO-K1. The cells were co-cultured with Human Umbilical Vein Endothelial Cells (HUVEC)(Ynebrten, Barois et al., 2015). Released CCL5-EGFP from CHO-K1 cells was readily taken up by nearby HUVEC cells, formed bright phase-separated condensates on the cell surface (**Figure 4A and Figure 4— figure supplement 1A**). With extended incubation, the condensates became fused (**Figure 4B**), in a similar way of CCL5-heparin condensates in solution (**Figure 1C**). The FRAP experiment showed reasonable fluorescence recovery after photobleaching, indicating liquid-like properties of the condensates on the cell surface (**Figure 4C**). To further illustrate the gradient formation of the CCL5-EGFP condensates, we cultured HUVEC in Matrigel and embedded tiny amounts (about 400 cells) of the CCL5-EGFP transfected CHO-K1 as a source of CCL5-EGFP. Confocal microscopy monitoring found decreased concentration of CCL5-EGFP with increasing distance of HUVEC from the source cells (CCL5-EGFP expressing CHO), confirming that CCL5-EGFP established a gradient by phase separation on its target cells (**Figure 4D and E**, **Figure 4— figure supplement 1B and C**). Further, both CHO-K1 and CHO-677 transfected with CCL5-EGFP were analyzed on the same

setting. The results show that a gradient was formed in CHO-K1 cells when co-cultured with CCL5-EGFP transfected CHO-K1 in Matrigel, while CHO-677 failed to form the concentration gradient (**Figure 4—figure supplement 1D**). Furthermore, the chemotactic activity was demonstrated by placing CCL5-EGFP transfected CHO-K1 in a lower chamber and THP-1 in the upper chamber. Counting the transmigrated THP-1 cells showed that the transfected CHO-K1 had stronger chemotactic activity compared with WT-CHO-K1(**Figure 4F**).

Ex vivo and in vivo demonstration of HS-dependent chemotactic activity of CCL5

To further verify our findings, we placed HUVEC or CHO cells in the lower chamber in the presence of CCL5 and human peripheral blood cells (PBMC) in the upper chamber of the Transwell device to detect transmigration of the inflammatory cells. Quantification of the transmigrated cells into the lower chamber after incubation for 4 hr revealed that neither CCL5 alone nor CHO-677 in the presence of CCL5 showed substantial chemotactic activity; in contrast, HUVEC and CHO-K1 in the presence of CCL5 showed significantly higher chemotactic activity. It should be pointed out that the total PBMC in the 3 blood samples varied dramatically, resulting in the great deviations in the number of transmigrated cells of each sample. Nevertheless, the trend of chemotactic activity under the different conditions is consistent in all samples (**Figure 5**).

To verify our findings in vivo, wild-type or mutant CCL5 was injected into the peritoneal cavity of Balb/c mice and peritoneal cells recruitment was monitored(Proudfoot et al., 2003). Wild-type CCL5 induced a robust increase of total cells in the peritoneal lavage to a level approximately 4-fold over the saline control (**Figure 6**). In comparison, though ^{44}AANA^{47}-CCL5 group had a higher cell number than saline control, the recruited cells was much less than CCL5. Heparin (1 mg/mL) co-injected with wild-type CCL5 attenuated the recruitment of inflammatory cells, which, again, can be ascribed to the competitive interaction with endogenous HS in binding to CCL5. Overall, these results demonstrate that the phase separation of CCL5 with HS indeed contribute to chemotaxis function of CCL5 in vivo.

Discussion

It has been suggested that an important component of the chemokine-signaling is formation of a haptotactic gradient through immobilization of chemokines on cell surface glycosaminoglycans (GAGs)(Helen P. Makarenkova, 2009). Interactions with GAGs facilitate gradient formation of chemokine, providing directional cues for migrating cells. Heparan sulfate (HS) is one member of the GAGs family and ubiquitously expressed on the cells surface of endothelial and epithelial cells, forming glycocalyx(Simon Davis & Parish, 2013). It is known that HS (and its analogue heparin) binds to a broad spectrum of cytokines, functioning as a co-receptor to mediate signaling activity of the cytokines(Xie & Li, 2019). Several inflammatory chemokines, including CCL5, bind to HS/heparin(Proudfoot et al., 2003); however, the molecular mechanisms of HS in CCL5-induced chemotactic activity has not been reported.

Phase separation is a common mechanism for protein assembly and compartmentalization, and it contributes to a variety of cellular processes, including the formation of membraneless organelles, signaling complexes, the cytoskeleton and numerous other supramolecular assemblies(Hyman, Weber et al., 2014, Zbinden, Pérez-Berlanga et al., 2020). Emerging evidence indicates that biomolecules in phase-separated liquid droplets are mobile and transitorily interact with surrounding molecules(Nakashima, Vibhute et al., 2019). CC chemokines are found to occur oligomerization in vivo, which may be modulated by GAGs(Hoogewerf, Kuschert et al., 1997). Indeed, a conserved and positively charged BBXB motif in the CC chemokines is the key to interact with negatively charged GAGs through multivalent weak ionic interactions(Liang, Wenguang et al.,

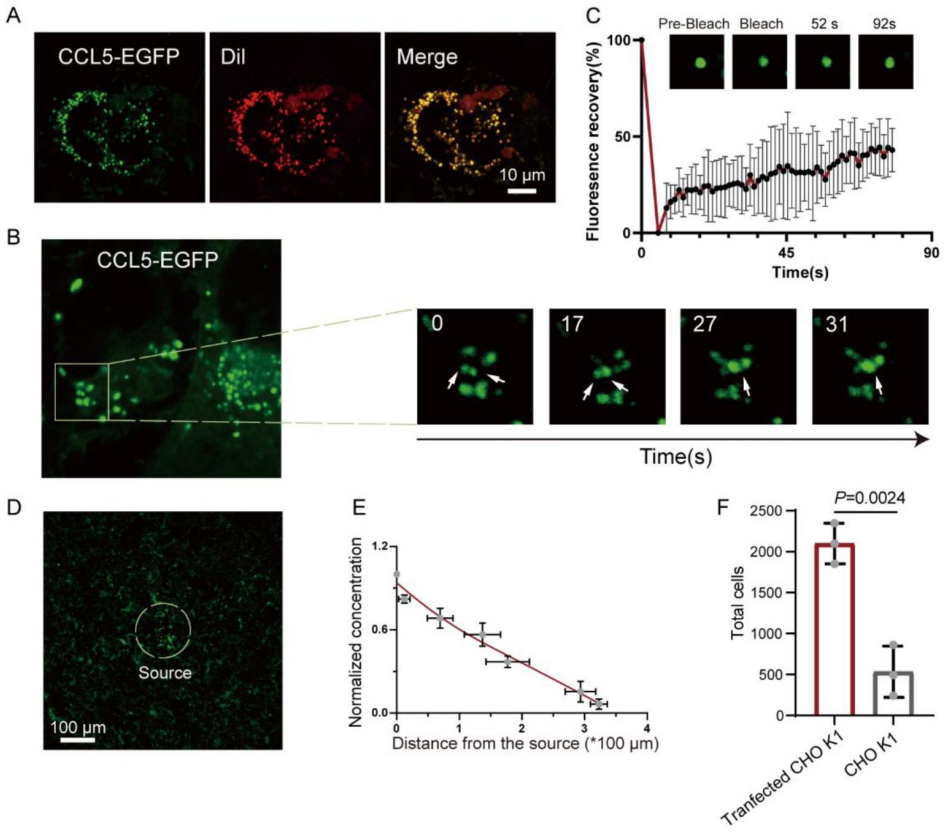

Figure 4

Formation of chemokine gradient on the cell surface by phase separation

(A) HUVEC cells were seeded on the plates for 24 h and stained with Dil (Cell membrane red fluorescent probe). After washing with PBS, CCL5-EGFP transfected CHO-K1 were added and co-cultured for 24 hr before taking the images (Scale bar=10 μm). (B) Fusion events of droplets formed by CCL5-EGFP on HUVEC cell surface. The white arrows in cropped images indicate the fusion of two droplets with time. (C) FRAP of the condensates formed by CCL5-EGFP in HUVEC cell surface, showing the intensity of fluorescence pre and after photobleaching (**Figure 4**-Source Data 1). The images of representative droplets in different recovery stages are shown. (D) HUVEC cells were seeded on the plate and firmly adhered before adding the 400 CCL5-EGFP transfected CHO-K1 cells placed in 50% of Matrigel. After 1 hr co-culture, confocal images were taking showing CCL5-EGFP diffusion on the surface of HUVEC. Dotted white circle indicates the source cell of CCL5-EGFP transfected CHO-K1. Scale bar=100 μm. (E) Quantification of the fluorescence intensity shows decreased signals of CCL5-EGFP as the distance from the source cells increased (**Figure 4**-Source Data 2). (F) The chemotaxis assay (same experimental conditions as described in **Figure 1**) shows higher activity of CCL5-EGFP transfected CHO-K1 cells than wild type CHO-K1 cells (**Figure 4**-Source Data 3). Data are mean ±s.d. n=3. Normal distribution was assessed by the Shapiro-Wilk (SW) normality test. *P* values were determined by unpaired two-tailed t-tests.

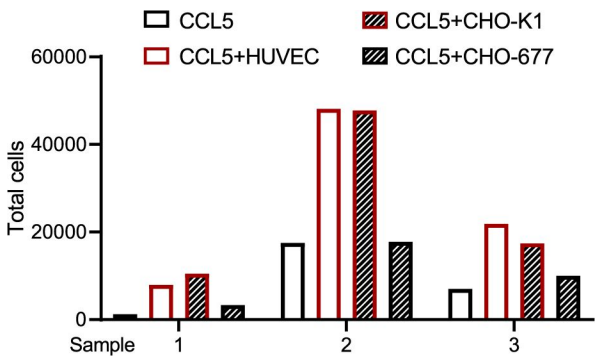

Figure 5

Transmigration of human peripheral blood cells

The cells (CHO-K1, CHO-677, or HUVEC) (1×10^5 cells/well) as indicated were plated onto the lower chamber for 24 hr (fully attached) and CCL5 was added for 1 hr. PBMC isolated from three healthy volunteer donors were placed in the upper chamber (total number of cells: Sample 1: 8.65^10^5; Sample 2: 2.31^10^6; Sample 3: 2.23^10^6). After culturing for 4 hr, the total transmigrated cells in the lower chamber were counted (**Figure 5** -Source Data 1).

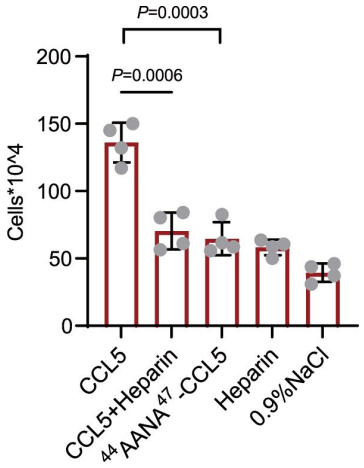

Figure 6

Chemokines phase separation promotes cell recruitment in vivo.

Mice were treated by intraperitoneal injection of CCL5 or the reagents as indicated. After 18h, the animals were sacrificed and peritoneal lavage was collected. Total cell number was counted by automated cell counter (**Figure 6** -Source Data 1). Data are mean ±s.d. n=4. Normal distribution was assessed by the Shapiro-Wilk (SW) normality test. *P* values were determined by unpaired two-tailed t-tests.

2016). Between the CCL5 dimers, Structural study revealed that the basic amino acids R44, K45, and R47 create two positively charged regions on the interface of dimer and interact with heparin(Lortat-Jacob, Grosdidier et al., 2002).

Using our established liquid-liquid phase separation (LLPS) method, we demonstrated that CCL5 phases separate on the cell surface via a heparan sulfate (HS) dependent mechanism, which is required for CCL5 chemotaxis. Mutations on the basic amino acids R44, K45, and R47 resulted in loosing binding of CCL5 to heparin beads, accordingly, lost its chemotactic activity. This finding also supports the fact that the phase separation of CCL5 with HS is mediated by weak electrostatic interactions, which facilitates the diffusion of CCL5 in solution to form a gradient.

It is well established that HS constitutes the major component of the glycocalyx on the endothelial cell surface(Oshima, King et al., 2021), and our earlier results showed that degradation of endothelial surface HS by heparanase impaired the function of MIP-2/CXCL2 induced leukocyte rolling, adhesion and transmigration(Massena, Christoffersson et al., 2010). From the finding of this study that the ratio of CCL5: heparin modulated droplets and condensates formation (**Figure 1A and B**), we may assume that HS on the cell surface should have a sufficient capacity to interact with chemokines, controlling their chemotactic activity and inflammatory cell migration. This assumption is supported by the finding that overexpressing CCL5 in CHO-K1 cells led to higher chemotactic activity, which may represent a scenario of acute inflammatory reaction.

Collectively, our studies provide the first evidence that CCL5 gradient formation and chemotactic activity are dependent on HS. The interaction between CCL5 and HS is through a weak multivalent electrostatic binding, which facilitates LLPS and diffusion of the chemokine (**Figure 7**). Notably, the results suggest that there may exist a physiological balance between CCL5 and HS, modulating the chemokine activity. This finding supports the hypothesis to develop chemokine-binding HS-mimetics that may competitively interfere with the HS-chemokine binding and phase separation, accordingly modulating inflammatory reactions.

Materials and Methods

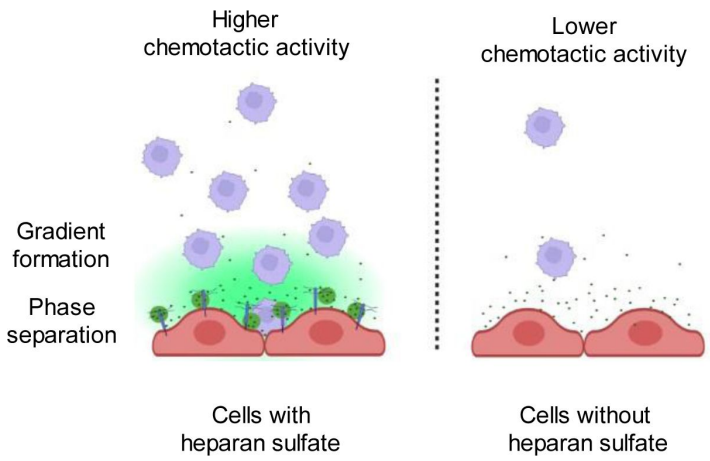

Figure 7

Schematic model of CCL5 phases separation along with heparan sulfate on the cell surface.

The inflammatory cells secrete chemokines that are immobilized by heparan sulfate in the form of Liquid-Liquid phase separation and subsequently diffused to form a concentration gradient.

sequence-based reagent	^{44}AANA^{47}-CCL5-mut-R	(Ynebrten et al., 2015)	PCR primers	TGGGTTGGCACACACTTGCGCGTTCGCCGCGGTGACAAAGACGACTGC
Recombinant DNA reagent	pET-28a	Miao Ling Plasmid	P51040	
Chemical compound, drug	Cy3-NHS	ATT Bioquest	1023	
Chemical compound, drug	heparin	GlycoNovo	C-HEPPIM	Mw:13.980 Da
cell line (Homo-sapiens)	HUVEC	Pricella	CL-0675	
Cell ine(Cricetulus griseus)	CHO-K1	GlycoNovo		
Cell ine(Cricetulus griseus)	CHO-677	GlycoNovo		
biological sample(Mus musculus)	BALB/cAnN Crl	Vital River	211	female

biological sample(Homo-sapiens)	Human peripheral blood cell	China-Japan Friendship Hospital		Freshly isolated from volunteers
software, algorithm	CHARMM-GUI	CHARMM-GUI	RRID:SCR_025037	
software, algorithm	GROMACS	GROMACS	RRID:SCR_014565	
software, algorithm	gmx_MMPBSA	GitHub	RRID:SCR_002630	
software, algorithm	LAS X software 3.5	Leica Application Suite X	RRID:SCR_013673	

Protein expression and purification

DNA sequences of wild-type human CCL5 were synthesized by General Biosystems, China. For the mutant of CCL5 (^{44}AANA^{47}-CCL5), site-directed mutations were introduced using the Quikchange mutagenesis kit (Beyotime Biotechnology, China) using mutagenesis primes: 5'-GCAGTCGTCTTTGTCACCGCGGCGAACGCGCAAGTGTGTGCCAACCCA-3'. CCL5 gene and ^{44}AANA^{47}-CCL5 gene were constructed in pET-28a vector (Miao Ling Plasmid, China) and transformed into *E.coli* Rosetta (DE3), (Biomed, China) for expression(Proudfoot et al., 1996, Proudfoot & Borlat, 2000). The products contained an additional Met residue at the N-terminus (Met-CCL5) and His tag or EGFP-fusion protein. Bacterium were grown to optical density of 0.6 at 37 °C and induced with 0.5 mM isopropyl-β-d-thiogalactopyranoside (IPTG) at 37 °C for 5 h. The cells were harvested by centrifugation (4000 rpm, 20 min at 4 °C), resuspended in lysis buffer (25 mM Tris-HCl, 100 mM NaCl, pH 8.0) and disrupted by sonication. The cell lysate was centrifuged at 12000 rpm for 35 min at 4 °C. To recover the recombinant CCL5 and ^{44}AANA^{47}-CCL5 proteins that were mainly distributed in the inclusion bodies, the pellet was collected and resuspended in denaturing buffer (100 mM Tris-HCl, 6 M guanidine-HCl, pH 8.0) and disrupted by sonication. Cell debris was removed by centrifugation (30000×g, 20 min at 4 °C). The proteins in the supernatant were refolded by dialysis in to refolding buffer (0.9 M guanidine-HCl, 100 mM Tris-HCl, 5 mM methionine, 5 mM cysteine, pH 8.0). After repeated change of dialysis buster, the protein debris was removed by centrifugation (30000×g, 20 min at 4 °C) and the supernatants were applied to His-Trap chelating column (GE Healthcare), washed by refolding buffer and eluted with the same buffer containing 30 mM imidazole. The purified proteins were dialyzed into 1.0 v/v% acetic acid aqueous solution twice and finally into 0.1%v/v trifluoroacetic acid aqueous solution. The purity of the recombinant CCL5 was analyzed by SDS-PAGE. After lyophilization, the CCL5 proteins were dissolved in water and stored in −80 °C.

For a better folding, the CCL5-EGFP and ^{44}AANA^{47}-CCL5-EGFP fusion proteins were expressed in *E.coli* OrigamiB(DE3) and induced by 0.5mM IPTG at 25 °C for 12 h. The proteins were purified by the same procedure as above.

Labeling of CCL5 with Cy3

Cy3 monosuccinimidyl ester (Cy3-NHS; ATT Bioquest, USA) was mixed with CCL5 to a final concentration of 1.5 mg/mL, pH 9 adjusted with 1M $NaHCO_3$. The mixture was incubated with shaking at 37 °C for 1 h, and then dialyzed in a 10 kDa dialysis tube (Thermo Fisher, USA) in ddH_2O. These Cy3 stained protein were mixed with unstrained protein at 1:40-1:20 ratio when used.

Cell culture

Human Umbilical Vein Endothelial Cells (HUVEC) were cultured in Dulbecco's modified Eagle's medium (DMEM) 1× with glucose (4.5 g/L), 10% fetal bovine serum (FBS), and 1% antibiotics (penicillin/streptomycin). CHO-K1/CHO-677 cells were cultured in F12K medium containing 10% FBS and 1% antibiotics. THP-1 were cultured in RPMI1640 medium containing 10% FBS and 1% antibiotics. All cells were cultured at 37 °C with 5% CO_2 in a humidified incubator.

Phase separation of CCL5 on cell surface

Cells were plated onto an eight-well Lab-Tek chambered coverglass (Thermo Fisher, USA) to around 70% confluency. Before imaging, the medium was discarded, and the cells were washed with PBS twice. Then, the protein solution diluted in the culture medium (500 nM) was added to the cells and incubation 1 hr in a cell incubator. Confocal microscopy image was captured with an inverted Leica DMi8 microscope, equipped with lasers for 489 nm, 554 nm excitation. Images were acquired using a 63×objective (oil immersion).

Z-stack for Living Cell 3-D rendering

Three-dimensional reconstruction for living cells were implemented with an inverted Leica DMi8 microscope. Images were acquired using the 63×oil immersion lens, a pinhole of 1 AU, 522 nm laser with 10% laser power, followed by setting the starting position and end position of Z-stack, 100~200 Nr. of Steps or 1 μm z-step size was selected.

Imaging of CCL5 phase separation *in vitro*

CCL5 were diluted to 4-6 mg/mL in KMEI buffer (150 mM KCl, 1 mM $MgCl_2$, 1 mM Ethylenebis (oxyethylenenitrilo) tetraacetic acid (EGTA) and 10 mM Imidazole, pH 6.5). For the co-phase separation of CCL5 and heparin (GlycoNovo, China, Mw:13.980 kDa), 20 μM of CCL5 were mixed with heparin and 5% w/v PEG-8000 in the assay buffer. The mixtures were incubated at 37 °C for 30 minutes then were loaded into a 96-well plate for imaging analysis. Images were captured with a Leica DMi8 confocal microscopy with a 63×objective (oil immersion) and LAS X software 3.5.

Turbidity assay

CCL5 proteins were mixed with various concentrations of heparin (0~1000 μM) and PEG-8000 (5% w/v) in the KMEI buffer. After incubation at 37 °C for 30 minutes the mixture was transferred to a 96-well plate and turbidity was measured by absorption at 620 nm using a Multiskan FC microplate reader (Thermo Fisher, USA). All samples were examined in triplicates (n = 3).

Molecular Docking

Heparin tetrasaccharide was docked to CCL5 (PDB number: 5coy) wild type and ^{44}AANA^{47}-CCL5 (PDB number: 1u4r) mutation by ClusPro server(Desta, Porter et al.). The docked models were selected according to the following standards: (1) close to the critical residues, i.e., aa 44-47, (2) having the lowest energy score. The selected CCL5-heparin tetrasaccharide complex was handled by Glycan Reader & Modeler module of CHARMM-GUI (Damm, Wolfgang et al., 1997, Gang, Liu et al., 2013) to set up the molecular dynamics simulation system and generate the input files.

The energy minimization (EM) simulations of the complexes were performed with GROMACS [*https://doi.org/10.1016/j.softx.2015.06.001*], and based on the EM trajectory, the binding free energy between CCL5 and heparin tetrasaccharide was calculated with gmx_MMPBSA(Van Wart, Durrant et al., 2014).

Diffusion of CCL5 on heparin beads and Ni-NTA beads

Heparin beads (Solarbio, China) and Ni-NTA beads (GenScript, China) 50 μL gel were mixed with 400 ng/μL of His-CCL5-EGFP and ^{44}AANA^{47}-CCL5-EGFP fusion proteins in a total volume of 500 μL. After incubation for 30 min in ice, the beads were dropped into Matrigel (ABWbionova, China) in 96-well plates kept on ice. Then the plates were moved to a 37 °C incubator for 12 h. Images were captured with a Zeiss Axiocam 506 color Fluorescence Microscope with a 5×, 10×objective and analyzed with ImageJ.

Chemotaxis assay

CHO-K1, mutant CHO-677 and HUVEC cells were cultured in the lower chamber of Transwell with 8 μm pores (Corning, USA) with a density 1×10^5 cells/well for 24 hr. Cells were washed three times with PBS and then 500 nM CCL5 or the mixed solution of 500 nM CCL5 and 1mg/mL heparin (GlycoNovo, China) in culture medium were added and incubation 1 hr in a cell incubator prior to the chemotactic assay. Then the THP-1 cells (3×10^5) in RPMI1640 medium were placed onto upper chamber of Transwell and incubated for 4 hr in a cell incubator, the transmigrated cells (in suspension) in the lower chamber were collected and counted by cell counter and inverted microscope.

For chemotaxis assay using beads, CCL5-EGFP and ^{44}AANA^{47}-CCL5-EGFP were first mixed with the beads as described above, washed and diluted with PBS and then placed on the bottom of lower chamber. The THP-1 cells were placed in the upper chamber. Cell migration analysis was performed as described above.

Diffusion of CCL5-EGFP in cell surface

CCL5-EGFP was constructed in pCDNA3.1-EGFP vector (General Biosystems, China) and transiently expressed in CHO-K1 cells. Briefly, the cells were seeded on 6-well plated at a density of 2.5×10^5 cells/well in 2 mL medium for 24 hr and then transfected using Lipo8000^{TM} Transfection Reagent (Beyotime Biotechnology, China). After 12 hr; transfected cells were detached by pancreatin (2000/mL). HUVEC cells were plated onto an eight-well Lab-Tek chambered coverglass (Thermo Fisher, USA) with a density 3×10^4 cells/well for 24 hr. The transfected CHO-K1 cells (2000/mL) in 200 μL medium were added to the eight-well coverglass coating HUVEC. After 1 hr, medium was aspirated and replaced with 50% of Matrigel. The transfected CHO-K1 was co-cultured with HUVEC cells for 12 hr, and images were captured with Leica DMi8 confocal microscopy with a 20×objective. For the capture of droplets on cell surface and FRAP, the cellular membrane of HUVEC was stained with Dil (Cell membrane red fluorescent probe, Beyotime, China) for 20 min and washed three times with PBS. Images were captured with a 100×objective (oil immersion).

Human peripheral blood cell transmigration

Blood (1 mL) was collected from three healthy volunteers (We did not collect demographics) in EDTA-tubes and centrifuged at 500×g (4°C for 5 min). After aspiration of plasma, the cells were treated with 10 mL of Red Blood Cell Lysis solution (Beyotime Biotechnology, China) for 10 min at 4°C and centrifuged. After removing supernatant, the cells were washed 3 times with PBS, and resuspended in 400 μL of RPMI1640 medium and counted. The cell (denoted as PBMC) suspension (100 μL) was added to the upper chamber for the chemotaxis assay as described above. All blood samples were obtained with informed consent, and the study was approved by the ethics review committee of China-Japan Friendship Hospital (2022-KY-050). All relevant ethical regulations of China-Japan Friendship Hospital and governmental regulations were followed.

Recruitment of inflammatory cells in peritoneal lavage

Eight-week-old female BALB/c mice were purchased from Charles River and kept in a pathogen-free environment with fed ad lib. The procedures for care and use of animals were approved by Beijing Municipal Science & Technology Commission, Administrative Commission of Zhongguancun Science Park (SYXK-2021-0056) and all applicable institutional and governmental regulations concerning the ethical use of animals were followed. Mice were injected intraperitoneally with 200 μL of NaCl (0.9%, lipopolysaccharide-free), 1 mg/ml heparin, 500 nM wild-type CCL5, 500 nM ^{44}AANA^{47}-CCL5 or 500 nM wild-type CCL5 added by 1mg/mL heparin diluted into 200 μL of NaCl (0.9%, lipopolysaccharide-free), respectively. At 18 h post-injection mice were sacrificed by cervical dislocation, the cells in the peritoneal cavity were collected by lavage of 5 mL ice-cold PBS. The total cells collected were counted with automated cell counter (Bodboge, China).

Acknowledgements

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Data and Materials availability statement

The authors confirm that the data supporting the findings of this study are available within the article and supplementary file.

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Conflict of interest

The authors declare that they have no conflicts of interest with the contents of this article.

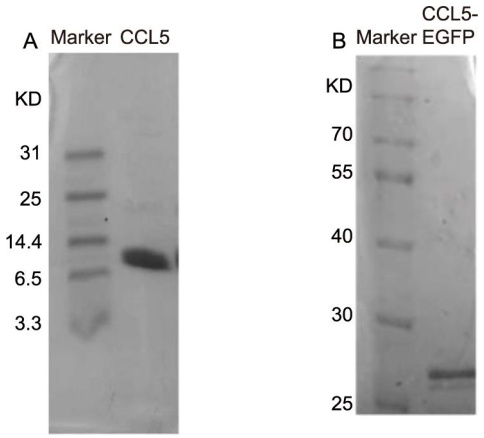

Figure1—Figure supplement 1

Protein expression and purification.

(A)-(B) SDS-PAGE showed the molecular weight and purity of CCL5 (A) and CCL5-EGFP (B).

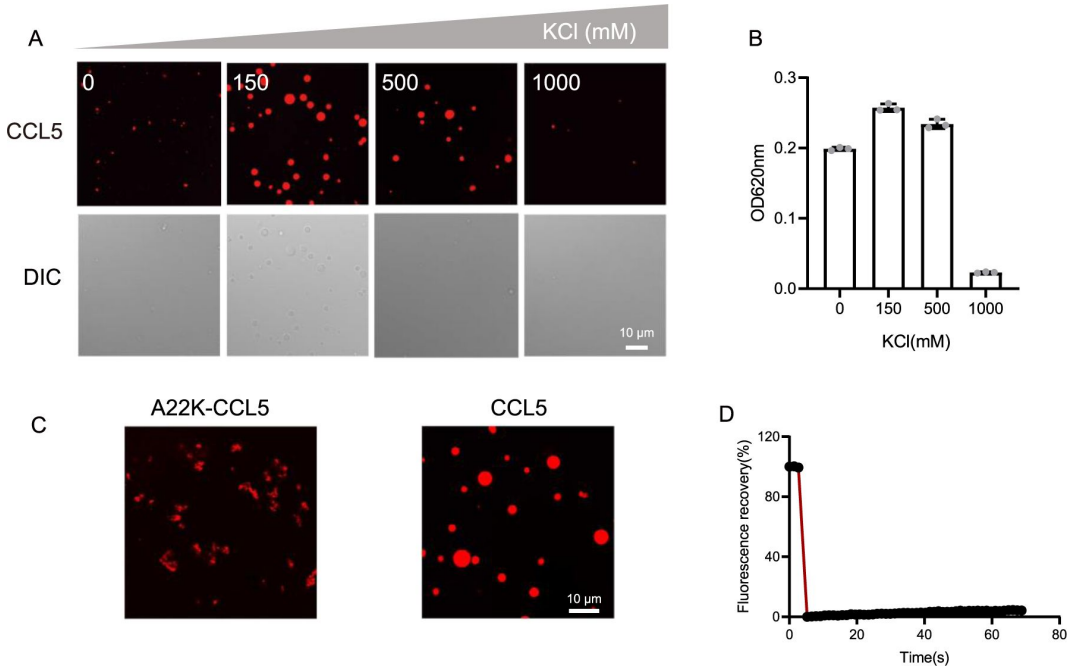

Figure 1—figure supplement 2

The effect of salt concentration on phase separation.

(A) Confocal images of assembling status of 20 μM CCL5-Cy3 mixed with heparin with increasing salt concentration. Scale bar=10 μm. (B) Turbidity changes with different increasing salt concentrations. (C) Confocal images of assembling status of A22K-CCL5 or CCL5 in the presence of heparin. Scale bar=10 μm. (D) FRAP of the aggregates formed by A22K-CCL5-Cy3, showing the intensity of fluorescence pre-and after photobleaching.

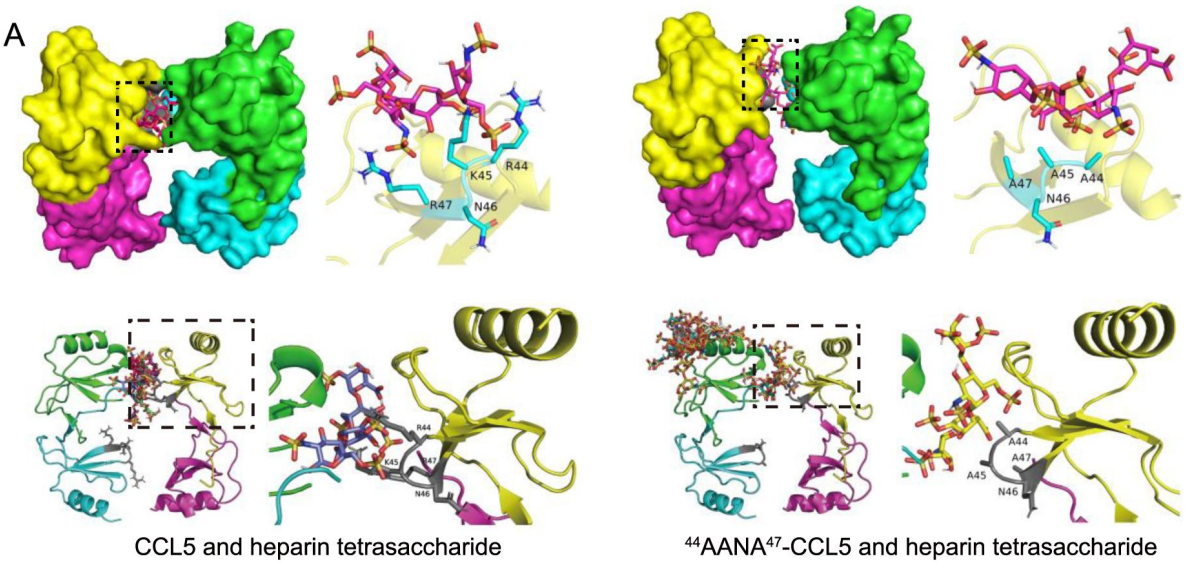

Figure 1—figure supplement 3

Comparison of binding of CCL5 and ^{44}AANA^{47}-CCL5 to heparin tetrasaccharide.

(A) Binding mode of heparin tetrasaccharide to ^{44}AANA^{47}-CCL5 or CCL5 shown by molecular docking. On the right are the details framed by the dotted line. (B) Comparison of binding free energy of different interacting forces between CCL5/^{44}AANA^{47}-CCL5 and heparin tetrasaccharide. The mutation led to change in electrostatic interaction forces.

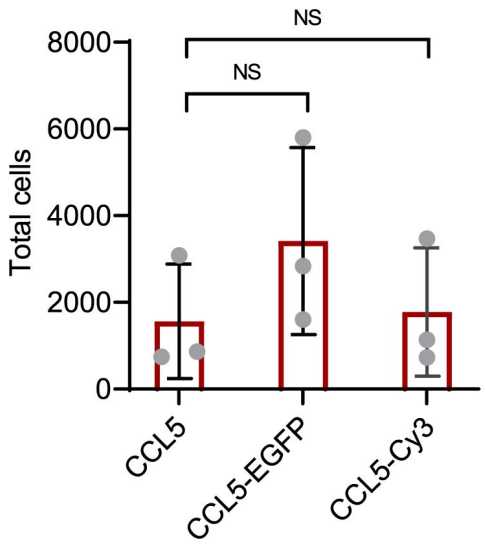

Figure 2—figure supplement 1

Chemotactic function of CCL5-EGFP.

Labeled CCL5 has similar activity as CCL5, 50 nM CCL5, CCL5-EGFP or CCL5-Cy3 were added to the lower chamber of the Transwell. THP-1 cells were added to upper chambers. Data are mean ± s.d. n=3. *P* values were determined by unpaired two-tailed t-tests. NS, Not Significant.

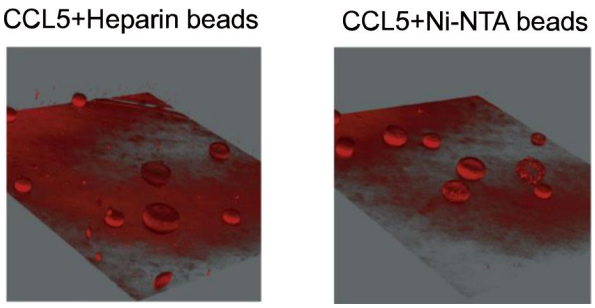

Figure 2—figure supplement 2

Diffusion of the CCL5-Cy3 in heparin-beads or Ni-NTA beads.

Beads bound with CCL5-Cy3 placed in Matrigel were imaged and reconstructed by Leica DMi8.

Figure 3—figure supplement 1

^{44}AANA^{47}-CCL5 didn't immobilize to the surface of CHO-K1.

CHO-K1 were treated with 500 nM ^{44}AANA^{47}-CCL5. From left to right, the pictures showed bright-field, fluorescent-field and overlay of two illuminations.

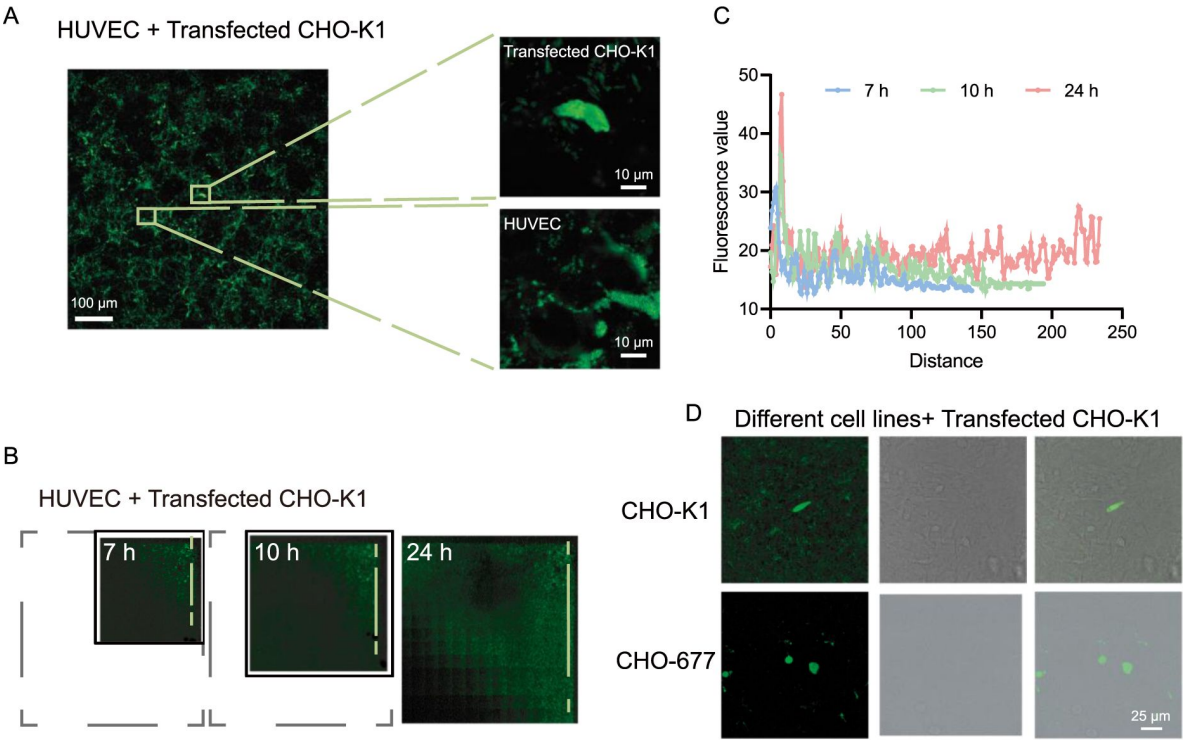

Figure 4—figure supplement 1

Diffusion of CCL5-EGFP on cell surface.

(A) Confocal images of Transfected CHO-K1 and HUVEC cells. Scale bar=100 μm. The yellow boxes are shown in higher magnification on the right. Scale bar=10 μm. CCL5-EGFP was expressed by transfected CHO-K1 and then adhere to the surface of HUVEC and formed phase separation. (B) The range of CCL5-EGFP diffusion on HUVEC cell surface at different time. The black dashed box showed the final image size. (C) Fluorescence intensity along the dotted lines indicated in **Figure 4**— figure supplement 1B. (D) Confocal images of Transfected CHO-K1 and CHO-K1 or CHO-677. CCL5-EGFP was expressed by transfected CHO-K1 and then adhere to the surface of CHO-K1 and formed phase separation but CHO-677 completely lack the signals. Scale bar=25 μm.

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Reviewer #2 (Public Review):

Although the study by Xiaolin Yu et al is largely limited to in vitro data, the results of this study convincingly improve our current understanding of leukocyte migration.

(1) The conclusions of the paper are mostly supported by the data and in the revised manuscript clarification is provided concerning the exact CCL5 forms (without or with a fluorescent label or His-tag) and amounts/concentrations that were used in the individual experiments. This is important since it is known that modification of CCL5 at the N-terminus affects the interactions of CCL5 with the GPCRs CCR1, CCR3 and CCR5 and random labeling using monosuccinimidyl esters (as done by the authors with Cy-3) is targeting lysines. The revised manuscript more clearly indicates for each individual experiment which form is used. However, a discussion on the potential effects of the modifications on CCL5 in the results and discussion sections is still missing.
(2) In general, authors used high concentrations of CCL5 in their experiments. In their reply to the comments they indicate that at lower CCL5 concentrations no LLPS is detected. This is important information since it may indicate the need for chemokine oligomerization for LLPS. This info should be added to the manuscript and comparison with for instance the obligate monomer CCL7 and another chemokine such as CXCL4 that easily forms oligomers may clarify whether LLPS is controlled by oligomerization.
(3) Statistical analyses have been improved in the revised manuscript.

https://doi.org/10.7554/eLife.93871.2.sa0

Author response:

The following is the authors' response to the original reviews.

> ***Public Reviews:***
>
> ***Reviewer #1 (Public Review):***
>
> *Summary:*
>
> *In their manuscript, Yu et al. describe the chemotactic gradient formation for CCL5 bound to - i.e. released from - glycosaminoglycans. The authors provide evidence for phase separation as the driving mechanism behind chemotactic gradient formation. A conclusion towards a general principle behind the finding cannot be drawn since the work focuses on one chemokine only, which is particularly prone to glycan-induced oligomerisation.*
>
> *Strengths:*
>
> *The principle of phase separation as a driving force behind and thus as an analytical tool for investigating protein interactions with strongly charged biomolecules was originally introduced for protein-nucleic acid interactions. Yu et al. have applied this in their work for the first time for chemokine-heparan sulfate interactions. This opens a novel way to investigate chemokine-glycosaminoglycan interactions in general.*

Response: Thanks for the encouragement of the reviewer.

> *Weaknesses:*
>
> *As mentioned above, one of the weaknesses of the current work is the exemplification of the phase separation principle by applying it only to CCL5-heparan sulfate interactions. CCL5 is known to form higher oligomers/aggregates in the presence of glycosaminoglycans, much more than other chemokines. It would therefore have been very interesting to see, if similar results in vitro, in situ, and in vivo could have been obtained by other chemokines of the same class (e.g. CCL2) or another class (like CXCL8).*

Response: We share the reviewer's opinion that to investigate more molecules/cytokines that interact with heparan sulfate in the system should be of interesting. We expect that researchers in the field will adapt the concept to continue the studies on additional molecules. Nevertheless, our earlier study has demonstrated that bFGF was enriched to its receptor and triggered signaling transduction through phase separation with heparan sulfate (PMID: 35236856; doi: 10.1038/s41467-022-28765-z), which supports the concept that phase separation with heparan sulfate on the cell surface may be a common mechanism for heparan sulfate binding proteins. The comment of the reviewer that phase separation is related to oligomerization is demonstrated in (Figure 1—figure supplement 2C and D), showing that the more easily aggregated mutant, A22K-CCL5, does not undergo phase separation.

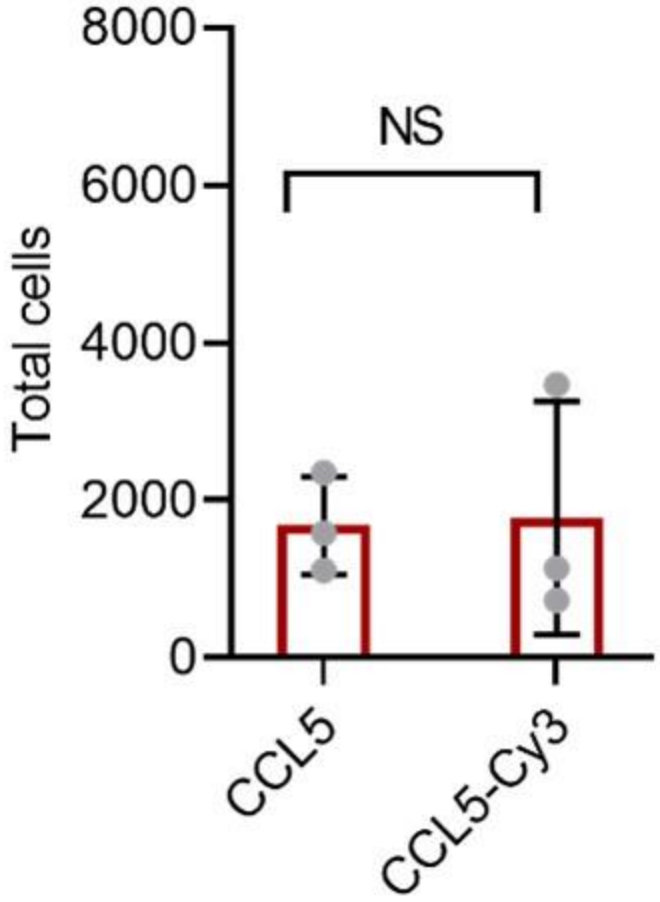
> *In addition, the authors have used variously labelled CCL5 (like with the organic dye Cy3 or with EGFP) for various reasons (detection and immobilisation). In the view of this reviewer, it would have been necessary to show that all the labelled chemokines yield identical/similar molecular characteristics as the unlabelled wildtype chemokine (such as heparan sulfate binding and chemotaxis). It is well known that labelling proteins either by chemical tags or by fusion to GFPs can lead to manifestly different molecular and functional characteristics.*

Response: We agree with the reviewer that labeling may lead to altered property of a protein, thus, we have compared chemotactic activity of CCL5 and CCL5-EGFP (Figure 2—figure supplement 1). To further verify this, we performed additional experiment to compare chemotactic activity between CCL5 and Cy3-CCL5 (see Author response image 1). For the convenience of readers, we have combined the original Figure 2—figure supplement 1 with the new data (Figure R1), which replaced original Figure 2—figure supplement 1.

Author response image 1.

Chemotactic function of CCL5-EGFP and CCL5-Cy3. Cy3-Labeled CCL5 has similar activity as CCL5, 50 nM CCL5 or CCL5-Cy3 were added to the lower chamber of the Transwell. THP-1 cells were added to upper chambers. Data are mean ± s.d. n=3. P values were determined by

unpaired two-tailed t-tests. NS, Not Significant.



Reviewer #2 (Public Review):

Although the study by Xiaolin Yu et al is largely limited to in vitro data, the results of this study convincingly improve our current understanding of leukocyte migration.

(1) The conclusions of the paper are mostly supported by the data although some clarification is warranted concerning the exact CCL5 forms (without or with a fluorescent label or His-tag) and amounts/concentrations that were used in the individual experiments. This is important since it is known that modification of CCL5 at the N-terminus affects the interactions of CCL5 with the GPCRs CCR1, CCR3, and CCR5 and random labeling using monosuccinimidyl esters (as done by the authors with Cy-3) is targeting lysines. Since lysines are important for the GAG-binding properties of CCL5, knowledge of the number and location of the Cy-3 labels on CCL5 is important information for the interpretation of the experimental results with the fluorescently labeled CCL5. Was the His-tag attached to the N- or C-terminus of CCL5? Indicate this for each individual experiment and consider/discuss also potential effects of the modifications on CCL5 in the results and discussion sections.

Response: We agree with the reviewer that labeling may lead to altered property of a protein, thus, we have compared chemotactic activity of CCL5 and CCL5-EGFP (Figure 2—figure supplement 1). To further verify this, we performed additional experiment to compare

chemotactic activity between CCL5 and Cy3-CCL5 (see Author response image 1). For the convenience of readers, we have combined the original Figure 2—figure supplement 1 with the new data (Author response image 1), which replaced original Figure 2—figure supplement 1.

The His-tag is attached to the C-terminus of CCL5, in consideration of the potential impact on the N-terminus.

> *(2) In general, the authors appear to use high concentrations of CCL5 in their experiments. The reason for this is not clear. Is it because of the effects of the labels on the activity of the protein? In most biological tests (e.g. chemotaxis assays), unmodified CCL5 is active already at low nM concentrations.*

Response: We agree with the reviewer that the CCL5 concentrations used in our experiments were higher than reported chemotaxis assays and also higher than physiological levels in normal human plasma. In fact, we have performed experiments with lower concentration of CCL5, where the effect of LLPS was not seen though the chemotactic activity of the cytokine was detected. Thus, LLPS-associated chemotactic activity may represent a scenario of acute inflammatory condition when the inflammatory cytokines can increase significantly.

> *(3) For the statistical analyses of the results, the authors use t-tests. Was it confirmed that data follow a normal distribution prior to using the t-test? If not a non-parametric test should be used and it may affect the conclusions of some experiments.*

Response: We thank the reviewer for pointing out this issue. As shown in Author response table 1, The Shapiro-Wilk normality test showed that only two control groups (CCL5 and 44AANA47-CCL5+CHO K1) in Figure 3 did not conform to the normal distribution. The error was caused by using microculture to count and calculate when there were very few cells in the microculture. For these two groups, we re-counted 100 μL culture medium to calculate the number of cells. The results were consistent with the positive distribution and significantly different from the experimental group (Author response image 3). The original data for the number of cells chemoattractant by 500 nM CCL5 was revised from 0, 247, 247 to 247, 123, 370 and 500 nM 44AANA47 +CHO-K1 was revised from 1111, 1111, 98 to 740, 494, 617. The revised data does not affect the conclusion.

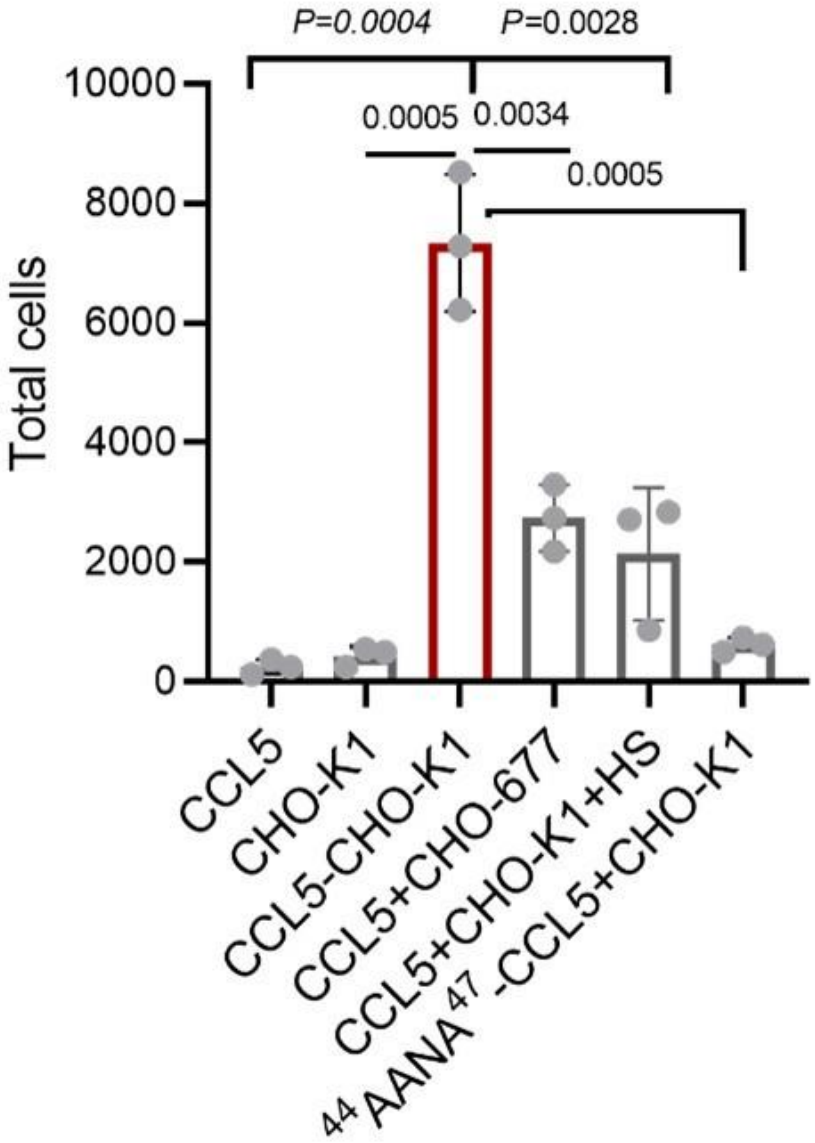
Author response table 1.

Table R1 Shapiro-Wilk test results of statistical data in the manuscript

Figure	Group	W	P value	Passed normality test(α=0.05)
1F	CCL5	0.9709	0.6725	Yes
	^{44}AANA^{47}-CCL5	0.9826	0.7470	Yes
2D	CCL5+Heparin beads	0.9425	0.5376	Yes
	CCL5+Ni beads	0.9776	0.7130	Yes
	^{44}AANA^{47}-CCL5+Heparin beads	0.9234	0.4606	Yes
3E	CCL5	0.75		
	CCL5 revision	**1**	**0.9955**	**Yes**
	CHO K1	0.8777	0.3176	Yes
	CCL5+CHO K1	0.9981	0.9174	Yes
	^{44}AANA^{47}-CCL5+CHO K1	0.75		
	^{44}AANA^{47}-CCL5+CHO K1 revision	**1**	**>0.9999**	**Yes**
	CCL5+CHO 677	1.0	0.9990	Yes
	CCL5+CHO K1+HS	0.7965	0.1062	Yes
4	Transfected CHO K1	1.000	＞0.9999	Yes
	CHO K1	0.9879	0.7896	Yes
6	CCL5	0.9436	0.6765	Yes
	CCL5+Heparin	0.8688	0.2928	Yes
	^{44}AANA^{47}-CCL5	0.8016	0.1052	Yes

Author response image 3.

Quantification of THP-1collected from the lower chamber. Data are mean ± s.d. n=3. P values were determined by unpaired two-tailed t-tests.



Recommendations for the authors:

Reviewer #1:

See the weaknesses section of the Public Review. In addition, the authors should discuss the X-ray structure of CCL5 in complex with a heparin disaccharide in comparison with their docked structure of CCL5 and a heparin tetrasaccharide.

Response: Our study, in fact, is strongly influenced by the report (Shaw, Johnson et al., 2004) that heparin disaccharide interaction with CCL5, which is highlighted in the text (page5, line100-102).

> ***Reviewer #2:***
>
> *(1) Clearly indicate in the results section and figure legends (also for the supplementary figures) which form and concentration of CCL5 is used.*

Response: The relevant missing information is indicated across the manuscript.

> *(2) Clearly indicate which GAG was used. Was it heparin or heparan sulfate and what was the length (e.g. average molecular mass if known) or source (company?)?*

Response: Relevant information is added in the section “Materials and Methods.

> *(3) Line 181: What do you mean exactly with "tiny amounts"?*

Response: “tiny amounts” means 400 transfected cells. This is described in the section of Materials and Methods. It is now also indicated in the text and legend to the figure.

> *(4) Lines 216-217: This is a very general statement without a link to the presented data. No combination of chemokines is used, in vivo testing is limited (and I agree very difficult). You may consider deleting this sentence (certainly as an opening sentence for the Discussion).*

Response: We appreciate very much for the thoughtful suggestion of the reviewer. This sentence is deleted in the revised manuscript.

> *(5) Why was 5h used for the in vitro chemotaxis assay? This is extremely long for an assay with THP-1 cells.*

Response: We apologize for the unclear description. The 5 hr includes 1 hr pre- incubation of CCL5 with the cells enable to form phase separation. After transferring the cells into the upper chamber, the actual chemotactic assay was 4 hr. This is clarified in the Materials and Methods section and the legend to each figure.

> *(6) Define "Sec" in Sec-CCL5-EGFP and "Dil" in the legend of Figure 4.*

Response: The Sec-CCL5-EGFP should be “CCL5-EGFP”, which has now been corrected. Dil is a cell membrane red fluorescent probe, which is now defined.

> *(7) Why are different cell concentrations used in the experiment described in Figure 5?*

Response: The samples were from three volunteers who exhibited substantially different concentrations of cells in the blood. The experiment was designed using same amount of blood, so we did not normalize the number of the cell used for the experiment. Regardless of the difference in cell numbers, all three samples showed the same trend.

> *(8) Check the text for some typos: examples are on line 83 "ratio of CCL5"; line 142 "established cell lines"; line 196 "peripheral blood mononuclear cells"; line 224 "to mediate"; line 226 "bind"; line 247 "to form a gradient"; line 248 "of the glycocalyx"; line 343 and 346 "tetrasaccharide"; line 409-410 "wild-type"; line 543 "on the surface of CHO-K1 and CHO-677"; line 568 "white".*

Response: Thanks for the careful reading. The typo errors are corrected and Manuscript was carefully read by colleagues.