

Compressed sensing based approach identifies modular neural circuitry driving learned pathogen avoidance

Reviewed Preprint

v1 • June 5, 2024

Not revised

Timothy Hallacy , Niels Ringstad, Sharad Ramanathan 

Biophysics Program, Harvard University, Cambridge, MA, USA • Department of Cell Biology, Skirball Institute of Biomolecular Medicine, New York University Grossman School of Medicine, New York, NY 10016, USA •

Department of Molecular and Cellular Biology, Department of Stem Cell and Regenerative Biology, John A.

Paulson School of Engineering and Applied Sciences, Harvard University, Cambridge, MA, USA

 https://en.wikipedia.org/wiki/Open_access Copyright information

Abstract

An animal's survival hinges on its ability to integrate past information to modify future behavior. The nematode *C. elegans* adapts its behavior based on prior experiences with pathogen exposure, transitioning from attraction to avoidance of the pathogen. A systematic screen for the neural circuits that integrate the information of previous pathogen exposure to modify behavior has not been feasible because of the lack of tools for neuron type specific perturbations. We overcame this challenge using methods based on compressed sensing to efficiently determine the roles of individual neuron types in learned avoidance behavior. Our screen revealed that distinct sets of neurons drive exit from lawns of pathogenic bacteria and prevent lawn re-entry. Using calcium imaging of freely behaving animals and optogenetic perturbations, we determined the neural dynamics that regulate one key behavioral transition after infection: stalled re-entry into bacterial lawns. We find that key neuron types govern pathogen lawn specific stalling but allow the animal to enter nonpathogenic *E. coli* lawns. Our study shows that learned pathogen avoidance requires coordinated transitions in discrete neural circuits and reveals the modular structure of this complex adaptive behavioral response to infection.

eLife assessment

This **important** study describes a neural circuit contributing to two behavioral processes affecting pathogen avoidance in the nematode *C. elegans*. The method used to identify specific contributing neurons is innovative and the experimental evidence supporting the major claims is **solid**. This study will be of interest to neuroscientists studying behavior, in particular in *C. elegans*.

<https://doi.org/10.7554/eLife.97340.1.sa3>

Introduction

Animals use past experiences to modify future behavior, and this behavioral plasticity is essential for an animal's fitness and survival. Previous studies suggest that small sets of neurons, such as those in the mushroom bodies of flies^{1,2,3,4} or in the human hippocampus^{5,6,7,8,9}, can play pivotal roles in integrating information from prior experiences and using it to influence subsequent behavior. Identification of these key neurons represents a crucial step towards understanding both the neural and molecular mechanisms responsible for encoding experiences and modulating behavior.

Despite its small nervous system, the nematode *Caenorhabditis elegans* displays a robust capacity to modify its behavior based on experience^{10,11,12,13,14}. One example of this is learned pathogen avoidance. Worms are initially attracted to pathogenic bacteria such as *Pseudomonas aeruginosa* (strain PA14). However, after several hours of exposure, animals associate infection with PA14 specific cues and change their behavior to avoid these bacteria^{15,16,17,18,19,20}. This behavioral transition reduces the chance of infection and thus increases the worm's odds of survival^{18,21,22,23}. How does the worm's nervous system encode this pathogen experience and use this information to change behavior? Previous studies uncovered a range of sensory cues that govern the worm's behavioral transition, ranging from chemosensory cues^{24,25,26,27} and the worm's innate immune response^{16,28} to mechanosensory inputs arising from the bacteria's biofilm^{29,30,31}. Olfactory stimuli have also been implicated in learned pathogen avoidance through serotonin modulation^{18,16}. The underlying neural circuits that encode information about past experiences and their dynamics remain unknown despite this. One strategy to find such neurons would be to carry out a comprehensive screen of the nervous system. However, such a screen is technically challenging due to the lack of neuron subtype-specific promoters and the necessity of systematic timed perturbations of neural activity during pathogen exposure to disrupt the animal's ability to learn to avoid pathogens in the future.

To discover neurons whose activity patterns governed experience-dependent pathogen avoidance, we performed a comprehensive screen of the nervous system using a compressed sensing based approach³² that overcomes these technical challenges. Through this screen, we discovered that the behavioral transition from attraction to avoidance of PA14 involves transitions in two subbehaviors - exit from the bacterial lawn and lawn re-entry. We found that distinct sets of neurons regulate each subbehavior. In particular, the aversion to re-entry is controlled by two neuronal types, AIY and SIA, which encode pathogen exposure through sustained downregulation of neural activity. Our data further indicated that AIY regulates PA14 specific aversion, and drives stalling at the edge of the pathogenic lawn. Finally, we used the identification of neural substrates of learned pathogen avoidance to explore how long-term changes in neural dynamics might be mediated by neuropeptide signaling between critical neurons that govern transitions in this behavior.

Results section

Prior pathogen exposure alters two behavioral modules to generate learned pathogen avoidance

Infection of the nematode *C. elegans* by pathogenic bacteria elicits learned pathogen avoidance behavior. Naïve animals dwell and feed on lawns of pathogenic *P. aeruginosa* (PA14), but after infection, animals avoid *P. aeruginosa* lawns^{15,29}. To obtain high-resolution measurements of

learned pathogen avoidance behavior, we monitored animals that had been placed on a lawn of pathogenic PA14 for 18 hours. We counted the fraction of animals that remained on the lawn over time (**Fig. 1A** [↗](#)) at 30min intervals. Within eight hours, half of the animals changed their foraging behavior and left the lawn of pathogenic bacteria. By contrast, nearly all the animals placed on a control lawn of non-pathogenic *E. coli* (OP50) bacteria remained on the lawn during 18 hours of monitoring (**Fig. 1A** [↗](#)).

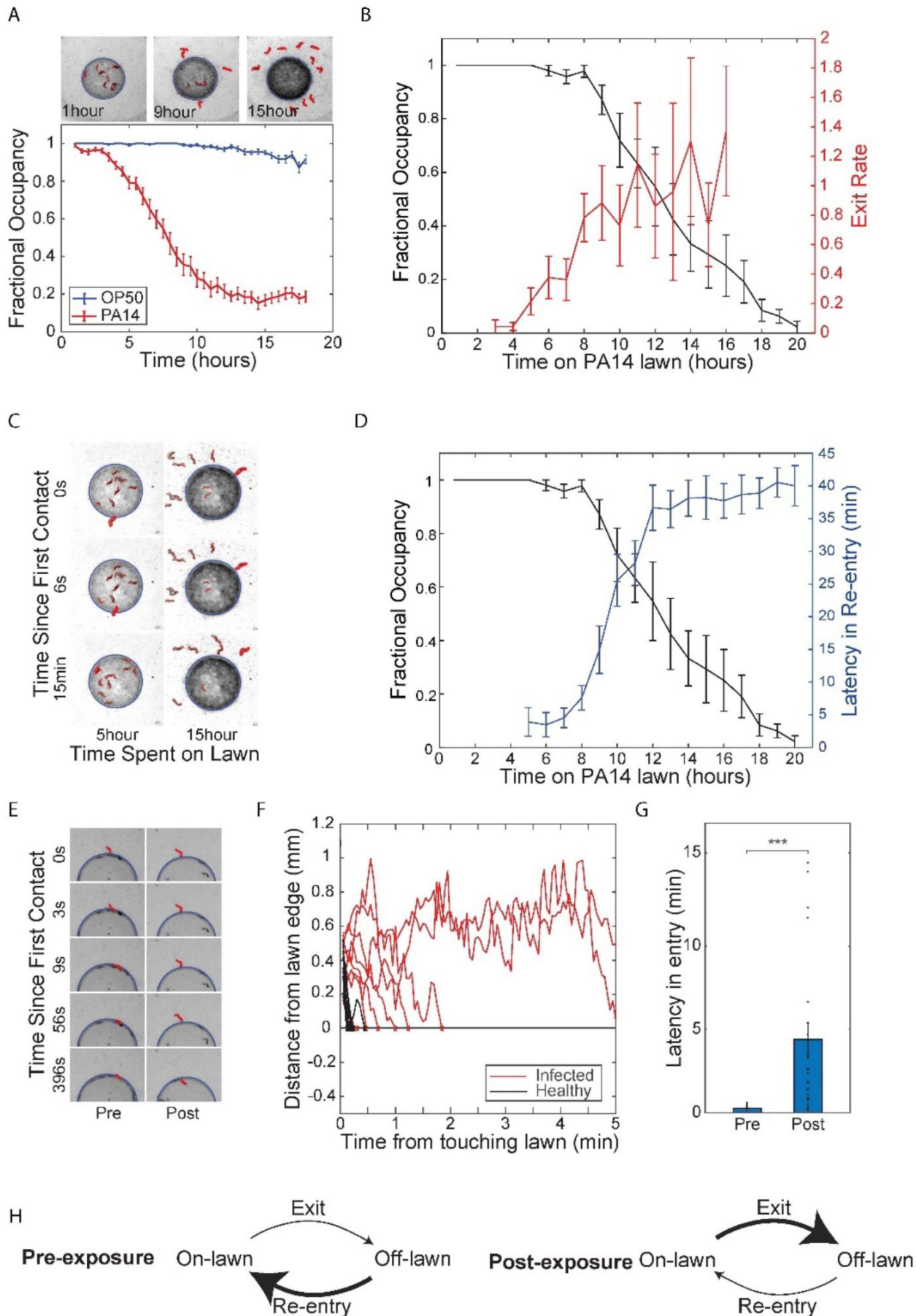


Fig. 1.

Exposure to pathogenic bacteria inhibits the ability of worms to re-enter the bacteria lawn.

(A) Above: representative sequence of phase contrast images of *P. aeruginosa* PA14 lawn evacuation at t=1, 9 and 15 hrs after deposition of *C.elegans* onto lawn. Evacuated worms are highlighted in red using imaging processing. Below: fractional occupancy of *C.elegans* on bacteria lawns plotted against time spent on pathogenic PA14 lawns (red) or non-pathogenic *E.coli* OP50 lawns (blue). Mean occupancy and SEM over n = 25 independent experiments with 10 worms each. (B) *C.elegans* fractional occupancy (black) and the rate constant of *C.elegans* exit (exits events/hour/animal, red) plotted against time spent on pathogenic PA14 lawns. The exit rate constant increased with time spent on the lawn. Mean and SEM over n = 5 samples of 10 worms each. (C) Representative phase contrast images of a worm attempting to re-enter PA14 lawn after 5 hours (left column) and 15 hours (right column) of PA14 exposure. Time labels denote time after first contact of worm with bacteria lawn. (D) Latency in re-entry (delay in the re-entry of the animal onto the bacteria lawn upon first contact with the lawn, red) and fractional occupancy (black) plotted against duration of exposure for worms on PA14 lawns. Mean and SEM over n = 5 samples of 10 worms each. (E) Representative sequence of phase contrast images of a worm (highlighted in red) attempting to enter a fresh PA14 bacterial lawn pre PA14 exposure (left) and post 15 hours of PA14 exposure (right). Time labels denote time after first contact of worm with bacteria lawn. (F) Representative plots of distance of center of mass of each animal from the edge of the PA4 lawn versus time after first contact with lawn, pre (black, n = 11) and post 15 hours of PA14 exposure (red, n = 9). (G) Latency of entry of worms pre (n=35) and 15 hours post (n=29) PA14 exposure. Post exposed worms increased latency of entry tenfold compared to pre-exposed control. (H) Model for lawn evacuation. Pre-exposed worms (left) have high re-entry rates and low exit rates. Exposure to pathogenic bacteria causes a transition in behavior (right), where worms increase exit rate and decrease re-entry rate, leading to net lawn evacuation.

The fractional occupancy of worms on a bacterial lawn is affected in principle by two processes - lawn exit and lawn re-entry. We determined how PA14 exposure affected each of these processes. After four hours in the presence of pathogen, the rate constant for the lawn exit (exits per animal per hour) increased dramatically from zero and reached a plateau of roughly 1 exit event per worm-hour after ten hours (Fig. 1B). We observed that worms that left lawns of pathogen after 10-16 hours of exposure repeatedly attempted to return to the lawn but stalled for long periods upon contact with the lawn edge (Fig. 1C). We quantified this re-entry defect by measuring the latency to re-entry, i.e., the delay between first contact with the lawn and re-entry of the animal into the lawn. Animals that had been exposed to pathogen for short periods rapidly re-entered the lawn after an exit with latencies of only a few seconds (Fig. 1D). After five hours of exposure to pathogen, the latency of re-entry began to increase and after twelve hours of exposure the mean latency to re-entry was 36.7 ± 3.5 mins (Fig. 1D). Increases in lawn-leaving rates and latency to re-entry were correlated and coincided with the observed evacuation of the lawn of pathogenic bacteria (Fig. 1D).

To demonstrate that the experience of pathogen exposure was driving the observed changes in behavior, as opposed to some change in the bacterial lawn caused by foraging animals, we exposed animals to pathogenic PA14 for 15 hours, collected those that had left the lawn, and then compared the latency of lawn entry of this cohort to the latencies of a cohort of naive animals that had not experienced pathogen. Naive worms rapidly entered a fresh lawn of pathogen whereas experienced worms displayed increased latency of entry into such fresh lawns (Fig. 1E-G). These results indicated that pathogen exposure changed the internal state of the worm to drive the observed behavioral transition from re-entering the law rapidly to stalled re-entry (Fig. 1H).

A compressed sensing based optogenetic screen to identify neurons that function in learned pathogen avoidance

We next sought to identify neurons that regulate interactions of *C. elegans* with lawns of pathogenic *P. aeruginosa*. Conventional approaches to identifying neurons required for a *C. elegans* behavior involve targeting individual neuron types by microablation^{33,34,35} or optogenetics^{36,37,38}. These approaches are time-consuming and can require the generation of large numbers of transgenic lines to individually interrogate specific neuron types for their role in a given behavior.

We previously showed that an optogenetic screen for neurons that drive a behavior can be performed more efficiently using multiplexed optogenetic manipulations of neurons followed by a compressed sensing analysis to infer individual key neuron types^{32,39,40}. We performed a compressed sensing based screen using a panel of 29 transgenic *C. elegans* lines, each expressing the light-gated ion channel Arch3 under a different promoter (**Supplementary Table 1**). The panel used for this study, focused on interneurons, and covered 54 classes of interneuron, 25 classes of sensory neuron, and 8 classes of motor neuron^{41,42,43}. We optically inhibited neurons during the early stages of pathogen exposure when animals presumably associate pathogen-specific cues with sickness. We then monitored the subsequent dynamics of lawn leaving and re-entry. Animals were transferred to pathogenic lawns, given 1 hour to settle, and then illuminated for two hours with pulses of 525 nm green light (1sec on/off, 5mW/mm²) to inhibit neurons expressing Arch3 in that line (**Fig. 2A**). The behavior of these animals was then recorded for a total of 18 hours at 3min intervals (**Fig. 2A**). We performed the same measurements of matched controls that had not been fed the opsin cofactor all-trans retinal and were thus insensitive to photoinhibition (**Fig. 2A**). To quantify the effect of optogenetic inhibition on behavior, we calculated for each strain a differential retention index - the difference in the area between the temporal lawn occupancy curves of transgenic animals exposed to inhibitory light and a paired no-ATR control (**Fig. 2B**). Six strains showed significant changes in differential retention index after neural silencing compared to controls (**Fig. 2B**). Silencing neurons in these strains during the first two hours of pathogen exposure augmented lawn-leaving over the next 18 hours (**Fig. S1**). These results indicated the existence of a neural mechanism that inhibits the association of pathogenicity with microbe-specific cues during the early stages (first two hours) of pathogen exposure.

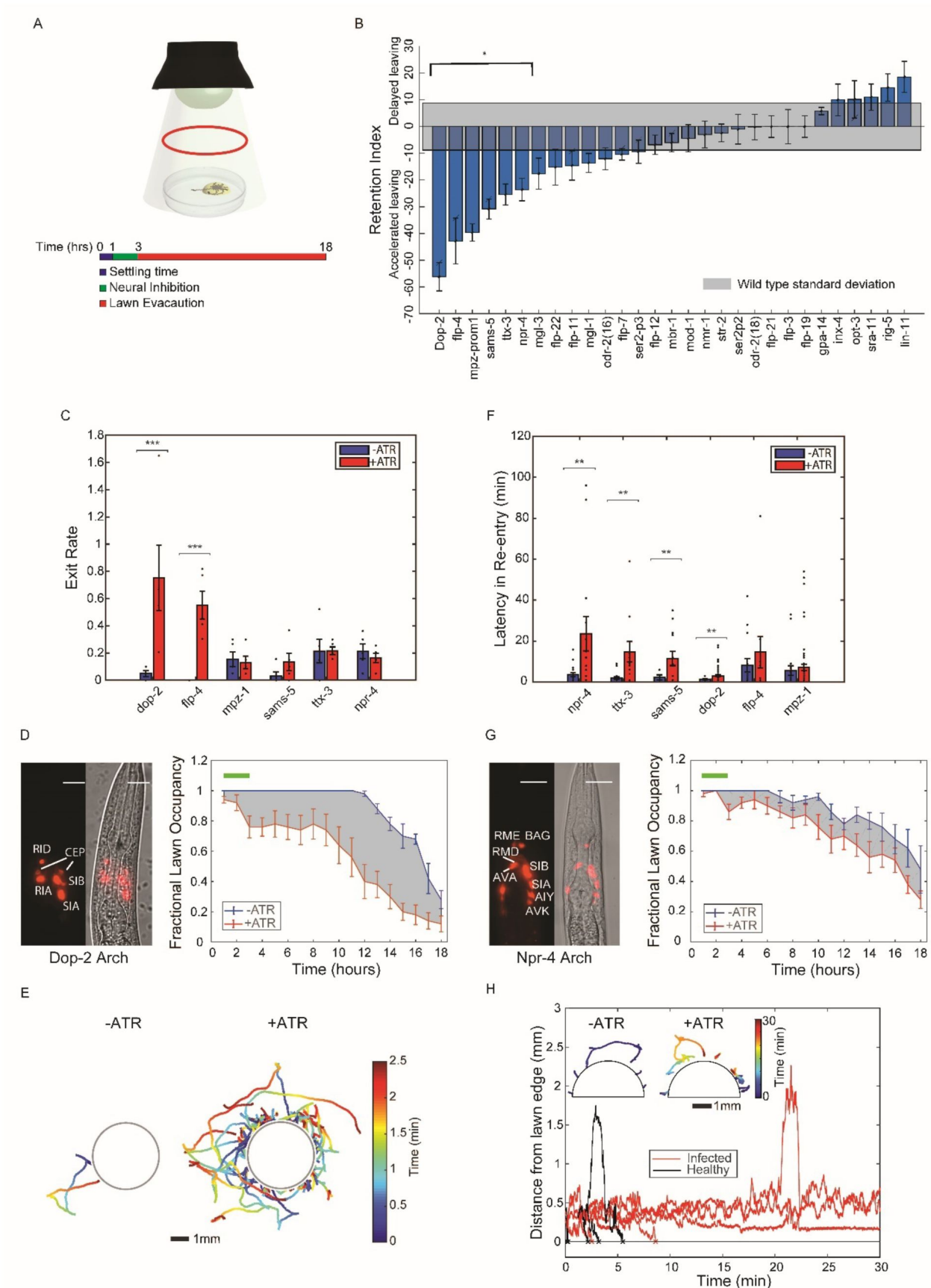


Fig. 2.

An optogenetic screen of the *C.elegans* nervous system to identify neurons controlling exit and re-entry into pathogenic bacteria lawns.

(A) Experimental protocol used to identify neurons that modulate long term changes in PA14 lawn evacuation dynamics. In each experiment, animals of one transgenic line expressing archaerhodopsin were placed onto a lawn of PA14 and allowed to settle for 1 hour (blue segment, in schematic). Worms were illuminated with green light (525nm) to inhibit activity for 2 hours in neurons expressing archaerhodopsin (green segment, in schematic). Worm evacuation was monitored over an additional 15 hours (red time segment, schematic). **(B)** Differential retention metric for 29 transgenic lines. The differential retention metric for each transgenic line was determined by measuring the difference in the area under lawn evacuation curves with and without neural inhibition (see methods). Differential retention metrics were compared against the standard deviation in retention metrics from uninhibited control worms (gray) to determine statistical significance (see methods). Six of the twenty nine lines (*dop-2*, *flp-4*, *mpz-prom1*, *sams-5*, *ttx-3*, and *npr-4*) showed statistically significant differential retention metrics. Mean and SEM over $n = 5$ samples of 10 worms each. **(C)** Rate constant of exit (exits events/hour/animal) for statistically significant transgenic lines (B). Four lines showed statistically significant changes in rate constant (*dop-2*, *sams-5*, *ttx-3*, and *npr-4*) between inhibition (red, +ATR) and no inhibition (blue, -ATR) control. Mean and SEM over $n = 5$ samples of 10 worms each. **(D)** Left: fluorescence and phase image of *Pdop-2::ARCH-mCherry* (C). Scale bar 15um. Right: fractional lawn occupancy of *Pdop-2::ARCH-mCherry* as a function of time (right) with (red, +ATR) and without (blue, -ATR) neural inhibition. **(E)** Trajectories of worms exiting the lawn either with (left) or without (right) inhibition of neurons expressing *dop-2* (D). Trajectories are color coded by time and are over $n = 2$ samples of 10 worms each. **(F)** Latency in re-entry (time of re-entry of the animal onto the bacteria lawn after first contact) of statistically significant transgenic lines (B). Four lines show statistically significant differences in latency in re-entry (*dop-2*, *sams-5*, *ttx-3*, and *npr-4*) between inhibition (red, +ATR) and no inhibition (blue, -ATR) control. **(G)** Representative fluorescence and phase image (left) of *Pnpr-4::ARCH-mCherry* (F). Scale bar 15um. Fractional lawn occupancy of this line as a function of time (right) with (red, +ATR) and without (blue, -ATR) neural inhibition. **(H)** Representative plots of the distance of individual *Pnpr-4::ARCH-mCherry* worms from the edge of the lawn with (red, $n = 5$) and without (black, $n = 5$) neural inhibition. Inset: representative trajectories ($n = 5$) color coded by time of *Pnpr-4::ARCH-mCherry* worms attempting to enter bacteria lawn with (left) and without (right) neural inhibition.

We next asked whether the accelerated pathogen avoidance observed upon neural silencing resulted from increased lawn exits, increased latency to re-entry, or both. Two Archaerhodopsin lines (*Pdop-2::Arch3* and *Pmpz-1::Arch3*) showed increased exit rates, while four lines (*Pflp-4::Arch3*, *Psams-5::Arch3*, *Pttx-3::Arch3*, and *Pnpr-4::Arch3*,) did not show changes in exit rates upon inhibition (**Fig. 2C, D**). Neural inhibition that increased lawn exit rates dramatically increased the number of tracks outside of the lawn (**Fig. 2E**). We next measured the effects of the early neural silencing on lawn re-entry. We found that four lines (*Pdop-2::Arch3*, *Pnpr-4::Arch3*, *Psams-5::Arch3* and *Pttx-3::Arch3*) showed significant increases in latency to re-entry in response to activation of Archaerhodopsin (**Fig. 2F, G**). For example, the inhibition of the *Pnpr-4::Arch3* line over the first two hours of the experiment dramatically decreased lawn re-entry over the entire time course, resulting in worm trajectories stalling at the lawn edge (**Fig. 2H**). These results indicated that inhibition of neurons during an early phase of learned pathogen avoidance could cause long-term changes in aversion to pathogenic bacteria through modulating distinct behaviors.

A small set of neurons influences the encoding of the memory of pathogen exposure

We next sought to identify specific neurons that influence learned pathogen avoidance. To identify neurons that control specific behaviors (**Fig. 3A**), one usually thinks of perturbing N neurons of the nervous system one at a time. These measurements can be visualized as an $N \times N$ set of

equations, which can be easily solved to give the relative contribution of each neuron to the phenotype (**Fig. 3B** [↗](#)). This approach requires as many measurements as the number of neurons being characterized. Utilizing a compressed sensing based approach, we can instead formulate our optogenetic screen results as an underdetermined set of equations $\mathbf{M}\vec{w} = \vec{p}$. The matrix, $\mathbf{M}$, is an incoherent measurement matrix of size 29 by 87 measurement matrix (**Fig. 3C** [↗](#)), with each row corresponding to each Archaelhodopsin line (experiments were performed on 29 lines) and each column corresponding to neural identity. The matrix element, $\mathbf{M}_{ij}$ is equal to 1 if the i th line drives expression in neuron j and is 0 otherwise. $\vec{w}$ corresponds to neural weights, *i.e.* how much each neuron contributes to the phenotype, and $\vec{p}$ is the phenotype vector. The relative contributions (weights) of 87 neuron types to a phenotype can be determined using an L1 norm from these 29 measurements.

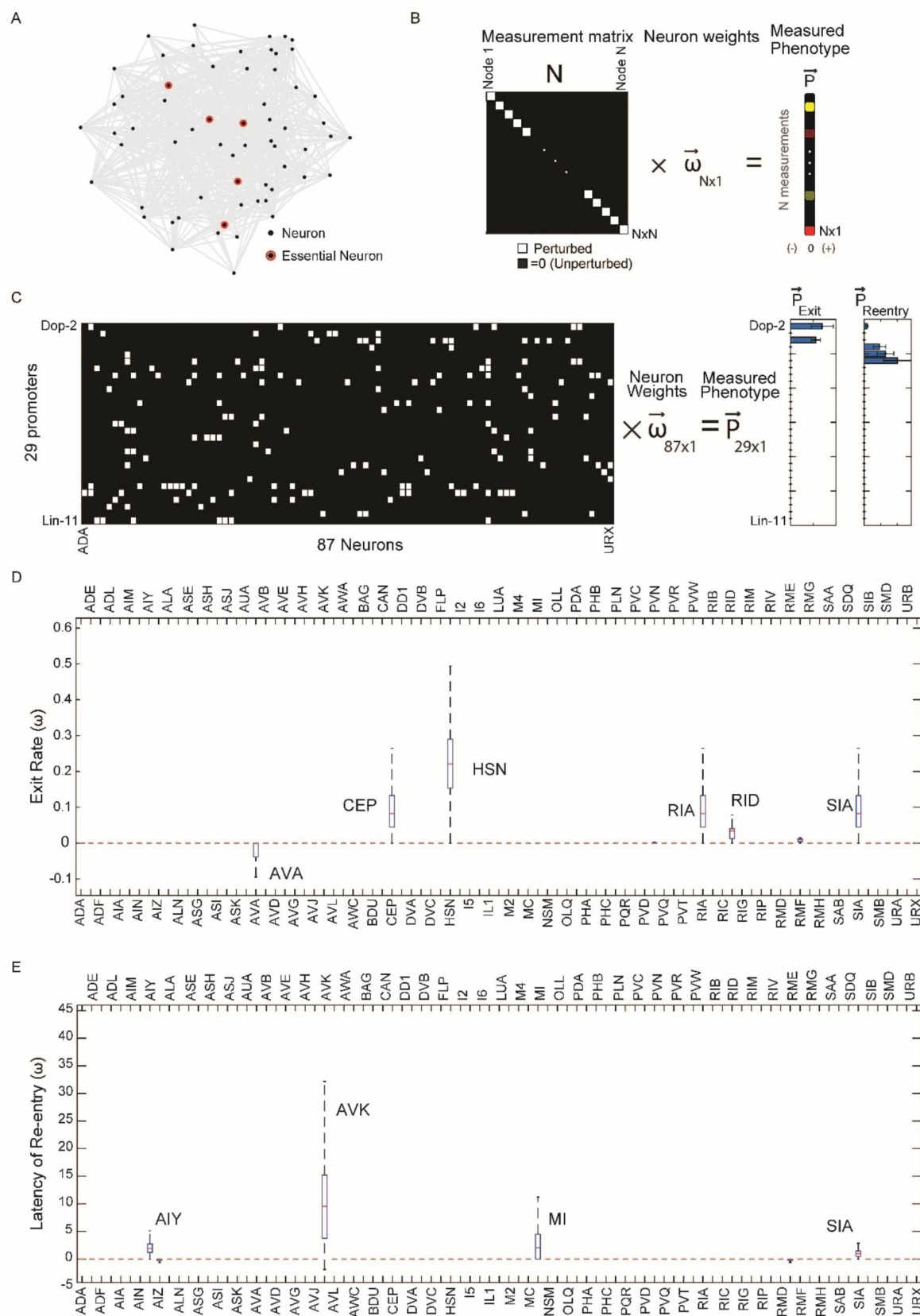


Fig. 3.

Compressed sensing analysis identified a set of neurons controlling worm re-entry and exit.

(A) Example of a neural network where behavior is controlled by a small number of key nodes (highlighted in red) that make up a small fraction of the total number of nodes (gray). (B) Single neuron perturbations can be framed as solving a N by N matrix equation to find the neural contributions to phenotypes. Each diagonal entry (white) corresponds to a single perturbation. Interrogating the entire nervous system to determine the contribution (weight, w) of each neuron to the behavior requires as many measurements as the number of neurons in the nervous system (N). (C) Using compressed sensing to determine neurons controlling entry and exit from the optogenetic screen by determining their weights (w) from an underdetermined set of equations. These equations can be represented as a 29×87 measurement matrix M (left). Rows are promoters and columns are neuron types. Matrix has an entry 1 (white) if the promoter drives expression in that neuron type, else 0 (black). Phenotype vectors P_{exit} and P_{entry} were obtained by taking the difference between the re-entry timescale or rate constant of exit with and without neural inhibition (see methods). From measurements of the phenotype of 29 lines (P_{exit} and P_{entry}), the weights of each of the 87 neurons (w) can be determined using compressed sensing (see methods). (D) Median neuron weight contributions to the exit phenotype from 10,000 lasso regression solutions from bootstrapping (see methods). Neurons with significant weights contributing to the exit phenotype (AVA, CEP, HSN, RIA, RID, SIA) are highlighted. (E) Median neuron weight contributions to the re-entry phenotype from 10,000 lasso regression solutions from bootstrapping (see methods). Neurons with significant weights contributing to the re-entry phenotype (AIY, SIA, AVK, MI) are highlighted.

We evaluated the phenotype vector $\bar{p}$ for the two behaviors of interest by quantifying the changes to the dynamics of re-entry and exit from the lawns of pathogen. For both exit and re-entry, we calculated differences between optogenetically inhibited and non-inhibited worms for each transgenic line that showed statistically significant effects of inhibition. For lawn exit, the phenotype was the increase in the rate constant of exit due to neural inhibition. For re-entry, the phenotype was the difference between the latency in re-entry caused by neural inhibition. Lines that did not show significant behavioral changes in response to inhibition were assigned a phenotype value of zero.

We inferred neural weights $\bar{w}$ from our matrix equation using Lasso regression^{44,45}, which allowed us to solve the underdetermined set of linear equations $M\bar{w} = \bar{p}$ while simultaneously imposing sparsity constraints on the weight vector by minimizing the sum of the mean squared error $\chi^2 = (M\bar{w} - \bar{p})^2$ and the L1 norm of the solutions, thus minimizing $(M\bar{w} - \bar{p})^2 + \lambda \|\bar{w}\|_1$ where λ is the sparsity parameter. Using this approach, we inferred a set of candidate neurons for lawn exit and lawn re-entry (Fig. 3D,E). We focused primarily on neurons governing lawn re-entry (Fig. 1F). For lawn re-entry, 4 key neurons were inferred over a wide range of sparsity parameters: AVK, SIA, AIY and MI. Some neurons were assigned negative weights by this analysis (suggesting that their inhibition promotes lawn re-entry). However, the contributions of these neurons decreased as the sparsity parameter increased, suggesting that these neurons were less important (Fig S2).

Compressed-sensing analysis of lawn-exit behavior identified contributions from six neuron classes to this behavior: CEPs, HSNs, RIAs, RIDs, and SIAs. Some of these neurons have previously been implicated in lawn retention or dwelling. RIAs are required for learned avoidance of *Pseudomonas* lawns^{46,26}. CEPs and HSNs are also known to promote dwelling on bacterial lawns through release of dopamine and serotonin, respectively⁴⁷. Notably, this analysis suggested that the neural circuit governing lawn-exits is distinct from the neural circuit governing re-entry.

We next performed several tests to determine the robustness of our solutions, focusing on lawn re-entry behavior. To determine whether variation in archaerhodopsin expression might affect identification of neurons that govern this behavior, we tested the solutions to corrupted versions of the measurement matrix. The four neurons AVK, SIA, AIY, and MI were robustly identified regardless of matrix corruption (**Fig. S3** [↗](#)). We next tested whether random removal of promoters would alter our solutions, *i.e.*, whether a small subset of strains was driving the identification of neurons. Neuron identification was robust to removal of up to 5 of the 29 promoters (**Fig. S4** [↗](#)). Finally, we determine false-positive and false-negative rates for compressed sensing based inference (**Fig. S5A** [↗](#)), as well as the recovery rate and true positive rate for each of the four neurons (AVK, SIA, AIY and MI) (**Fig. S5B-E** [↗](#)). While all 4 neurons identified have high recovery rates, MI, SIA and AVK can have true recovery rates below 50%. Thus, we would expect at least one of these neurons to be a false positive. To directly test and determine the roles of the neurons implicated by compressed sensing, we next focused on measuring the activities from these neuron types in freely moving animals.

Neurons identified by compressed sensing encode the experience of pathogen exposure as a reduction in neural activity

We measured calcium signals in three neuron subtypes (AVK, SIA, and AIY) in unrestrained worms before and after exposure to pathogenic *Pseudomonas*. We excluded MI from this analysis because of its role as a pharyngeal motor neuron; perturbation of MI might have affected lawn re-entry by modulating bacteria ingestion⁴⁸ [↗](#).

To accurately measure calcium dynamics in AVK, SIA, and AIY we used a custom real-time image-stabilization microscope capable of tracking and measuring neural dynamics in freely moving worms (**Fig. 4A** [↗](#)). The microscope can track worms with 1 μ m precision in all three dimensions while performing rotational stabilization by tracking a marker neuron (AWC^{on}, *Pstr-2::mKO*). We demonstrated the capability of this system by imaging neural activity in freely moving *C. elegans* at high magnification for up to an hour without affecting animal behavior³² [↗](#).

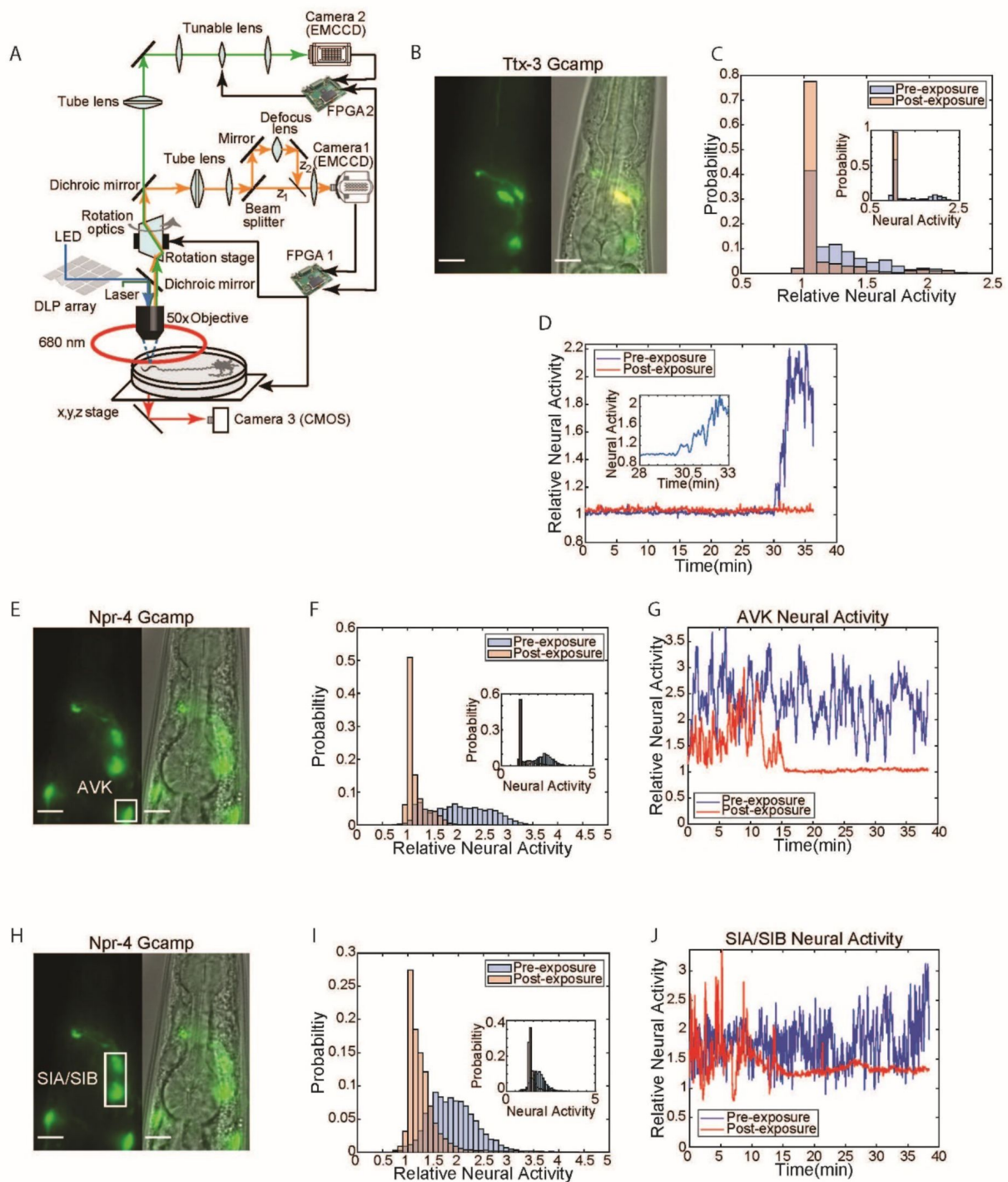


Fig. 4.

Identified neurons responsible for re-entry show reduction in calcium activity following PA14 exposure.

(A) Schematic of tracking and image stabilization microscope allowing for simultaneous measurement of GCaMP activity and worm position with 1µm precision (orange: mKO emission, light green: GCaMP emission, blue: GCaMP excitation, dark green: mKO excitation). An mkOrange labeled marker neuron is imaged through Camera 1. This imaging data is transmitted to FPGA 1 for processing. Processed position information is used to control the x,y,z (stage) and rotational optics to stabilize the worm within the field of view. GCaMP imaging data is acquired via Camera 2. A tunable lens being controlled by FPGA 2 scans through the worm to allow for acquisition of GCaMP images at multiple focal planes. A DLP mirror array, controlled by a PC, is used to target light on specific neurons through structured illumination. (B) Fluorescent and phase image of *Pttx-3::GCaMP* line used to image AIY neural activity. Scale bar 10µm. (C) Histogram of AIY neural activity pre (blue) and post (red) 24 hours of PA14 exposure normalized to pre-exposure baseline (see methods). Data taken from n = 4 worms over 36 min. Inset: histogram of neural activity from a single worm pre (blue) and post (red) 24 hour PA14 exposure. (D) AIY neural activity over 36min for a single worm pre (blue) and post (red) 24 hours of PA14 exposure normalized to pre-exposure baseline (see methods). Inset: zoom in of AIY neural activity between 28 to 33.5minutes to illustrate neural activity transition in naive worms. (E) Fluorescent and phase image of *Pnpr-4::GCaMP* line used for AVK imaging. Scale bar 10µm. (F) Histogram of AVK neural activity pre (blue) and post (red) 24 hours of PA14 exposure normalized to pre-exposure baseline (see methods). Data taken from n = 7 worms over 36 min. Inset: histogram of neural activity from a single worm pre (blue) and post (red) 24 hour PA14 exposure. (G) AVK neural activity for a single worm pre (blue) and post (red) 24 hours of PA14 exposure as a function of time normalized to pre-exposure baseline (see methods). (H) Fluorescent and phase image of *Pnpr-4::GCaMP* used to image SIA neural activity. Scale bar 10µm. (I) Histogram of AVK neural activity pre (blue) and post (red) 24 hours of PA14 exposure normalized to pre-exposure baseline (see methods). Data was taken from n = 8 worms over 36 min. Inset: histogram of neural activity from a single worm (blue) and post (red) 24 hour PA14 exposure. (J) SIA neural activity for a single worm pre (blue) and post (red) 24 hours of PA14 exposure as a function of time normalized to pre-exposure baseline (see methods).

We tracked neurons in transgenic animals expressing the calcium sensor GCaMP6s in AVK, SIA, or AIY interneurons. Naive animals (pre-pathogen exposure) were imaged on an empty agar plate for approximately 40 minutes. These worms were then placed onto a PA14 lawn for 24 hours, recovered to an empty assay plate and imaged for approximately 40 minutes to determine how pathogen exposure affected neural activity. *Pnpr-4::GCaMP6s* was utilized to measure the activity of AVKs and SIAs, and *Pttx-3::GCaMP6s* was used to measure the activity of AIYs. We found that AIY (Fig. 4C, D), AVK (Fig. 4F, G), and SIA (Fig. 4I, J) all showed reduced neural activity after pathogen exposure. These results were evident both in analyses of populations of animals (Fig. 4C, F, I), but were also clearly observed in individual worms (Fig. 4C, F, I inset and Fig. 4D, G, J). Pathogen exposure had different effects on different interneurons. AIY neurons of naive animals showed a significant increase in calcium approximately 30 minutes after transfer to assay plates (Fig. 4D). By contrast, AIY neurons of animals that had been exposed to pathogen remained quiescent. AVK and SIA neurons of naive animals displayed continuous high-frequency calcium signals. Post-exposure, AVKs and SIAs displayed long periods of quiescence. This data indicated that the activity of neurons that regulate a behavior critical for learned pathogen avoidance is strongly affected by exposure to pathogen. These neurons thus appear to encode the history of exposure to pathogenic bacteria in their neural activity state.

Modulation of candidate neurons validates their role in inhibition of pathogen lawn re-entry

We next tested how manipulating the activity of neurons identified by compressed sensing as regulators of lawn re-entry affects how naïve animals interact with bacterial lawns. We inhibited each of the three candidate neuron types using Archaelrhodopsin and measured how acute neuronal inhibitions affected re-entry into a lawn of pathogen. We performed parallel measurements of a control set of worms that had not been treated with the opsin cofactor ATR. To target specific neurons, we used a DLP mirror array to restrict illumination to cells of interest as previously described³².

We found that acute inhibition of AIY in naïve animals increased the latency of re-entry onto pathogenic *Pseudomonas*, mimicking the effect of prior pathogen exposure (Fig. 5B). This effect was not the result of a general defect in locomotion or lawn-entry behavior; entry into lawns of non-pathogenic *E. coli* was not affected by AIY inhibition (Fig. 5B). Inhibition of AVK failed to produce any effect on re-entry behavior (Fig. 5D). Inhibition of *npr-4*-expressing neurons also increased latency to lawn entry on PA14 (Fig. 5E). To validate that this effect was due to SIA, we projected patterned light onto *Pnpr-4::Arch3* animals (Fig. 5F) to selectively inhibit SIA/SIB. Inhibition of SIA using this method deterred entry of worms onto PA14 lawns (Fig. 5G), resulting in increased latency in entry (Fig. 5H). Overall, our results show that two out of the three key neurons identified by compressed sensing were able to elicit a change in lawn entry dynamics through inhibition. Together, our neural imaging and selective inhibition suggested that reduced activity of AIY, SIA, and SIB drive the reduction in lawn occupancy triggered by pathogen exposure.

We next tested whether activation of these neurons would suffice to reverse this behavioral switch. To test this, we optogenetically activated AIY, SIA and SIB using transgenes expressing the light gated ion channel channelrhodopsin-2 driven by the two promoters *Pttx-3::ChR2* and *Pnpr-4::ChR2*. Transgenic lines were fed ATR and placed on lawns of pathogenic bacteria for 24 hours to induce lawn evacuation. Blue light (467 nm, 1mW/mm²) was then used to activate these neurons and the rate of re-entry onto the lawn was quantified. For both *Pttx-3::ChR2* (Fig. 5J) and *Pnpr-4::ChR2* (Fig. 5L), activation of these neurons dramatically increased the re-entry rate of experienced animals onto the PA14 lawn. Animals that re-entered the lawn also rapidly exited the lawn. Thus, the increased re-entry rate did not result in sustained increases in lawn occupancy (Fig. 5K). These results further illustrate that lawn exit and re-entry are controlled by a distinct set of neurons (Fig. 2C, D) and demonstrate that both behavioral modules must change in order to evacuate the bacterial lawn.

Discussion

After experiencing pathogenic bacteria, worms switch their foraging behavior and evacuate the bacterial lawn. We found that this learned pathogen avoidance behavior is driven in part by changes to lawn re-entry behavior. Unlike naïve animals, which rapidly re-enter a lawn of pathogen after they exit, animals previously exposed to pathogen dramatically delay re-entry upon encountering the pathogenic bacteria lawn. Such contact-dependent pathogen aversion depends in a graded manner on the extent of pathogen exposure; the latency to lawn re-entry increases with increased time of pathogen exposure. Contact-dependent inhibition of lawn re-entry is a previously unappreciated behavioral response to pathogen exposure revealed by our study. This behavior is distinct from associative olfactory learning and modulation of chemotactic behaviors reported by other studies^{16,26}. In our study, continuous monitoring of worms exposed to pathogen revealed that animals that leave the pathogen lawn are capable of chemotaxis back to the lawn but stall upon contacting the lawn and do not re-enter. We further

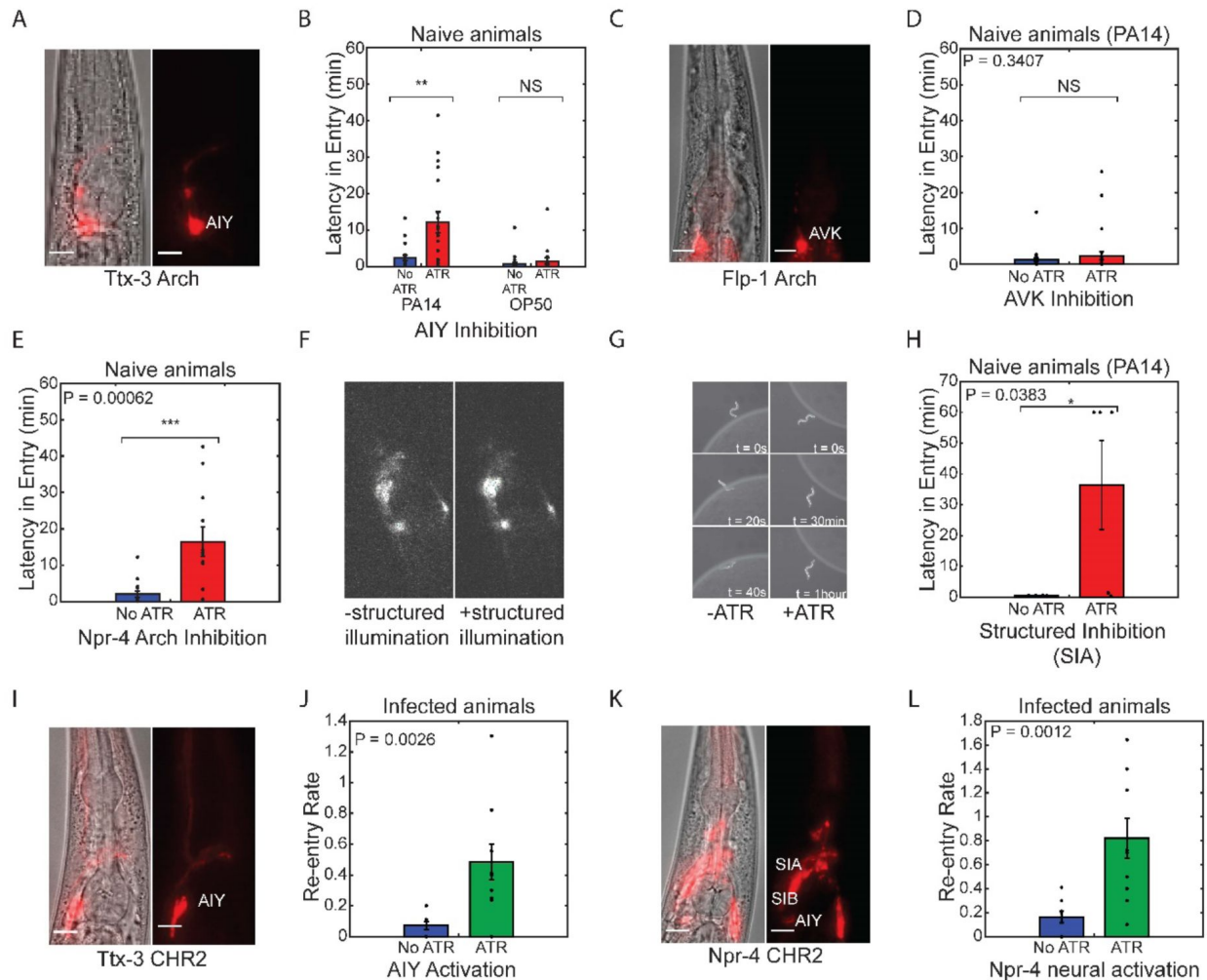


Fig. 5.

Entry onto pathogenic bacteria can be controlled through modulation of key neurons.

(A) Fluorescent and phase image of *Ptx-3::ARCH* line used for neuron specific AIY inhibition. Scale bar 10μm. (B) Latency in entry (delay in the entry of the animal onto the bacteria lawn upon first contact) of naive worms onto PA14 (n > 18) and OP50 (n > 17) lawns with (red) and without (blue) AIY neural inhibition. Neural inhibition resulted in significant increases in latency in entry onto PA14, but not OP50. (C) Fluorescent and phase image of *Pflp-1::NpHR* line used for neuron specific AVK inhibition. Scale bar 10μm. (D) Latency in entry of worms onto PA14 lawns with (red) and without (blue) AVK neural inhibition (n > 27). (E) Latency in entry of naive worms onto PA14 (n > 11) and OP50 (n > 16) with (red) and without (blue) inhibition of *npr-4* expressing neurons. Neural inhibition resulted in significant increases in latency in entry onto both PA14 and OP50. (F) Fluorescent images of *Pnpr-4::ARCH* with (+ATR) and without (-ATR) targeted illumination of SIA/SIB neuron cluster. (G) Representative sequence of phase contrast images of a naive worm attempting to enter PA14 bacteria lawn with (+ATR) and without (-ATR) SIA neural inhibition. Time labels denote time after first contact of worm with bacteria lawn. (H) Latency in entry of naive worms onto PA14 with (red) and without (blue) SIA neural inhibition. Neural inhibition resulted in significant increase in latency in entry for PA14. Mean and SEM over n = 5 samples of 10 worms each. (I) Fluorescent and phase image of *Ptx-3::CHR2* line used for neuron specific AIY activation. Scale bar corresponds to a length of 10μm. (J) Re-entry rate constant of evacuated worms following 24 hours of exposure to PA14 lawns with (green) and without (blue) AIY neural activation. Mean and SEM of the rate constant over n = 5 samples of 10 worms each (see methods). (K) Fluorescent and phase image of *Pnpr-4::CHR2* line used for neural activation. Scale bar corresponds to a length of 10μm. (L) Re-entry rate constant of evacuated worms following 24 hours of exposure to PA14 lawns with (green) and without (blue) *npr-4* neural activation. Mean and SEM of the rate constant over n = 5 samples (see methods).

found that neurons previously identified as being important in aversive olfactory learning, e.g. AIB, RIA, AIZ and RIM²⁶, were not essential for control of lawn re-entry. Our study indicated that the transition in pathogen lawn re-entry behavior occurs through modulation of two key neurons, AIY and SIA, which both decrease their neural activity in response to pathogen exposure.

If contact-dependent lawn aversion represents a modality of pathogen aversion distinct from aversive olfactory learning, what sensory system governs this behavior? While we have yet to establish the signals involved in this process, some hints as to what might be responsible for this aversion can be inferred from looking at synaptic inputs to the neurons that we identified as key for this behavior. SIAs are connected to two sensory neurons that might play a role in driving neural activity changes following pathogen exposure – URX and CEP⁴⁹. URX is a potent regulator of foraging behavior⁵⁰, is one of the few neurons with contact with the pseudocoelomic fluid of *C. elegans*, and has been previously linked to regulation of metabolic signals and innate immune responses^{51,52}. CEPs mediate mechanosensory detection of bacteria and potentially inhibit locomotion^{47,53}. By being downstream of these two neurons, SIA might integrate chemosensory and mechanosensory stimuli, two signals known to be important in modulating lawn evacuation from prior studies^{24,30}. Other neurons that we identified – the AIYs – are part of the olfactory learning circuit and may thus represent a chemotactic component of this contact dependent pathogen aversion^{26,46}. In addition, AIY acts as a central hub neuron that is downstream of multiple sensory neurons and may thus also act as an integrator for multiple sensory modalities^{54,55,56}. Interestingly, investigation of molecular modulators of this aversion behavior by looking at neuropeptides that are highly and uniquely expressed in our neurons of interest reveals a candidate neuropeptide PDF-2 which is highly expressed in AIY⁵⁷. Knocking out PDF-2 increases contact dependent lawn avoidance (**Fig. S7B, C**). PDF-2 has been implicated in gut to neuron signaling through the Rictor/TORC2 pathway⁵⁸, suggesting a potential mechanism through which pathogen infection data could be communicated to AIY to influence PDF-2 signaling to modulate behavior.

One remarkable aspect of the neurons that control lawn aversion is the fact that early perturbation of these neurons (within the first one to three hours of the animal's deposition on the pathogenic lawn) produce long term changes in pathogen avoidance behavior. Effects of this early neural inhibition could be seen in differences in lawn occupancy 15 hours later, suggesting that early suppression of neural activity patterns have long term consequences of behavior. How could SIA and AIY produce such long-term effects? One possible explanation is that reduction in AIY and SIA neural activity might serve as an internal cue. The reduction in activity could drive a bistable circuit including AIY and SIA, causing extended suppression of their activity and a long-term change in their neural activity patterns. This bistability could be accomplished through positive autoregulatory feedback. PDF-2, which appears to be involved in bacteria re-entry, might provide such a mechanism to provide this feedback mechanism. Both PDF-2 and its receptor PDFR-1 are expressed in AIY⁵⁹, providing a potential feedback loop for bistability.

While contact dependent re-entry represents a novel form of pathogen aversion, it is not the only behavioral transition driving lawn evacuation. Using our compressed sensing based approach, we were able to rapidly assay the nervous system to not only discover the key neurons controlling entry, but also exit from the pathogen lawn. We find there is little overlap between the sets of neurons controlling these two subbehaviors. Consistent with this we were able to modulate the subbehavior independently. Together, these results suggest that neural control of lawn evacuation is highly modular, with different sets of neurons governing the individual behavioral transitions needed for lawn evacuation. This high degree of modularity in control over net pathogen aversion is similar to that seen in several forms of aversive olfactory learning of *C. elegans* in response to pathogenic bacteria. For example, imprinting of a memory of pathogenic bacteria in larvae stage worms requires distinct sets of neurons for the establishment of the memory and the expression of that memory⁶⁰, while aversive learning in adult worms carry distinct neural circuits for naïve versus learnt olfactory preference²⁶.

Method details

Strains

All lines used in this work are listed in supplementary tables. Lines used for the measurement matrix are in [Supplementary Table 1](#). Lines used for neural activation and halorhodopsin can be seen in [Supplementary Table 2](#). Finally, lines used for neural imaging can be seen in [Supplementary Table 3](#). PDF-2 mutant used was wSR897: nlp-37(tm4780).

Lawn evacuation assays

PA14 lawn evacuation assay plates were prepared as follows. PA14 was inoculated into LB media and allowed to grow for 15 hours without shaking at 37°C until culture reached an OD of 0.1 to 0.2. 5uL of this culture was then pipetted onto NGM Agar plates and the colonies were allowed to grow for 18 hours at room temperature. Each colony was surrounded by a ring of filter paper to prevent worms from escaping to the edges of the plate. OP50 lawn evacuation plates were prepared in the same manner as PA14 evacuation plates, however, OP50 bacteria was grown for 18 hours at 37°C with shaking. This longer growth period was used to roughly match the thickness of the PA14 and OP50 colonies.

To begin lawn evacuation, 10 worms were transferred onto the bacteria lawn and given one hour to settle. All worms used for this assay were 1 day old hermaphroditic adults grown on OP50 bacteria. The assay was then imaged at 3fps for a total of 18 hours to assay lawn avoidance. Occupancy rate was calculated as (Number of worms on lawn)/(Total number of worms). Worms were counted as being on the lawn if any part of the body was in contact with the lawn.

Lawn re-entry assay

PA14 bacteria colonies were generated as described for the lawn evacuation assay. Assays were initiated by placing 5-10 worms 5mm away from the lawn edge, and then imaging the worms at 3fps for 1 hour. Worms were infected prior to the assay by placing them onto a PA14 lawn evacuation colony for 15 hours. Worms that evacuated the colony over this time period were removed from the plate and used in the assay. Control worms were left on an OP50 lawn during this timeframe.

Optogenetic assay for lawn evacuation

All transgenic lines used for this assay were generated by fusing Archaelhodopsin-3 to the relevant promoters via fusion PCR and then injecting the resulting constructs into worms. Worms used for optogenetic assays were fed on the rhodopsin cofactor all-trans retinal (ATR) for +12hours before the assay. One day old adults were used for all behavioral assays.

Plates were set up as described for the lawn evacuation assays above. Following this, we performed optogenetic inhibition with pulsed (1sec on, 1 sec off) 5mW/mm² green light for 2 hours using a ScopeLED G250 to activate archaelhodopsin. Assays were imaged for a total of 18 hours either at 1 frame per 3minutes for the full assay, or 3fps to assay entry and exit sub-behaviors.

Targeted inhibition of neurons during re-entry

Targeted inhibition of SIA and SIB was carried out as described in previous work. Worms were cultured for 12+hours on ATR. Worms were then placed 5 mm away from the edge of a PA14 lawn generated as described above. SIA and SIB were located at the center of the fluorescent pattern

Pnpr-4::Arch3 line. A circular pattern of light was projected from the DLP projector to selectively target SIA/SIB. Worms were tracked in the frame of view and imaged until they fully entered the PA14 colony or for a total of 1 hour after the worms first contacted the lawn.

Quantification of entry and exit rate for optogenetic screening

Lawn exit rate was calculated by evaluating the number of exit events during the 2 hour timeframe of neural inhibition. Lawn evacuation movies were analyzed in 1 minute intervals and any exit events occurring (as defined by a worm completely leaving contact with the bacteria) were noted. The exit events per hour were calculated from this. Lawn entry timescale was also evaluated by measuring the time from first contact of the worm to the bacteria lawn to time of full entry of the worm onto the lawn.

GCaMP imaging and analysis

Calcium imaging was performed using a custom built microscope as previously described in past work at 15.625 frames per second. Worms were first imaged on an empty NGM plate for 40 minutes. Imaged worms were then transferred onto a colony of PA14 bacteria as used for the lawn evacuation assay and allowed to remain there for 24 hours. Following this, infected worms were imaged again for another 40 minutes.

GCaMP intensity information was extracted using custom software written in MATLAB to extract intensity data given segments encapsulating the neurons of interest. GCaMP imaging data was compiled for all imaged worms. Worm to worm variability in GCaMP expression was normalized by dividing all data by the bottom 5 percentile of fluorescence intensity in healthy worms. GCaMP data was smoothed over a 6 sec window.

Re-entry with neural activation

PA14 lawn evacuation plates were prepared as described above, and 10 ATR treated worms expressing channelrhodopsin-2 under the relevant promoters were seeded onto each colony. Channelrhodopsin was activated using 1mW/mm^2 blue light for 1 hour. Re-entry rate (defined as the rate at which worms fully entered the lawn) and contact rate were evaluated and quantified over this time.

Promoter removal and additions

The robustness of the solutions to removal of promoters was tested as follows. 1 to 5 promoters were removed from the measurement matrix at random. The resulting measurement matrix and phenotype vector were used to infer neuronal weights for the re-entry phenotype. This process was repeated 200 times for each of the 1 to 5 promoters, or 1000 times in total for 1 to 5 promoters. Our results demonstrated robustness of AIY, AVK, SIA and MI to these removals.

Robustness of solutions to corruption of measurement matrix

The robustness of the solutions to corruption of the measurement matrix was tested by randomly altering a small fraction of the measurement matrix and re-inferring the neural contributions. 10% of the non-zero entries in the measurement matrix were altered to a value between 0 and 0.5. 1000 such corrupted measurement matrices were generated and solutions to each corrupted matrix were inferred via Lasso regression. Our results demonstrated that corruption generally did not result in alteration of the inferred neurons, with AIY, AVK, MI and SIA robustly being inferred even with matrix corruption.

Calculation of inverse participation ratio and relative expression

Single cell RNA seq data from S. R. Taylor et al. was used to calculate the average expression of each neuropeptide within each neuron type. The relative expression of each of the neuropeptides in each neuron was calculated by dividing expression in each neuron by the maximum expression over all neurons. Inverse participation ratio (IPR) was calculated as:

$$IPR = \frac{1}{\sum_i \left(\frac{E_i}{\sum_i E_i} \right)^2}$$

Where E_i is the expression in the i th neuron.

Acknowledgements

We would like to thank members of the Ringstad and Ramathan lab for their feedback and advice on the manuscript. This work is supported by 5R01NS117908-03 (SR, NR).

Declaration of Interests

The authors declare no competing interests.

Supplementary Materials

Fig. S1.

Lawn evacuation following neural inhibition.

Evacuation dynamics of all 29 transgenic lines following 2 hours of neural inhibition.

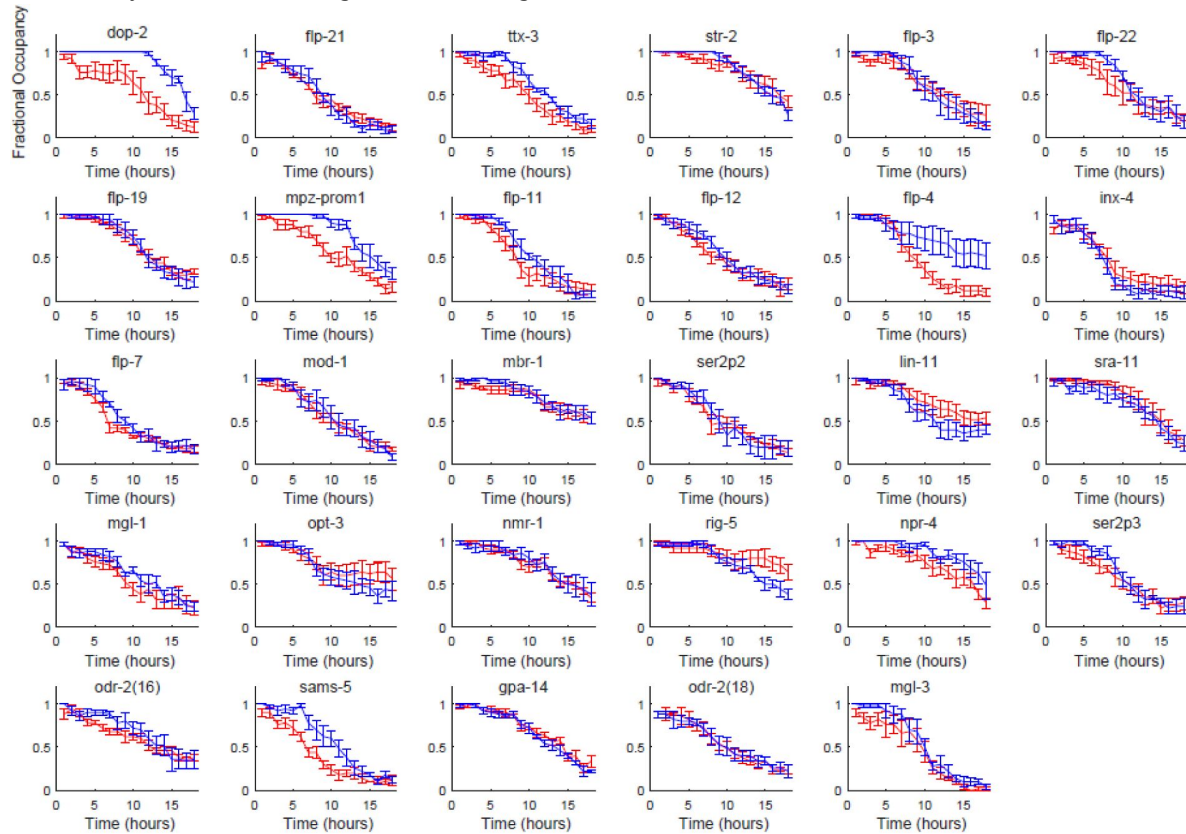
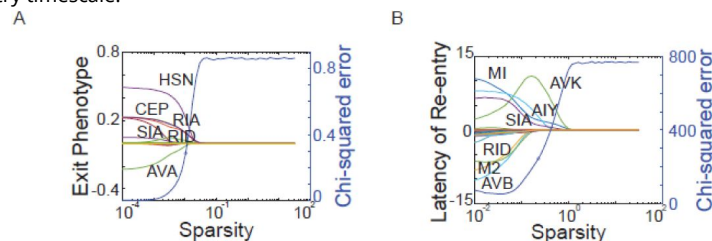


Fig. S2.

Compressed sensing solutions with sparsity parameters for lawn exit and entry

(A) Median neuron weights from 10,000 lasso regression solutions over three orders of magnitude of sparsity parameters for lawn exit rate. (B) Median neuron weights from 10,000 lasso regression solutions over three orders of magnitude of sparsity parameters for lawn entry timescale.



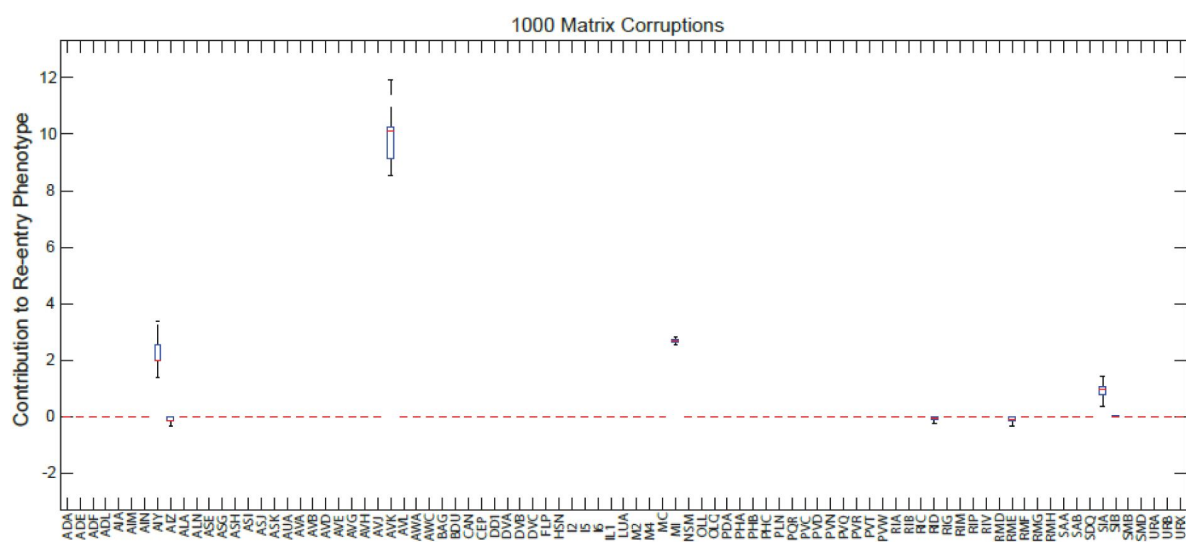


Fig. S3.

Validation of compressed sensing solutions via to Arch expression efficiency through measurement matrix corruption

Solutions obtained following perturbing the measurement matrix to mimic differences in expression of Archaerhodopsin (see methods).

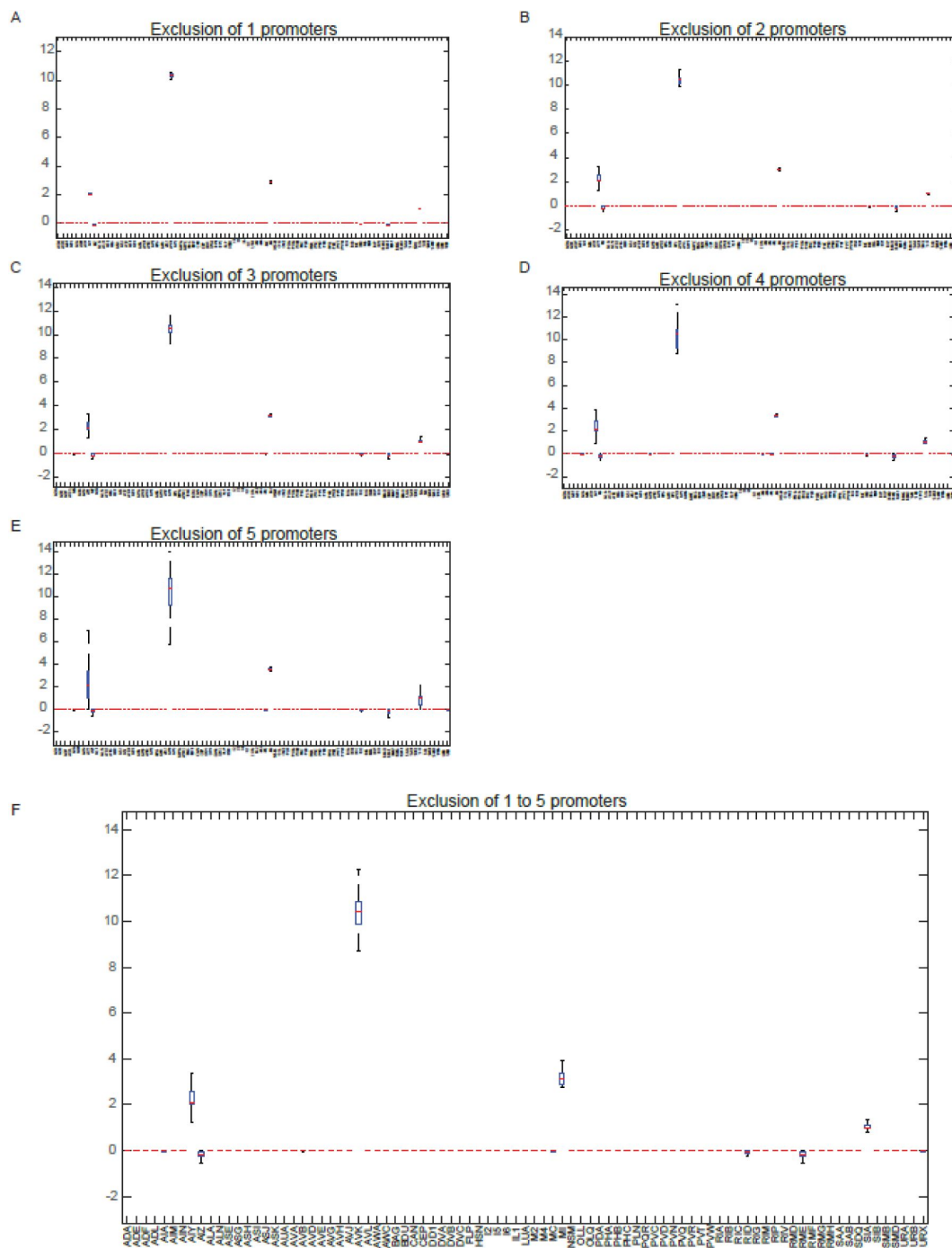


Fig. S4.

Validation of robustness of compressed sensing solutions to choice of measurements through promoter removal

To validate that solutions to lawn entry were robust to choice of measurements, 1 (A) to 5 (E) promoters were removed at random from the measurement matrix (see methods) and solutions were evaluated. Our results showed that our solutions were robust to such removals. (F) Aggregated solutions across 1-5 to removals.

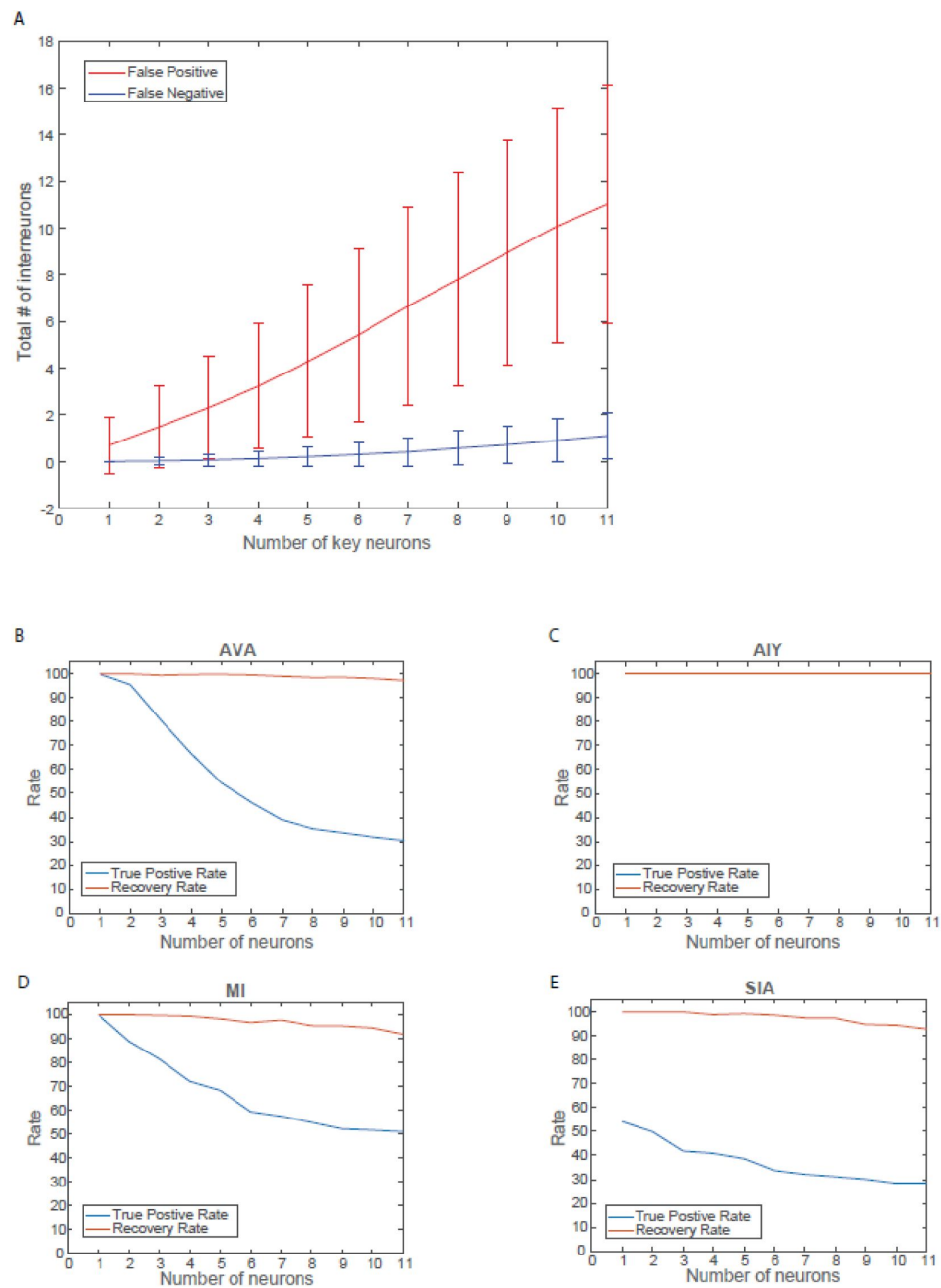


Fig. S5.

Recovery and false positive rate for measurement matrix of choice

(A) False positive and negative rate for recovery of measurement matrix for a randomly simulated set of key neurons (see methods). False positive and recovery rates for each key neuron identified as being important in lawn entry behavior: AVK (B), AIY (C), MI (D) and SIA (E).

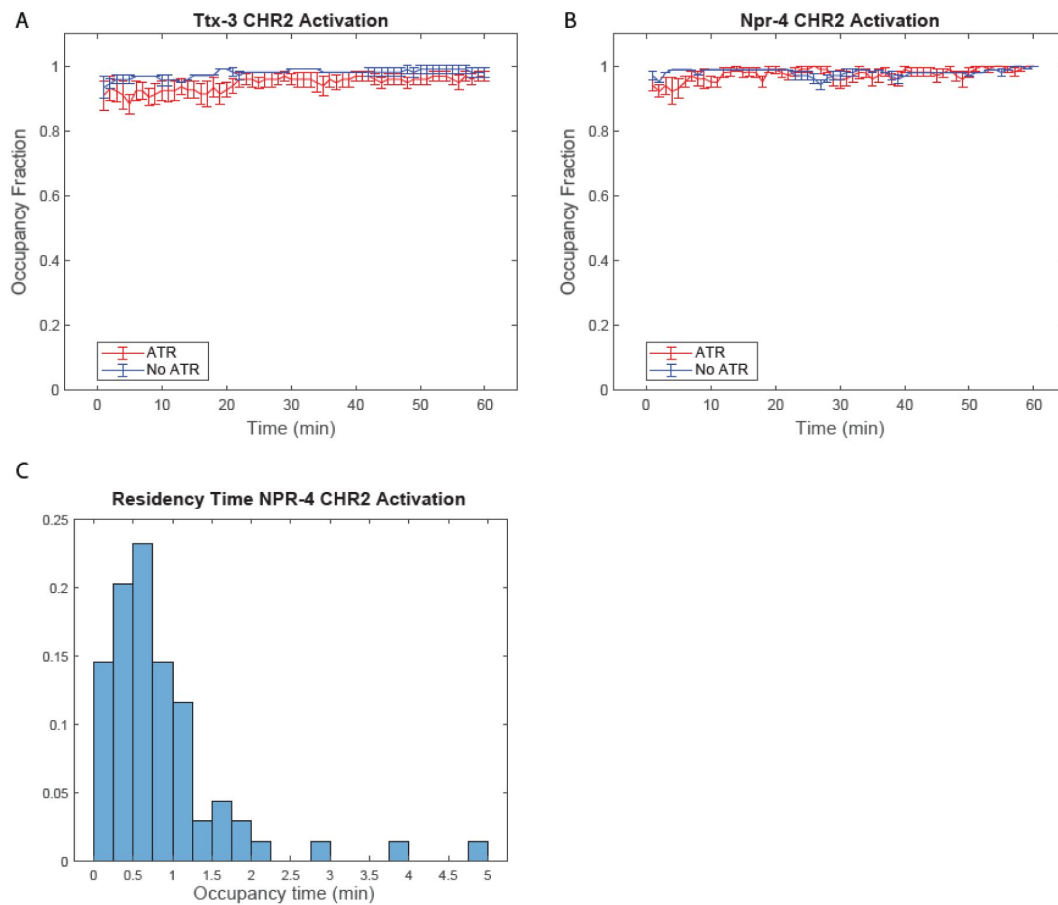


Fig. S6.

Activation of re-entry neurons fails to impact lawn occupancy

Activation of AIY (**A**) and Npr-4 expressing neurons (**B**) fails to significantly increase lawn occupancy in evacuated colonies. (**C**) Worms that re-enter lawn after Npr-4 neural activation show very low residency time on PA14 lawns.

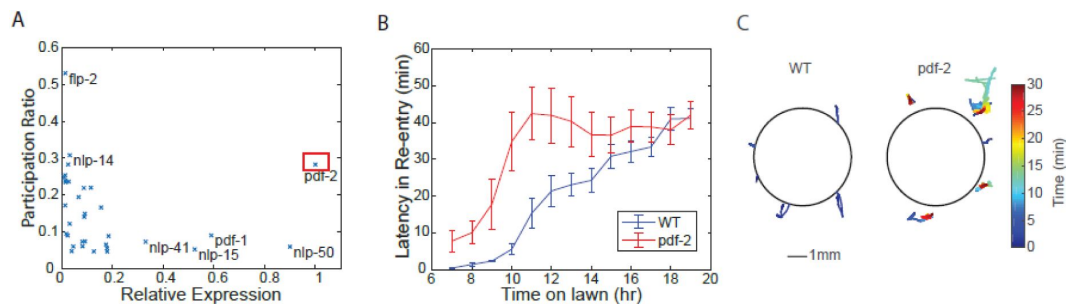


Fig. S7.

Candidate neuropeptide identification from re-entry neurons

(A) Scatterplot of neuropeptides plotted on an axis of maximum relative expression in one of the two candidate neurons (AIY and SIA) versus the specificity of their expression (as measured by calculated participation ratio, see methods). *pdf-2* is highlighted as a neuropeptide of interest as it showed both high expression and high specificity. **(B)** Plot of worm latency of re-entry onto lawns as a function of time spent on PA14 lawn for loss of function mutant of the neuropeptide *pdf-2* (red) and wild type worms (blue). Mutant worms showed systematic increases in latency of re-entry compared to wild type control. **(C)** Representative center of mass trajectories of worms at the edge of the PA14 lawn taken from 12 to 14hours of PA14 exposure for wild type worms (left) and PDF-2 mutant worms (right) color coded by time

Promoters	Neurons
wSR853: [pdop-2::Arch-tagRFP]; srals[pstr-2::mkO]; lite-1(ce314)x	CEP; SIA; SIB; RIA; RID; ADE
wSR517: [pflp-21::Arch-tagRFP, pBX]; pha-1(e2123)III; lite-1(ce314)x	URX; URA; RMG; MC; M2; AIY
sraEx280[pttx-3::Arch-tagRFP; pBX]; pha-1(e2123)III; lite-1(ce314)x	AIY
wSR230: [pstr-2::Arch-tagRFP; pBX]; pha-1(e2123)III; lite-1(ce314)x	AWC
wSR347: [pflp-3::Arch-GFP; pflp-3::mkO; pBX]; pha-1(e2123)III; lite-1(ce314)x	IL1; OLL; URB; PQR
wSR469: [pflp-19::Arch-tagRFP]; pha-1(e2123)III; lite-1(ce314)x	AWA; URX
wSR495: [pmpz-1prom2::Arch-tagRFP]; pha-1(e2123)III; lite-1(ce314)x	M4; NSM; MC; RMH; RMF; RMD; HSN; SDQ; PVC; PVQ; PVN
wSR545: [pflp-11::Arch-tagRFP, pBX]; pha-1(e2123)III; lite-1(ce314)x	AUA; BAG; DVB; LUA; PHC; PVC; SAB; URX
wSR523: [pflp-12::Arch-tagRFP, pBX]; pha-1(e2123)III; lite-1(ce314)x	BAG; SAA; SMB; AVH; AVJ
wSR496: [pflp-4::Arch-tagRFP]; pha-1(e2123)III; lite-1(ce314)x	NSM; ADL; I5; I6; AWC; FLP
wSR512: [pinx-4::Arch-tagRFP, pBX]; pha-1(e2123)III; lite-1(ce314)x	ADA; ADE; AIN; AUA; AVJ; DVC; FLP; PHA; PHB; PVR; PVT; RIC; RIG; RIM; RIP
wSR500: [pflp-7::Arch-tagRFP]; pha-1(e2123)III; lite-1(ce314)x	ALA; AVG; PHB; PDA; PVW; RIC; SAA
wSR683: [pmod-1::Arch-tagRFP, pBX]; srals467[pstr-2::mkO] III; lite-1(ce314)x; pha-1(e2123) III	RID; RME; AIZ; AIY; DD1
wSR448: [pmb-1::Arch-tagRFP]; pha-1(e2123)III; lite-1(ce314)x	AWC; AIM; RIC; AIN
wSR274: [pser-2prom2::Arch-tagRFP; pBX]; pha-1(e2123)III; lite-1(ce314)x	RME; AIZ; RID; AIY; BDU
wSR454: [plin-11::Arch-tagRFP]; pha-1(e2123)III; lite-1(ce314)x	ADF; ADL; AIZ; RIC; AVG; AVH; AVJ
wSR186: [psra-11::Arch-GFP, psra-11::mkO]; lite-1(ce304) x	AIY; AVB; AIA
wSR453: [pmgl-1::Arch-tagRFP]; pha-1(e2123)III; lite-1(ce314)x	AIA; RMD; NSM
wSR345: [popt-3::Arch-GFP; popt-3::mkO; pBX]; pha-1(e2123)III; lite-1(ce314)x	DVA; AVE; ASJ; OLQ; AIM; CAN
wSR507: [pnmr-1::Arch-tagRFP]; pha-1(e2123)III; lite-1(ce314)x	AVA; AVD; AVE; RIM; AVG
wSR241: [prig-5::Arch-tagRFP; pBX]; pha-1(e2123)III; lite-1(ce314)x	RMD; SMD; I2; MC; M4
wSR352: [pnpr-4::Arch-GFP; pnpr-4::mkO; pBX]; pha-1(e2123)III; lite-1(ce314)x	SIA; SIB; RIC; AVA; RMD; AIY; AVK; BAG
wSR550: [pser-2prom3::Arch-tagRFP, pBX]; pha-1(e2123)III; lite-1(ce314)x	OLL; PVD
wSR224: [podr-2(16)::Arch-tagRFP; pBX]; pha-1(e2123)III; lite-1(ce314)x	SMD; RME
wSR486: [psams-5::Arch-tagRFP]; pha-1(e2123)III; lite-1(ce314)x	MI; PVQ

Supplementary Table 1.

Archaeorhodopsin lines that constitute the measurement matrix

Supplementary Table 1. (continued)

wSR468: [pgpa-14::Arch-tagRFP; pbx]; pha-1(e2123)III; lite-1(ce314)x	ASI; ASJ; ASH; ASK; ADE; PHA; PHB; ALA; AVA; CAN; DVA; PVQ; RIA
wSR288: [podr-2(18)::Arch-tagRFP; pBX]; pha-1(e2123)III; lite-1(ce314)x	SMB; RME; ALN; PLN; RIG
wSR535: [pmgl-3:Arch-tagRFP, pBX]; pha-1(e2123)III; lite-1(ce314)x	NSM; ADF; ASE; AWC; RIB; RIC; BAG
wSR499: [pflp-22::Arch-tagRFP]; pha-1(e2123)III; lite-1(ce314)x; line-3	AIM; ASG; AVA; AVG; AVL; CEP; PVD; PVW; RIC; AIZ; RIV; SMD; URA

Supplementary Table 2.**Channelrhodopsin and Halo lines**

Promoters	Neurons
wSR281: [pttx-3::ChR2-tagRFP; pBX]; pha-1(e2123); lite-1(ce314)x	AIY
wSR491:[pnpr-4::ChR2-tagRFP]; pha-1(e2123)III; lite-1(ce314)x	SIA; SIB; RIC; AVA; RMD; AIY; AVK; BAG
ZX888: zxIs16 [pflp-1::NpHR::eCFP; lin-15+]	AVK

Supplementary Table 3.**GCaMP Lines**

Promoters	Neurons Imaged
wSR762: pnpr-4::GCaMP6s; pstr-2::mKO; lite-1(ce314)x	AVK, SIA
wSR490: sraEx490[pttx-3::GCaMP6s(mammalian)]; sraIs467[pstr-2::mKO]; lite-1(ce314)	AIY

References

1. Zhao C., Widmer Y.F., Diegelmann S., Petrovici M.A., Sprecher S.G., Senn W (2021) **Predictive olfactory learning in *Drosophila*** *Sci. Rep* **11** <https://doi.org/10.1038/s41598-021-85841-y>
2. De Belle J.S., Heisenberg M. (1994) **Associative Odor Learning in *Drosophila* Abolished by Chemical Ablation of Mushroom Bodies** *Science* **263**:692–695 <https://doi.org/10.1126/science.8303280>
3. Pascual A., Pr  at T (2001) **Localization of Long-Term Memory Within the *Drosophila* Mushroom Body** *Science* **294**:1115–1117 <https://doi.org/10.1126/science.1064200>
4. Heisenberg M (2003) **Mushroom body memoir: from maps to models** *Nat. Rev. Neurosci* **4**:266–275 <https://doi.org/10.1038/nrn1074>
5. Vargha-Khadem F., Gadian D.G., Watkins K.E., Connelly A., Van Paesschen W., Mishkin M. (1997) **Differential Effects of Early Hippocampal Pathology on Episodic and Semantic Memory** *Science* **277**:376–380 <https://doi.org/10.1126/science.277.5324.376>
6. Bird C.M., Burgess N (2008) **The hippocampus and memory: insights from spatial processing** *Nat. Rev. Neurosci* **9**:182–194 <https://doi.org/10.1038/nrn2335>
7. Kov  cs K.A (2020) **Episodic Memories: How do the Hippocampus and the Entorhinal Ring Attractors Cooperate to Create Them?** *Front. Syst. Neurosci* **14** <https://doi.org/10.3389/fnsys.2020.559186>
8. Duff M.C., Covington N.V., Hilverman C., Cohen N.J (2020) **Semantic Memory and the Hippocampus: Revisiting, Reaffirming, and Extending the Reach of Their Critical Relationship** *Front. Hum. Neurosci* **13** <https://doi.org/10.3389/fnhum.2019.00471>
9. FeldmanHall O., Montez D.F., Phelps E.A., Davachi L., Murty V.P (2021) **Hippocampus Guides Adaptive Learning during Dynamic Social Interactions** *J. Neurosci* **41**:1340–1348 <https://doi.org/10.1523/JNEUROSCI.0873-20.2020>
10. Gourgou E., Adiga K., Goettemoeller A., Chen C., Hsu A.-L (2021) ***Caenorhabditis elegans* learning in a structured maze is a multisensory behavior** *iScience* **24** <https://doi.org/10.1016/j.isci.2021.102284>
11. Amano H., Maruyama I.N (2011) **Aversive olfactory learning and associative long-term memory in *Caenorhabditis elegans*** *Learn. Mem* **18**:654–665 <https://doi.org/10.1101/lm.222441>
12. Eliezer Y., Deshe N., Hoch L., Iwanir S., Pritz C.O., Zaslaver A (2019) **A Memory Circuit for Coping with Impending Adversity** *Curr. Biol* **29**:1573–1583 <https://doi.org/10.1016/j.cub.2019.03.059>
13. Zhang X., Zhang Y (2012) **DBL-1, a TGF-  , is essential for *Caenorhabditis elegans* aversive olfactory learning** *Proc. Natl. Acad. Sci* **109**:17081–17086 <https://doi.org/10.1073/pnas.1205982109>

14. Ardiel E.L., Rankin C.H (2010) **An elegant mind: Learning and memory in *Caenorhabditis elegans*** *Learn. Mem* **17**:191–201 <https://doi.org/10.1101/lm.960510>
15. Meisel J.D., Kim D.H (2014) **Behavioral avoidance of pathogenic bacteria by *Caenorhabditis elegans*** *Trends Immunol* **35**:465–470 <https://doi.org/10.1016/j.it.2014.08.008>
16. Zhang Y., Lu H., Bargmann C.I (2005) **Pathogenic bacteria induce aversive olfactory learning in *Caenorhabditis elegans*** *Nature* **438**:179–184 <https://doi.org/10.1038/nature04216>
17. Chen Z., Hendricks M., Cornils A., Maier W., Alcedo J., Zhang Y (2013) **Two Insulin-like Peptides Antagonistically Regulate Aversive Olfactory Learning in *C. elegans*** *Neuron* **77**:572–585 <https://doi.org/10.1016/j.neuron.2012.11.025>
18. Shivers R.P., Kooistra T., Chu S.W., Pagano D.J., Kim D.H (2009) **Tissue-Specific Activities of an Immune Signaling Module Regulate Physiological Responses to Pathogenic and Nutritional Bacteria in *C. elegans*** *Cell Host Microbe* **6**:321–330 <https://doi.org/10.1016/j.chom.2009.09.001>
19. Singh J., Aballay A (2019) **Microbial Colonization Activates an Immune Fight-and-Flight Response via Neuroendocrine Signaling** *Dev. Cell* **49**:89–99 <https://doi.org/10.1016/j.devcel.2019.02.001>
20. Kaletsky R., Moore R.S., Vrla G.D., Parsons L.R., Gitai Z., Murphy C.T (2020) ***C. elegans* interprets bacterial non-coding RNAs to learn pathogenic avoidance** *Nature* **586**:445–451 <https://doi.org/10.1038/s41586-020-2699-5>
21. Kim D.H. *et al.* (2002) **A Conserved p38 MAP Kinase Pathway in *Caenorhabditis elegans*** *Innate Immunity Science* **297**:623–626 <https://doi.org/10.1126/science.1073759>
22. Tan M.-W., Mahajan-Miklos S., Ausubel F.M (1999) **Killing of *Caenorhabditis elegans* by *Pseudomonas aeruginosa* used to model mammalian bacterial pathogenesis** *Proc. Natl. Acad. Sci* **96**:715–720 <https://doi.org/10.1073/pnas.96.2.715>
23. Reddy K.C., Andersen E.C., Kruglyak L., Kim D.H (2009) **A Polymorphism in *npr-1* Is a Behavioral Determinant of Pathogen Susceptibility in *C. elegans*** *Science* **323**:382–384 <https://doi.org/10.1126/science.1166527>
24. Meisel J.D., Panda O., Mahanti P., Schroeder F.C., Kim D.H (2014) **Chemosensation of Bacterial Secondary Metabolites Modulates Neuroendocrine Signaling and Behavior of *C. elegans*** *Cell* **159**:267–280 <https://doi.org/10.1016/j.cell.2014.09.011>
25. Hao Y., Yang W., Ren J., Hall Q., Zhang Y., Kaplan J.M (2018) **Thioredoxin shapes the *C. elegans* sensory response to *Pseudomonas* produced nitric oxide** *eLife* **7** <https://doi.org/10.7554/eLife.36833>
26. Ha H., Hendricks M., Shen Y., Gabel C.V., Fang-Yen C., Qin Y., Colón-Ramos D., Shen K., Samuel A.D.T., Zhang Y (2010) **Functional Organization of a Neural Network for Aversive Olfactory Learning in *Caenorhabditis elegans*** *Neuron* **68**:1173–1186 <https://doi.org/10.1016/j.neuron.2010.11.025>
27. Harris G., Shen Y., Ha H., Donato A., Wallis S., Zhang X., Zhang Y (2014) **Dissecting the Signaling Mechanisms Underlying Recognition and Preference of Food Odors** *J. Neurosci* **34**:9389–9403 <https://doi.org/10.1523/JNEUROSCI.0012-14.2014>

28. Lee K., Mylonakis E (2017) **An Intestine-Derived Neuropeptide Controls Avoidance Behavior in *Caenorhabditis elegans*** *Cell Rep* **20**:2501–2512 <https://doi.org/10.1016/j.celrep.2017.08.053>
29. Reddy K.C., Hunter R.C., Bhatla N., Newman D.K., Kim D.H (2011) ***Caenorhabditis elegans* NPR-1-mediated behaviors are suppressed in the presence of mucoid bacteria** *Proc. Natl. Acad. Sci* **108**:12887–12892 <https://doi.org/10.1073/pnas.1108265108>
30. Chang H.C., Paek J., Kim D.H (2011) **Natural polymorphisms in *C. elegans* HECW-1 E3 ligase affect pathogen avoidance behaviour** *Nature* **480**:525–529 <https://doi.org/10.1038/nature10643>
31. Bai H., Zou W., Zhou W., Zhang K., Huang X (2021) **Deficiency of Innate Immunity against *Pseudomonas aeruginosa* Enhances Behavioral Avoidance via the HECW-1/NPR-1 Module in *Caenorhabditis elegans*** *Infect. Immun* **89**:e00067–21 <https://doi.org/10.1128/IAI.00067-21>
32. Lee J.B., Yonar A., Hallacy T., Shen C.-H., Milloz J., Srinivasan J., Kocabas A., Ramanathan S (2019) **A compressed sensing framework for efficient dissection of neural circuits** *Nat. Methods* **16**:126–133 <https://doi.org/10.1038/s41592-018-0233-6>
33. Avery L., Horvitz H.R (1989) **Pharyngeal pumping continues after laser killing of the pharyngeal nervous system of *C. elegans*** *Neuron* **3**:473–485 [https://doi.org/10.1016/0896-6273\(89\)90206-7](https://doi.org/10.1016/0896-6273(89)90206-7)
34. Gray J.M., Hill J.J., Bargmann C.I (2005) **A circuit for navigation in *Caenorhabditis elegans*** *Proc. Natl. Acad. Sci. U. S. A* **102**:3184–3191 <https://doi.org/10.1073/pnas.0409009101>
35. Kobayashi J., Shidara H., Morisawa Y., Kawakami M., Tanahashi Y., Hotta K., Oka K (2013) **A method for selective ablation of neurons in *C. elegans* using the phototoxic fluorescent protein, KillerRed** *Neurosci. Lett* **548**:261–264 <https://doi.org/10.1016/j.neulet.2013.05.053>
36. Liu M., Kumar S., Sharma A.K., Leifer A.M (2022) **A high-throughput method to deliver targeted optogenetic stimulation to moving *C. elegans* populations** *PLOS Biol* **20** <https://doi.org/10.1371/journal.pbio.3001524>
37. López-Cruz A., Sordillo A., Pokala N., Liu Q., McGrath P.T., Bargmann C.I (2019) **Parallel Multimodal Circuits Control an Innate Foraging Behavior** *Neuron* **102**:407–419 <https://doi.org/10.1016/j.neuron.2019.01.053>
38. Sordillo A., Bargmann C.I (2021) **Behavioral control by depolarized and hyperpolarized states of an integrating neuron** *eLife* **10** <https://doi.org/10.7554/eLife.67723>
39. Candes E.J., Wakin M.B (2008) **An Introduction To Compressive Sampling** *IEEE Signal Process. Mag* **25**:21–30 <https://doi.org/10.1109/msp.2007.914731>
40. Donoho D.L (2006) **Compressed sensing** *IEEE Trans. Inf. Theory* **52**:1289–1306 <https://doi.org/10.1109/tit.2006.871582>
41. Hobert O., Glenwinkel L., White J (2016) **Revisiting Neuronal Cell Type Classification in *Caenorhabditis elegans*** *Curr. Biol* **26**:R1197–R1203 <https://doi.org/10.1016/j.cub.2016.10.027>
42. White JG, Southgate E, Thomson JN, Brenner S (1986) **The structure of the nervous system of the nematode *Caenorhabditis elegans*** *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **314**:1–340 <https://doi.org/10.1098/rstb.1986.0056>

43. Xu M., Jarrell T.A., Wang Y., Cook S.J., Hall D.H., Emmons S.W (2013) **Computer assisted assembly of connectomes from electron micrographs: application to *Caenorhabditis elegans*** *PLoS One* **8**:e54050–e54050 <https://doi.org/10.1371/journal.pone.0054050>
44. Tibshirani R (2011) **Regression shrinkage and selection via the lasso: a retrospective** *J. R. Stat. Soc. Ser. B Stat. Methodol* **73**:273–282 <https://doi.org/10.1111/j.1467-9868.2011.00771.x>
45. Tibshirani R (1996) **Regression Shrinkage and Selection Via the Lasso** *J. R. Stat. Soc. Ser. B Methodol* **58**:267–288 <https://doi.org/10.1111/j.2517-6161.1996.tb02080.x>
46. Liu H., Wu T., Canales X.G., Wu M., Choi M.-K., Duan F., Calarco J.A., Zhang Y (2022) **Forgetting generates a novel state that is reactivatable** *Sci. Adv* **8** <https://doi.org/10.1126/sciadv.abi9071>
47. Sawin E.R., Ranganathan R., Horvitz H.R (2000) **C. elegans Locomotory Rate Is Modulated by the Environment through a Dopaminergic Pathway and by Experience through a Serotonergic Pathway** *Neuron* **26**:619–631 [https://doi.org/10.1016/S0896-6273\(00\)81199-X](https://doi.org/10.1016/S0896-6273(00)81199-X)
48. Vidal B., Gulez B., Cao W.X., Leyva-Díaz E., Reilly M.B., Tekieli T., Hobert O (2022) **The enteric nervous system of the *C. elegans* pharynx is specified by the *Sine oculis*-like homeobox gene *ceh-34*** *eLife* **11** <https://doi.org/10.7554/eLife.76003>
49. Chen B.L., Hall D.H., Chklovskii D.B (2006) **Wiring optimization can relate neuronal structure and function** *Proc. Natl. Acad. Sci* **103**:4723–4728 <https://doi.org/10.1073/pnas.0506806103>
50. Laurent P., Soltesz Z., Nelson G.M., Chen C., Arellano-Carbajal F., Levy E., de Bono M. (2015) **Decoding a neural circuit controlling global animal state in *C. elegans*** *eLife* **4** <https://doi.org/10.7554/eLife.04241>
51. Styer K.L., Singh V., Macosko E., Steele S.E., Bargmann C.I., Aballay A (2008) **Innate Immunity in *Caenorhabditis elegans* Is Regulated by Neurons Expressing NPR-1/GPCR** *Science* **322**:460–464 <https://doi.org/10.1126/science.1163673>
52. Hussey R., Littlejohn N.K., Witham E., Vanstrum E., Mesgarzadeh J., Ratanpal H., Srinivasan S (2018) **Oxygen-sensing neurons reciprocally regulate peripheral lipid metabolism via neuropeptide signaling in *Caenorhabditis elegans*** *PLOS Genet* **14** <https://doi.org/10.1371/journal.pgen.1007305>
53. Tanimoto Y., Zheng Y.G., Fei X., Fujie Y., Hashimoto K., Kimura K.D (2016) **In actio optophysiological analyses reveal functional diversification of dopaminergic neurons in the nematode *C. elegans*** *Sci. Rep* **6** <https://doi.org/10.1038/srep26297>
54. Chalasani S.H., Chronis N., Tsunozaki M., Gray J.M., Ramot D., Goodman M.B., Bargmann C.I (2007) **Dissecting a circuit for olfactory behaviour in *Caenorhabditis elegans*** *Nature* **450**:63–70 <https://doi.org/10.1038/nature06292>
55. Ashida K., Hotta K., Oka K (2019) **The Input-Output Relationship of AIY Interneurons in *Caenorhabditis elegans* in Noisy Environment** *iScience* **19**:191–203 <https://doi.org/10.1016/j.isci.2019.07.028>
56. Kuhara A., Ohnishi N., Shimowada T., Mori I (2011) **Neural coding in a single sensory neuron controlling opposite seeking behaviours in *Caenorhabditis elegans*** *Nat. Commun* **2** <https://doi.org/10.1038/ncomms1352>

57. Taylor S.R. *et al.* (2021) **Molecular topography of an entire nervous system** *Cell* **184**:4329–4347 <https://doi.org/10.1016/j.cell.2021.06.023>
58. O'Donnell M.P., Chao P.-H., Kammenga J.E., Sengupta P (2018) **Rictor/TORC2 mediates gut-to-brain signaling in the regulation of phenotypic plasticity in *C. elegans*** *PLOS Genet* **14** <https://doi.org/10.1371/journal.pgen.1007213>
59. Flavell S.W., Pokala N., Macosko E.Z., Albrecht D.R., Larsch J., Bargmann C.I (2013) **Serotonin and the Neuropeptide PDF Initiate and Extend Opposing Behavioral States in *C. elegans*** *Cell* **154**:1023–1035 <https://doi.org/10.1016/j.cell.2013.08.001>
60. Jin X., Pokala N., Bargmann C.I (2016) **Distinct Circuits for the Formation and Retrieval of an Imprinted Olfactory Memory** *Cell* **164**:632–643 <https://doi.org/10.1016/j.cell.2016.01.007>

Editors

Reviewing Editor

Ilona Grunwald Kadow

University of Bonn, Bonn, Germany

Senior Editor

K VijayRaghavan

National Centre for Biological Sciences, Tata Institute of Fundamental Research, Bangalore, India

Reviewer #1 (Public Review):

This study identifies two behavioral processes that underlie learned pathogen avoidance behavior in *C. elegans*: exiting and re-entry of pathogenic bacterial lawns. Long-term behavioral tracking indicates that animals increase the prevalence of both behaviors over long-term exposure to the pathogen *Pseudomonas aeruginosa*. Using an optogenetic silencing screen, the authors identify groups of neurons, whose activity regulates lawn occupancy. Surprisingly, they find that optogenetic inhibition of neurons during only the first two hours of pathogen exposure can establish subsequent long-term changes in pathogen aversion. By leveraging a compressed sensing approach, the authors define a set of neurons involved in either lawn exit or lawn re-entry behavior using a constrained set of transgenic lines that drive Arch-3 expression in overlapping groups of neurons. They then measure the calcium activity of the candidate neurons involved in lawn re-entry in freely moving animals using GCaMP, and observe a reduction in their neural activity after exposure to a pathogen. Optogenetic inhibition of AIY and SIA neurons during acute pathogen exposure in naïve animals delays lawn entry whereas activating these neurons in animals previously exposed to pathogen enhances lawn entry, albeit transiently.

This work is missing several controls that are necessary to substantiate their claims. My most important concern is that the optogenetic screen for neurons that alter pathogenic lawn occupancy does not have an accompanying control on non-pathogenic OP50 bacteria. Hence, it remains unclear whether these neuronal inhibition experiments lead to pathogen-specific or generalized lawn-leaving alterations. For strains that show statistical differences between - and + ATR conditions, the authors should perform follow-up validation experiments on non-pathogenic OP50 lawns to ensure that the observed effect is PA14-specific. Similarly, neuronal inhibition experiments in Figures 5E and H are only performed with naïve animals on PA14 - we need to see the latency to re-entry on OP50 as well, to make general conclusions about these neurons' role in pathogen-specific avoidance.

My second major concern is regarding the calcium imaging experiments of candidate neurons involved in lawn re-entry behavior. Although the data shows that AIY, AVK, and SIA/SIB neurons all show reduced activity following pathogen exposure, the authors do not relate these activity changes to changes in behavior. Given the well-established links between these cells and forward locomotion, it is essential to not only report differences in activity but also in the relationship between this activity and locomotory behavior. If animals are paused outside of the pathogen lawn, these neurons may show low activity simply because the animals are not moving forward. Other forward-modulated neurons may also show this pattern of reduced activity if the animals remain paused. Given that the authors have recorded neural activity before and after contact with pathogenic bacteria in freely moving animals, they should also provide an analysis of the relationship between proximity to the lawn and the activity of these neurons.

This work is missing methodological descriptions that are necessary for the correct interpretation of the results shown here. Figure 2 suggests that the determination of statistical significance across the optogenetic inhibition screen will be found in the Methods, but this information is not to be found there. At various points in the text, authors refer to "exit rate", "rate constant", and "entry rate". These metrics seem derived from an averaged measurement across many individual animals in one lawn evacuation assay plate. However "latency to re-entry" is only defined on a per-animal basis in the lawn re-exposure assay. These differences should be clearly stated in the methods section to avoid confusion and to ensure that statistics are computed correctly.

This work also contains mislabeled graphs and incorrect correspondence with the text, which make it difficult to follow the authors' claims. The text suggests that *Pdop-2::Arch3* and *Pmpz-1::Arch3* show increased exit rates, whereas Figure 2 shows that *Pflp-4::Arch3* but not *Pmpz-1::Arch3* has increased exit rate. The authors should also make a greater effort to correctly and clearly label which type of behavioral experiment is used to generate each figure and describe the differences in experimental design in the main text, figure legends, and methods. Figure 2E depicts trajectories of animals leaving a lawn over a 2.5-minute interval but it is unclear when this time window occurs within the 18-hour lawn leaving assay. Likewise, Figure 2H depicts a 30-minute time window which has an unclear relationship to the overall time course of lawn leaving. This figure legend is also mislabeled as "Infected/Healthy", whereas it should be labeled "-/+ ATR".

This work raises the interesting possibility that different sets of neurons control lawn exit and lawn re-entry behaviors following pathogen exposure. However, the authors never directly test this claim. To rigorously show this, the authors would need to show that lawn-exit-promoting neurons (CEPs, HSNs, RIAs, RIDs, SIAs) are dispensable for lawn re-entry behavior and that lawn re-entry promoting neurons (AVK, SIA, AIY, MI) are dispensable for lawn exit behavior in pathogen-exposed animals. The authors identify AVK neurons as important for modulating lawn re-entry behavior by brief inhibition at the start of pathogen exposure but fail to find that these neurons are required for increased latency to re-entry in naïve animals (Figure 5D). Recent work from Marquina-Solis et al (2024) shows that chronic silencing of these neurons delays pathogen lawn leaving, due to impaired release of *flp-1* neuropeptide. Authors may wish to connect their work more closely with the existing literature by investigating the behavioral process by which AVK contributes to lawn evacuation.

If the authors work through these criticisms, this work can become an important contribution to the field of pathogen learning in *C. elegans*. However, in its current form, this work remains incomplete.

<https://doi.org/10.7554/eLife.97340.1.sa2>

Reviewer #2 (Public Review):

In this manuscript, Hallacy et al. used a compressed sensing-based optogenetic screening method to investigate the crucial neurons that regulate pathogenic avoidance behavior in *C. elegans*. They further substantiate their findings using complementary optogenetic activation and imaging techniques to confirm the roles of the key neurons identified through extensive screening efforts. Notably, they identified AIY and SIA as pivotal neurons in the dynamic process of pathogenic avoidance. Their significant discovery is the delayed or stalled reentry process, which drives avoidance behavior; to my knowledge, this dynamic has not been previously documented. Additionally, the successful integration of quantitative optogenetic tools and compressed sensing algorithms is noteworthy, demonstrating the potential for obtaining highly quantitative data from the *C. elegans* nervous system. This approach is quite rare in this field, yet it represents a promising direction for studying this simple nervous system.

However, the paper's main weakness lies in its lack of a detailed mechanism explaining how the delayed reentry process directly influences the actual locomotor output that results in avoidance. The term 'delayed reentry' is used as a dynamic metric for quantifying the screening, yet the causal link between this metric and the mechanistic output remains unclear. Despite this, the study is well-structured, with comprehensive control experiments, and is very well constructed.

<https://doi.org/10.7554/eLife.97340.1.sa1>

Reviewer #3 (Public Review):**Summary:**

Using a compressed sensing-based approach applied previously by the author's group, the authors conducted an initial screen for neurons that when optogenetically down-regulated, influenced learned pathogen avoidance consisting of two component behaviors, exit from the bacterial lawn and lawn re-entry. Authors found that 4 classes of neurons AVK, SIA, AIY, and MI were inferred over a wide range of sparsity parameters, thereby indicating the importance of lawn re-entry. They found six classes of neurons required for lawn exit. The authors then went on to further analyze the neurons for the re-entry behavior, and conducted calcium imaging of those neurons in the freely behaving animals. They found that the activities of AIY and SIA neurons decreased after the animals that had been exposed to the pathogenic bacteria tried to re-enter the bacterial lawn. They also found that when those neurons of the animals that had not been exposed to pathogenic bacteria were downregulated by optogenetics, those operated animals increased the latency of the re-entry, which is a similar behavioral modification to that of the animals that had been exposed to the pathogen. Conversely, those neurons of the animals that were exposed to pathogenic bacteria were up-regulated by optogenetics, those animals showed a shortened latency of the re-entry, which is similar to the behavior observed in the animals not exposed to pathogen.

Strengths:

This is overall a very nice piece of work. Most importantly, an initial screening of neurons was conducted by a compressed sensing-based approach previously applied by the same group. It is also worth emphasizing that this compressed analysis is applicable when the behavior of interest involves a small number of neurons, as the authors pointed out in the Introduction Session. Therefore, the readers should keep in mind that the validation and significance of this work heavily depend on the justification of scarcity parameters that the authors chose. Nevertheless, this work is well justified because neurons identified by the

initial screening were thoroughly analyzed by various methods including calcium imaging and optogenetic manipulation of neuronal activities and behavioral analyses using an animal-tracking system.

Weaknesses:

My only concern is that the authors should be more careful about describing their "compressed sensing-based approach". Authors often cite their previous Nature Methods paper, but should explain more because this method is critical for this manuscript. Also, this analysis is based on the hypothesis that only a small number of neurons are responsible for a given behavior. Authors should explain more about how to determine scarcity parameters, for example.

<https://doi.org/10.7554/eLife.97340.1.sa0>